# **761 Compact IC**





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# **761 Compact IC**

Program «761 PC Software 1.1»



8.761.1063 Instructions for Use



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3	Feedthrough for aspirating tubin	ng3	<b>12</b> P	Pump head	7,9,177
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5	Feedthrough for capillaries	3 <b>4</b>	14	Connection capillary	7,9
6	Connection for drain tube			nlet capillary for detector block	7,9,37,40
7	Feedthrough for capillaries	Δ		Detector block	
8	Connection to purge valve			Suppressor module	
9	Mains pilot lamp			ubing cartridge	
10	Bottle rack			Contact pressure lever	
11	Opening for detector cable	5		· Holding clamp	
12	Opening for inlet capillaries	5		Snap-action lever	
13	Opening for outlet capillaries	5 <b>5</b>		Pump drive	
14	Connection for drain tube	5		· Mounting pin	
15	Knurled screw	5		Compression fitting 17,18,3	
16	Rear panel opening	5		Compression fitting	
17	Detachable rear panel	5		Capillary	
18	Transport security screws	5		Filter-Screw of Filter Unit	
19	Mains switch	5,20		Filter	
20	Mains connection plug	5,20		Filter-Housing of Filter Unit	
21	Fuse holder	5,20		Pulsation dampener	
22	RS232 interface	5		Connection to injection valve .	
23	Connection for detector block	6		Connection to purge valve	
24	Remote interface	5		Aspirating tubing	
25	Serial number	£		ubing nipple	
26	Inlet capillary for injector	_		Threaded stopper	
27	Mounting rail			Bottle attachment	
28	Column connection capillary7,9,32	,34,37,40		Eluent bottle	
29	Sample loop		<b>38</b> A	Aspirating filter	28
30	Connection capillary to syringe	7,9		CO <sub>2</sub> absorber	
31	Rotary nipple for aspirating tube	e7,9 <b>7</b>		Cotton wool	
32	Injection valve7,9	,26,37,40	<b>71</b> S	GJ clip	28
33	PEEK coupling	7,9,40 <b>7</b>	<b>72</b> A	Absorber tube	28
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37	Connection capillary	7,9,26 <b>7</b>	<b>76</b> 2	2 Steel meshes	32
38	Connection capillary	7,9 <b>7</b>	<b>77</b> P	Precolumn cartridge	32
39	Purge valve	7,9,26 <b>7</b>	<b>78</b> S	Steel spacer	32

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# 1 Introduction

## 1.1 Instrument description

The **761 Compact IC** is a PC-controlled system for ion chromatographic analyses. The two following versions are available:

- 2.761.0010 Compact IC without suppressor module
- 2.761.0020 Compact IC with suppressor module

The extremely compact housing of the 761 Compact IC contains everything needed to carry out ion chromatography at the highest quality level:

- **Injection valve** for individual injections or for use with a sample changer such as the Metrohm 766 IC Sample Processor
- **High-pressure pump** extremely low-pulsation double piston pump with a flow range from 0.2 ... 2.5 mL/min and a maximum pressure of 25 MPa (250 bar)
- **Pulsation dampener** even with low-level pressure variations the pulsation dampener reliably protects the column against damage
- Column chamber the perfect insulation of the housing provides not only thermally stable conditions for the separation column but also shields the system against electromagnetic interference
- **Columns** whether anion columns with or without suppression, separation columns for cations or organic acids the 761 Compact IC can accommodate them all
- **Suppressor** the Metrohm Suppressor Module (MSM) is already integrated in the 2.761.0020 Compact IC pressure-resistant, with fully automatic regeneration, highest performance and optimal reproducibility
- **Peristaltic pump** integrated two-channel peristaltic pump with a flow rate of 0.5 ... 0.6 mL/min for regeneration and rinsing of the suppressor module built into the 2.761.0020 Compact IC
- **Detector** conductivity detector with outstanding temperature stability. The detector temperature varies by less than 0.01°C and can be optimally adapted to the ambient conditions.

All components which come into contact with eluent and sample are metal-free.



The **operation** of the 761 Compact IC takes place via a PC connected to the RS232 interface with the help of the control and evaluation program **<761 Compact IC**». This PC program can be used to create systems for recording and evaluating chromatograms. Time programs can also be created in which a large number of instrument functions can be triggered for each program step. It is also possible to use programmable signals to control external instruments such as the 750 Autosampler or the 766 IC Sample Processor via the remote interface.

About 80 prepared system configurations for more than 300 applications are already permanently stored and additional new applications can be downloaded at any time from the Internet under <a href="https://www.metrohm.ch">www.metrohm.ch</a>».

The operating software for the 761 Compact IC meets all the requirements you could place today on a modern integration software: single or multi-point calibration, internal or external standard, selectable algorithms for non-linear calibration, various integration modes with integration parameters and integration events, different methods for peak recognition, peak editor, free scaling, superimposing several chromatograms, use of sample tables and batch reprocessing; a powerful and GLP-conform report generator with output interfaces for monitor, printer and external databases.

The independent «Autodatabase» PC programm supplied can be used to save and handle results and chromatograms produced by the «761 Compact IC» program in a database. With «Autodatabase», data can be sorted, filtered and searched with the help of different criteria. In addition, data and curves can be printed out according to user-defined report templates.

**2** 761 Compact IC

## 1.2 Parts and controls

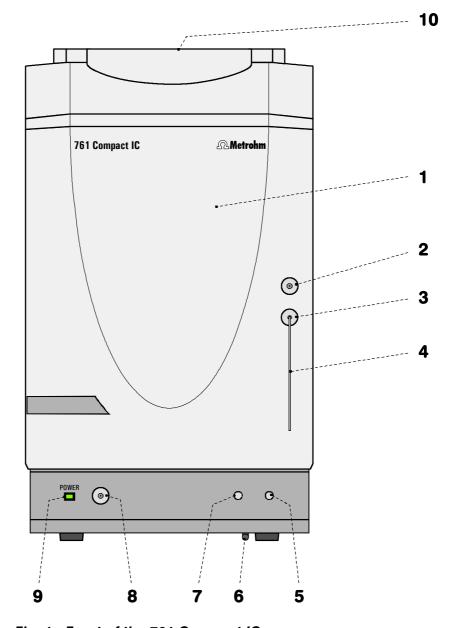


Fig. 1: Front of the 761 Compact IC

1	Door to interior	6	Connection for drain tube for discharge of spilled liquid from the interior
2	Connection for 6.2816.020 Syringe for aspiration of the sample	7	Feedthrough for capillaries
3	Feedthrough for aspirating tubing	8	Connection to purge valve
4	Aspirating tubing for sample	9	Mains pilot lamp lit up when instrument switched on
5	Feedthrough for capillaries	10	Bottle rack for holding supply bottles with eluent, regeneration solution, and rinsing solution

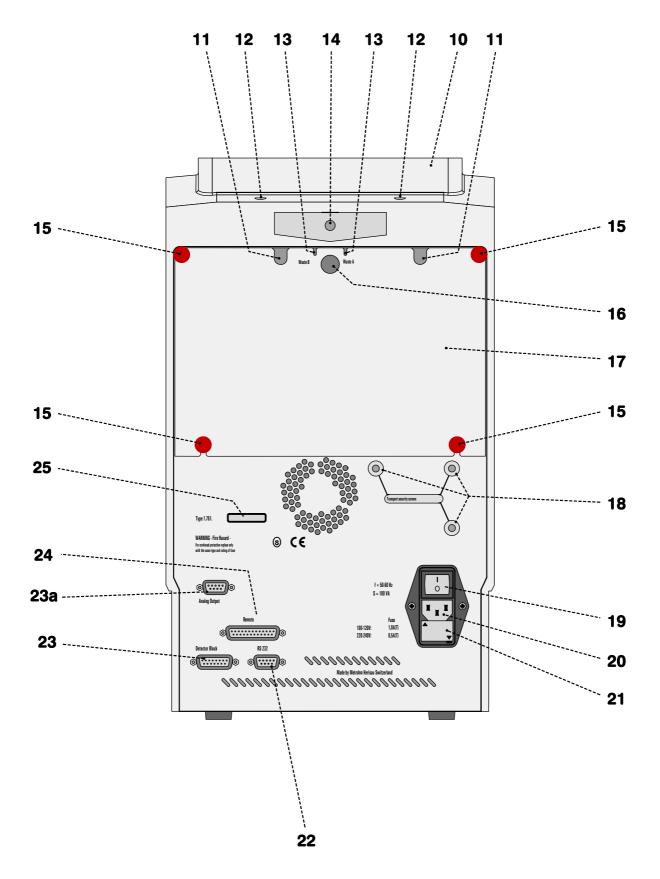


Fig. 2: Rear of the 761 Compact IC

compartment

10	Bottle rack for holding supply bottles with eluent, regeneration solution, and rinsing solution	18	Transport security screws to secure the pump head when the instrument is transported
11	Opening for detector cable	19	Mains switch to switch instrument on and off: $I = ON  0 = OFF$
12	Opening for inlet capillaries for supply of eluent, regeneration solution, and rinsing solution into the inner compartment	20	Mains connection plug mains connection, see section 2.4
13	Opening for outlet capillaries for discharge of eluent, regeneration solution, and rinsing solution from the inner compartment	21	Fuse holder changing the fuses, see section 2.4
14	Connection for drain tube for discharge of spilled liquid from the bottle rack	22	RS232 interface connection of the PC
15	Knurled screw for fastening the rear panel 17	23	Connection for detector block
	J I	23a	Analog output for measuring signal
16	Rear panel opening (closed with plastic stopper) for additional supply and discharge lines to and from the inner compartment	24	Remote interface remote I/O lines for connection of external devices
17	Detachable rear panel access to upper part of inner	25	Serial number

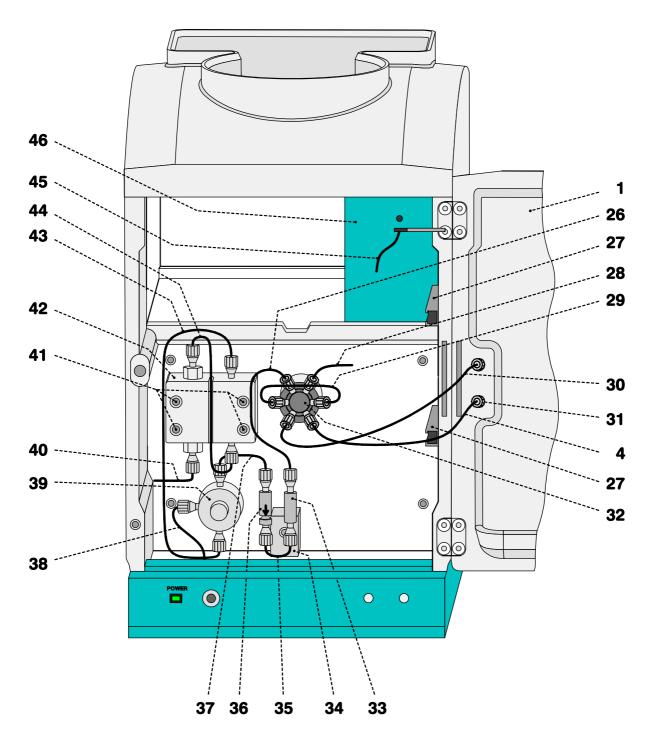


Fig. 3: Interior of the 2.761.0010 Compact IC (with permanently attached accessories and 1.733.0110 Detector block)

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1	Door to interior	36	Filter unit PEEK (6.2821.120)
4	Aspirating tubing for aspirating the sample; 6.1803.020 PTFE tubing, length $L = 52$ cm	37	Connection capillary 6.1831.010 PEEK capillary, length $L=13$ cm
26	Inlet capillary for injector 6.1831.010 PEEK capillary, length $L=24~\mathrm{cm}$	38	Connection capillary 6.1831.010 PEEK capillary, length $L=15$ cm
27	Mounting rail for 6.2027.0X0 column holder	39	Purge valve
28	Column connection capillary 6.1831.010 PEEK capillary, length $L=30~\mathrm{cm}$	40	Aspirating capillary Connection for 6.1834.010 aspirating tubing
29	Sample loop 10 μL 6.1825.230 PEEK sample loop	41	Fastening screws for pump head <b>42</b>
30	Connection capillary to syringe 6.1803.020 PTFE tubing, length $L=30~\mathrm{cm}$	42	Pump head (6.2824.100)
31	Rotary nipple for aspirating tube for fixing the aspirating tube	43	Connection capillary Connection pump head – purge valve, fixed mounting
32	Injection valve	44	Connection capillary in pump head, fixed mounting
33	PEEK coupling (6.2744.040)	45	Inlet capillary for detector block PEEK capillary, fixed mounting
34	Leak detector	46	Detector block (1.732.0110)

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**Connection capillary** 6.1831.010 PEEK capillary, length L = 13 cm

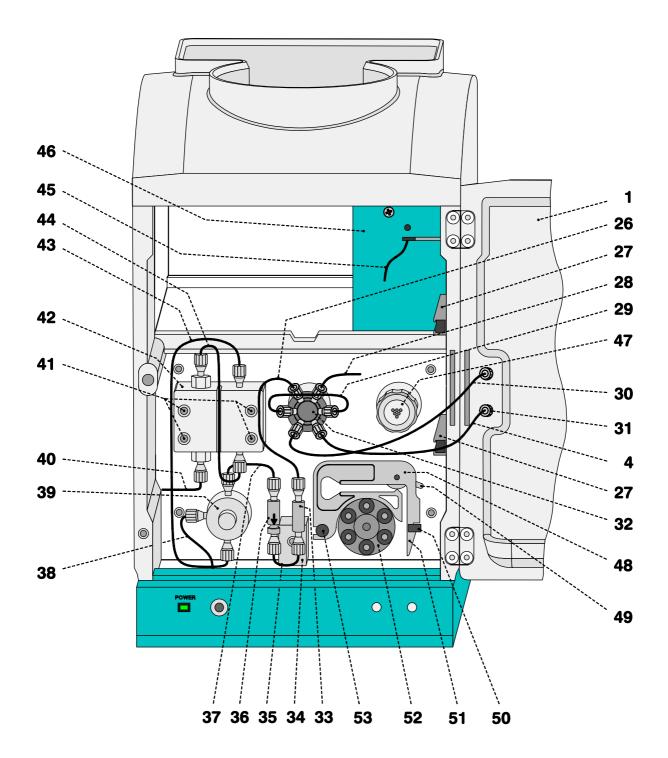


Fig. 4: Interior of the 2.761.0020 Compact IC (with permanently attached accessories and 1.733.0110 Detector block)

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1	Door to interior	39	Purge valve
4	Aspirating tubing for aspirating the sample; 6.1803.020 PTFE tubing, length $L = 52$ cm	40	Aspirating capillary Connection for 6.1834.010 aspirating tubing
26	Inlet capillary for injector 6.1831.010 PEEK capillary, length $L=24~\mathrm{cm}$	41	Fastening screws for pump head <b>42</b>
27	Mounting rail for 6.2027.0X0 column holder	42	Pump head (6.2824.100)
28	Column connection capillary 6.1831.010 PEEK capillary, length $L=30~\mathrm{cm}$	43	Connection capillary Connection pump head – purge valve, fixed mounting
29	Sample loop 20 μL 6.1825.210 PEEK sample loop	44	Connection capillary in pump head, fixed mounting
30	Connection capillary to syringe 6.1803.020 PTFE tubing, length $L=30~\mathrm{cm}$	45	Inlet capillary for detector block PEEK capillary, fixed mounting
31	Rotary nipple for aspirating tube for fixing the aspirating tube	46	Detector block (1.732.0110)
32	Injection valve	47	Suppressor module (inlet and outlet capillaries are not shown)
33	PEEK coupling (6.2744.040)	48	<b>Tubing cartridge (6.2755.000)</b> for 6.1826.060 pump tubing
34	Leak detector	49	Contact pressure lever for adjusting the contact pressure
35	Connection capillary 6.1831.010 PEEK capillary, length $L = 13$ cm	50	Holding clamp for locking the tubing cartridge into place
36	Filter unit PEEK (6.2821.120)	51	Snap-action lever for releasing the tubing cartridge
37	Connection capillary 6.1831.010 PEEK capillary, length $L=13$ cm	52	Pump drive roller head with contact rollers
38	Connection capillary 6.1831.030 PEEK capillary, length $L = 20$ cm	53	Mounting pin for attaching the tubing cartridges



## 1.3 Information on the Instructions for Use



Please read through these Instructions for Use carefully before you put the 761 Compact IC into operation. The Instructions for Use contain information and warnings to which the user must pay attention in order to assure safe operation of the instrument.

## 1.3.1 Organization

These **8.761.1063 Instructions for Use** for the 761 Compact IC provide a comprehensive overview of the installation, startup procedure, operation, fault rectification and technical specifications of this instrument. The Instructions for Use are organized as follows:

Section 1 Introduction

General description of instrument, parts and controls and safety notes

Section 2 Installation

Installation of instrument, accessories,

and external devices

Section 3 Operating tutorial

Introduction to the operation using an example

Section 4 Operation

Detailed description of the operation

Section 5 Notes – Maintenance – Faults

Notes on ion chromatography, maintenance, fault rectification, diagnostic tests, validation

Section 6 Appendix

Technical data, standard equipment, options, warranty, declarations of conformity, index

To find the required information on the instruments, you will find it an advantage to use either the **Table of contents** or the **Index** at the back.

As a supplement to the Instructions for Use, the **Metrohm Monograph 8.732.2003 "Ion chromatography"** is also supplied. This provides an introduction to the theoretical fundamentals and general information on separating columns and sample pretreatment.

The **8.732.2013 IC Applications Collection** is also supplied; this contains all the **Application Notes** on the subject of ion chromatography. Each of these applications can be carried out directly with the 761 Compact IC by loading the system file with the same name. The Applications Collection can be updated at any time by downloading the latest applications from the Internet under «www.metrohm.ch».

You will find detailed information on the separating columns available from Metrohm and on special IC applications in the relevant "Application Bulletins", which are available on request free of charge from your Metrohm agency.

T61 Compact IC

## 1.3.2 Notation and pictograms

The following notations and pictograms (symbols) are used in these Instructions for Use:

Range	Menu item, parameter or entry value
SYSTEM STATE	Program window
<0K>	Button
[ Ctrl ]	Key
35	Part or control of 761
12	Part or control of 750
<u>26</u>	Part or control of 766
	Hazard This symbol draws attention to a possible danger to life or of injury if the associated directions are not followed correctly.
	Warning This symbol draws attention to possible damage to instruments or instrument parts if the associated directions are not followed correctly.
•	Caution This symbol marks important information. First read the associated directions before you continue.
0	Comment This symbol marks additional information and tips.



## 1.4 Safety notes

## 1.4.1 Electrical safety

While electrical safety in the handling of the 761 Compact IC is assured in the context of the specifications IEC 1010-1 (protection class 1, degree of protection IP20), the following points should be noted:

#### Mains connection



Setting of the **mains voltage**, checking the **mains fuse** and the **mains connection** must be effected in accordance with the instructions in section 2.4.

## • Opening the 761 Compact IC



If the 761 Compact IC is connected to the power supply, the instrument must not be opened nor must parts be removed from it, otherwise there is a danger of coming into contact with components which are live. Hence, always disconnect the instrument from all voltage sources before you open it and ensure that the **mains cable is disconnected from mains connection 20**!

## Protection against static charges



Electronic components are sensitive to static charging and can be destroyed by discharges. Before you touch any of the components inside the 761 Compact IC, you should earth yourself and any tools you are using by touching an earthed object (e.g. housing of the instrument or a radiator) to eliminate any static charges which exist.

## 1.4.2 General precautionary rules

### Handling of solvents



Check all lines of the IC system periodically for possible leaks. Follow the relevant instructions regarding the handling of flammable and/or toxic solvents and their disposal.

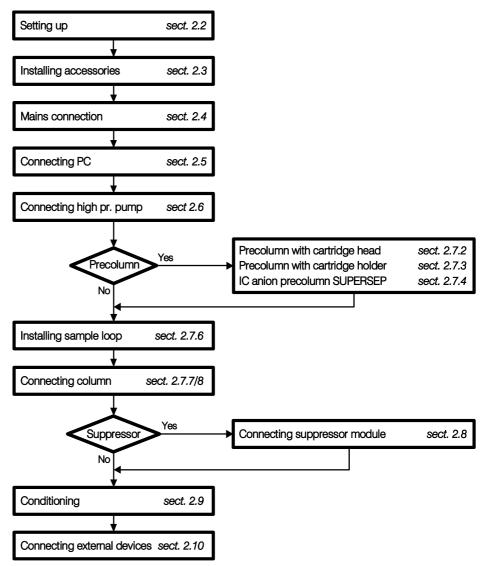
12

# 2 Installation

## 2.1 Overview

## 2.1.1 Flow chart

The following flow chart provides an overview of all installation work. You will find more detailed information in the relevant sections.



## 2.1.2 Connections in the 761 Compact IC

The two following illustrations show the internal connections in the 761 Compact IC in schematic form. The meanings of the various numbered components are given in the detailed illustrations and descriptions in sections 2.2 - 2.10.

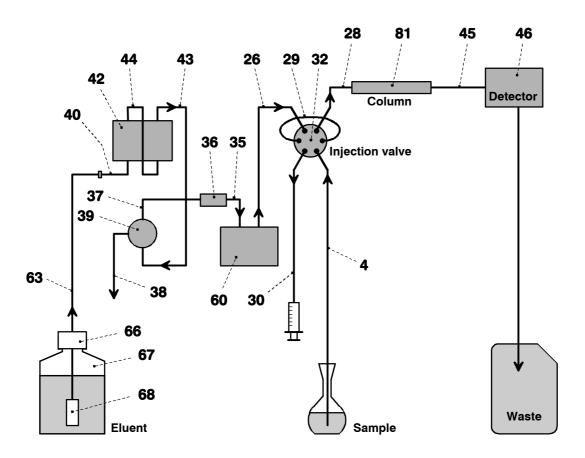


Fig. 5: Connecting diagram for 2.761.0010 Compact IC without suppressor

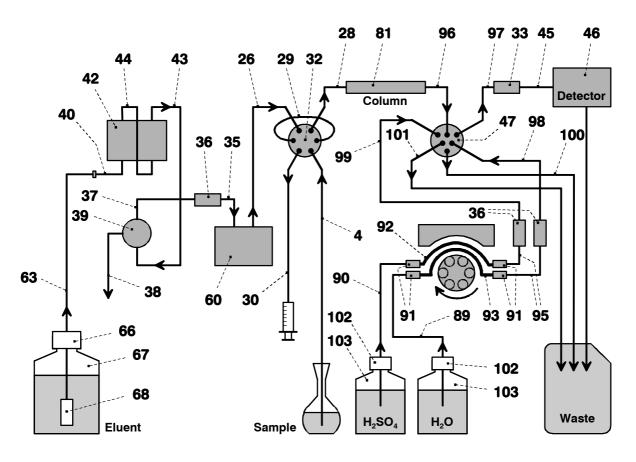


Fig. 6: Connecting diagram for 2.761.0020 Compact IC with suppressor

## 2.2 Setting up the instrument

## 2.2.1 Packaging

The 761 Compact IC is supplied together with the separately packed accessories in special packagings containing shock-absorbing foam linings designed to provide excellent protection. The instrument itself is packed in an evacuated polyethylene bag to prevent the ingress of dust. Please store all these special packagings as only they assure transport of the instrument free from damage.

## 2.2.2 Check

After receipt, immediately check whether the shipment is complete and has arrived without damage (compare with delivery note and list of accessories in section 6.2). In the case of transport damage, see instructions in section 6.4.1 "Warranty".

### 2.2.3 Location

Position the instrument in the laboratory at a location convenient for operation, free from vibrations and protected against a corrosive atmosphere and contamination by chemicals.



To avoid disturbing temperature influences on the insulated column compartment, the instrument must be protected against direct sunlight.

## 2.3 Attaching the accessories

## 2.3.1 Connection of detector block

The metal-free **1.732.0110 Detector block** belongs to the scope of supply of the 761 Compact IC; it must be inserted in the instrument and connected up. Proceed as follows:

#### 1 Note the cell constant

 The cell constant c = XX,X /cm is printed on the rear of the detector block. Note this value; it must subsequently be entered in the software in order to ensure that an exact display of the conductivity is obtained (see section 2.5.3).

#### 2 Install detector block

- Unscrew the four knurled screws **15** from the top rear panel **17** of the 761 Compact IC and remove rear panel (see *Fig. 2*).
- Position detector block 46 from the back in the space provided in the 761 Compact IC and push fully to the front (see Fig. 3 and Fig. 4).

- Insert the cable permanently attached to the detector block
   46 in one of the openings 11 and the outlet capillary in one of the openings 13 of the rear panel 17.
- Replace rear panel 17 and screw to the 761 Compact IC using the four knurled screws 15.

## 3 Connect detector block

 Plug the gray connecting cable permanently attached to the detector block 46 into connection 23 "Detector Block" of the 761 Compact IC and fasten to the instrument by tightening the screws in the cable connector (see Fig. 2).

## 4 Connect waste container

• Lead the outlet capillary of the detector block **46** to a sufficiently large waste container and fix in place.

## 2.3.2 Connection of syringe and aspirating tubing

For manual filling of the sample loop **29** mounted on the injection valve, the 6.2816.020 Syringe and the PTFE aspirating tubing **4** already screwed to the valve are needed. These accessories are mounted or adjusted as follows:

## 1 Connect syringe

• Push 6.2816.020 Syringe (without needle) as far as it will go into connection socket **2** (see *Fig.* 1).

## 2 Adjust aspirating tubing

- Loosen the rotary nipple **31** screwed onto the interior side of feedthrough **3** (see *Fig.* 3 and *Fig.* 4).
- Pull PTFE aspirating tubing **4** (see *Fig.* 3 and *Fig.* 4) by hand out of feedthrough **3** as far as desired.
- Retighten rotary nipple 31 on the interior side of feedthrough
   3 to fix the aspirating tubing 4 in place.

## 2.3.3 Connection of the drain tube for the inner compartment

The 761 Compact IC has a connection at the front to which a drain tube for discharged liquids in the inner compartment can be attached. Proceed as follows:

#### 1 Connect drain tube

• Mount 6.1816.020 Silicone tubing on connection nipple **6** (see *Fig. 1*).

## 2 Lead drain tube to collecting vessel

 Lead the other end of the drain tube to a suitable collecting vessel and fix in place.

T61 Compact IC

## 2.3.4 Connection of the drain tube for bottle rack

The 761 Compact IC has a connection at the rear to which a drain tube for discharged liquids in the bottle rack can be attached. Proceed as follows:

#### 1 Connect drain tube

 Mount 6.1816.020 Silicone tubing on connection nipple 14 (see Fig. 2).

## 2 Lead drain tube to collecting vessel

• Lead the other end of the drain tube to a suitable collecting vessel and fix in place.

## 2.3.5 Connection of PEEK capillaries

For the connections between high-pressure pump and detector block **6.1831.010 PEEK capillaries** (i.d. = 0.25 mm, e.d. =  $^{1}/_{16}$ ") are used which are connected using either **6.2744.010 PEEK compression fittings** (long) or **6.2744.070 PEEK compression fittings** (short). These PEEK connectors can also be used to connect 6.1822.010 PTFE microcapillaries (i.d. = 0.3 mm). Proceed as follows:



Capillaries fitted with new connectors must have a perfectly flat cut surface. To cut PEEK or PTFE capillaries it is best to use the **6.2621.080 Capillary tubing cutter**.

## 1 Mount compression fitting

Slide a compression fitting **54** (6.2744.010) or a compression fitting **55** (6.2744.070) over the end of the capillary **56** to be fastened as shown in *Fig.* 7.

## 2 Insert capillary in connection

Push capillary end in the corresponding connection as far as it will go (to avoid dead volume).

## 3 Tighten compression fitting

Tighten compression fitting **54** or **55** by hand (never use tools).

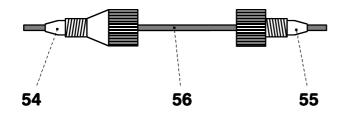


Fig. 7: Connectors for capillaries

- **54** Compression fitting (6.2744.010)
- **55** Compression fitting (6.2744.070)

## 56 Capillary

6.1831.010 PEEK capillary or 6.1822.010 PTFE microcapillary

## 2.3.6 Filter unit PEEK

One **6.2821.120 Filter unit PEEK** (see *Fig. 8*) is already installed between the high-pressure pump and the injection valve at the 761 Compact IC. This filter unit serves to avoid contamination by abrasive particles of the piston seals.

The two other filter units PEEK supplied with the 2.761.0020 Compact IC (with suppressor) are installed between the pump tubings of the peristaltic pump and the inlet capillaries for regeneration and rinsing solution (see *section 2.8.2*). These filter units serve to protect the suppressor module from foreign particles and bacterial growth.

The Filter unit PEEK **36** consists of the filter-housing **59**, the filter-screw **57** and the 6.2821.130 filter **58**. For the connection of capillaries **56** PEEK compression fittings **54** (6.2744.010 or 6.2744.070) must be used. New filters **58** are available as an option with the ordering number 6.2821.130 (set of 10).



For the connection of the filter unit, please note the flow direction arrow printed on the housing.

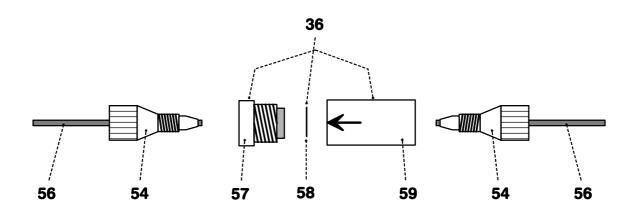


Fig. 8: 6.2821.120 Filter unit PEEK

36	Filter unit PEEK (6.2821.120)	57	Filter-Screw of Filter Unit Part of 6.2821.120 Filter unit	
54	Compression fitting (6.2744.010)	58	<b>Filter 6.2821.130</b> Part of 6.2821.120 Filter unit	
56	Capillary 6.1831.010 PEEK capillary or 6.1822.010 PTFE microcapillary	59	Filter-Housing of Filter Unit Part of 6.2821.120 Filter unit	

18

## 2.4 Mains connection



Follow the instructions below for connecting to the power supply. If the instrument is operated with a mains voltage set wrongly and/or wrong mains fuse, there is a danger of fire!

## 2.4.1 Setting the mains voltage

Before switching on the 761 Compact IC for the first time, check that the mains voltage set on the instrument (see *Fig. 9*) matches the local mains voltage. If this is <u>not</u> the case, you must reset the mains voltage on the instrument as follows:

#### 1 Disconnect mains cable

Disconnect mains cable from mains connection plug **20** of the 761 Compact IC.

## 2 Remove fuse holder

Using a screwdriver, loosen fuse holder **21** below the mains connection plug **20** and take out completely.

## 3 Check and change fuse if necessary

Carefully take the fuse installed for the desired mains voltage out of fuse holder **21** and check its specifications (the position of the fuse in the fuse holder is marked by the white arrow imprinted next to the mains voltage range):

**100...120 V 1.0 A (slow-blow)** Metrohm No. U.600.0016 **220...240 V 0.5 A (slow-blow)** Metrohm No. U.600.0013

## 4 Insert fuse

Change fuse if necessary and reinsert in fuse holder 21.

#### 5 Install fuse holder

Depending on the desired mains voltage, insert fuse holder **21** in the 761 Compact IC so that the corresponding mains voltage range can be read normally and the adjacent white arrow points to the white bar imprinted below the fuse holder (see *Fig. 9*).

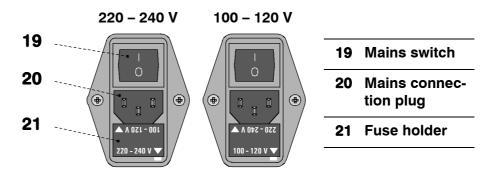


Fig. 9: Setting the mains voltage

## **2.4.2** Fuses

One of the two fuses 1 A/slow-blow for 100...120 V or 0.5 A/slow-blow for 220...240 V is installed in fuse holder **21** of the 761 Compact IC as standard.



Ensure that the instrument is never put into operation with fuses of another type, otherwise there is danger of fire!

For checking or changing fuses, process as described in section 2.4.1.

### 2.4.3 Mains cable and mains connection

### **Mains cable**

The instrument is supplied with one of three mains cables

- 6.2122.020 with plug SEV 12 (Switzerland, ...)
- 6.2122.040 with plug CEE(7), VII (Germany, ...)
- 6.2133.070 with plug NEMA 5-15 (USA, ...)

which are three-cored and fitted with a plug with an earthing pin. If a different plug has to be fitted, the yellow/green lead (IEC standard) must be connected to protective earth (protection class 1).



Any break in the earthing inside or outside the instrument can make it a hazard!

#### **Mains connection**

Plug the mains cable into mains connection plug **20** of the 761 Compact IC (see *Fig.* 9).

## 2.4.4 On/off switching of the instrument

The 761 Compact IC is switched on and off using mains switch **19**. When the instrument is switched on, the mains pilot lamp **9** lights up.

**20** 761 Compact IC

## 2.5 Connection to the PC

## 2.5.1 Connecting cable



Always switch off 761 Compact IC and PC before you connect the two instruments with the 6.2134.100 Cable.

Connect the RS232 interface **22** at the 761 Compact IC to one of the serial COM ports at the PC using the 6.2134.100 Cable (9 pin/9 pin). If only a 25-pin COM interface is available on the PC then the 6.2125.110 Adapter cable or a commercially available adapter must be used.

## 2.5.2 Software installation



These Instructions for Use describe the operation of a single 761 Compact IC connected to a PC. If several instruments should be operated simultaneously with one PC, the «IC Net 2.0» PC program must be installed for this purpose (details see Instructions for Use for «IC Net»).

The PC program **«761 Compact IC 1.1»** is required for the operation of the 761 Compact IC; this is contained on the 6.6030.013 CD included in the accessories. This program runs under Windows 95, Windows 98 and Windows NT operating systems and is installed as follows:

### 1 Install program

- Insert 6.6030.013 Installation CD into CD drive.
- Select <Start> and Run. Browse for the setup.exe file on the installation CD and click on <OK>. Follow the instructions given in the setup program.

The software package will be installed in the desired directory. Icons are created in the program folder and in the startup folder. In addition to the program files, the following folders are installed:

Data	Folder for storage of chromatogram files (*.chw) and batch reprocessing files (*.bar)
Devices	Folder for storage of device files (*.dev)
Methods	Folder for storage of method files (*.mtw)
Reports	Folder for storage of report files (*.txt) and graphic files (*.wmf)
Systems	Folder with subfolders with system files (*.smt) and sample queue files (*.que).

## 2 Registration

 Please send us your 8.761.8007 Registration card as soon as possible. Only registered users will get updated program versions at a special price.





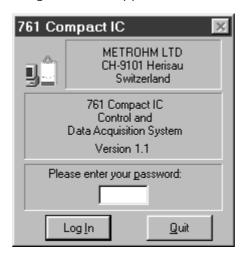
The installed files (incl. system and method files) are generally <u>not</u> write-protected. To prevent these files from being deleted by mistake, switch on the write-protection or make a backup copy in another directory.

## 2.5.3 Basic settings

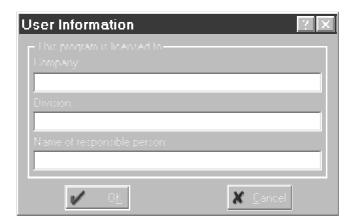
When the program is started for the first time several basic settings must be made for the 761 Compact IC. Proceed as follows:

## 1 Start program

 Double-click the software icon to start the program. The program window with the opening picture is opened and the Log In window appears on the screen:



 Do <u>not</u> enter any password here, just click on <Log <u>I</u>n>. The following window appears:



• Enter company, division and name and click on **<OK>**. This window appears only one time after software installation.

## 2 COM port settings

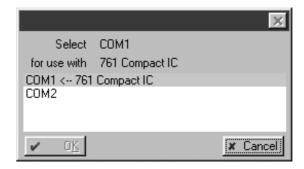
This step must only be carried out if a different **COM interface from COM1** is used for connection to the 761 Compact IC.



 Click on Options / 761 Compact IC:COM1 to open the Links window:



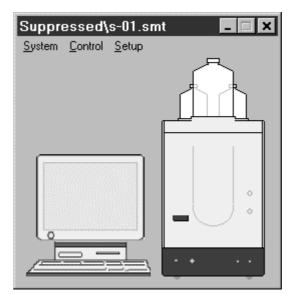
 Click on COM1 using the right mouse button and select the Change menu item to open the following window listing all available COM ports at the PC:



- Select the desired COM port to which the 761 Compact IC has been connected and click on <OK>. The window is closed.
- Close the **Links** window by clicking on **<OK>**.

## 3 Open a system

Click on File / Open / System in the main window. Open either the subfolder Non-suppressed (for 2.761.0010 Compact IC without suppressor) or the subfolder Suppressed (for 2.761.0020 Compact IC with suppressor). Select the system file n-01.smt or s-01.smt and click on <Open>. The corresponding system window is opened:





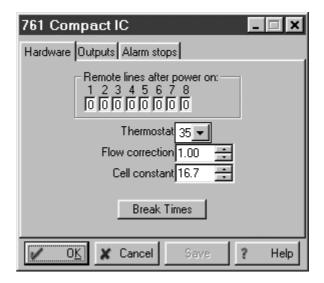
761 Compact IC

- Select the Connect to workplace menu item of the Control menu in this window.
- If the connection between PC and Compact IC is working the message Hardware settings file for 761 unit with serial number '####' not found! Create? will appear. Click <Yes> in order to create the configuration file ####.761 for this instrument.
- If the connection between PC and Compact IC does not work then the message Detection of hardware failed[761 Compact IC [COM#]] appears in the SYSTEM STATE window. In this case check whether the instrument has been switched on, whether the connection cable is connected up properly and whether the COM interface has been set correctly (see point 2). Then repeat point 3.

## 4 Hardware settings

Of the general hardware settings only the input of the cell constant is described. Standard settings can normally be used for all other parameters.

 Click the 761 icon using the right mouse button and select the Hardware item. The Hardware settings window is opened:



- In the **Cell constant** field enter the cell constant which is printed on the 1.732.0110 Detector block (see section 2.3.1).
- Click on **<OK>** to close the window and save the settings.

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## 2.6 High-pressure pump



In order to avoid damage to the pump it must never be operated dry. Each time that the pump is switched on always first check that the eluent supply has been connected up correctly and that sufficient eluent is present in the eluent bottle.

## 2.6.1 Removing the transport security screws

In order to prevent the pump drive from being damaged during transport the pump head is fitted with three transport security screws **18** (see *Fig. 2*). These transport security screws must be removed before the high-pressure pump is started up. Also remove the red sticker attached to the pump head.



In order to avoid damage to the pump head these three security screws should be attached to the pump head each time that it is to be transported.

## 2.6.2 Installing the pulsation dampener

To protect the column material against pressure drops caused by the injector, the **6.2620.150 Pulsation dampener MF** has to be installed between the high-pressure pump and the injection valve of the 761 Compact IC. Proceed as follows (see *Fig. 10*):

## 1 Install pulsation dampener

 Position the pulsation dampener 60 in the interior of the 761 Compact IC on the base.

## 2 Connection to the pump

• Unscrew PEEK capillary **35** of coupling **33** and attach it to connection **62** of the pulsation dampener **60**.

## **3** Connection to injection valve

• Unscrew PEEK capillary **26** of coupling **33** and attach it to connection **61** of the pulsation dampener **60**.



The pulsation dampener is filled with isopropanol and must be rinsed with eluent before connection to a separating column (see section 2.6.4).



The 6.2620.150 Pulsation dampener can be operated in both directions.

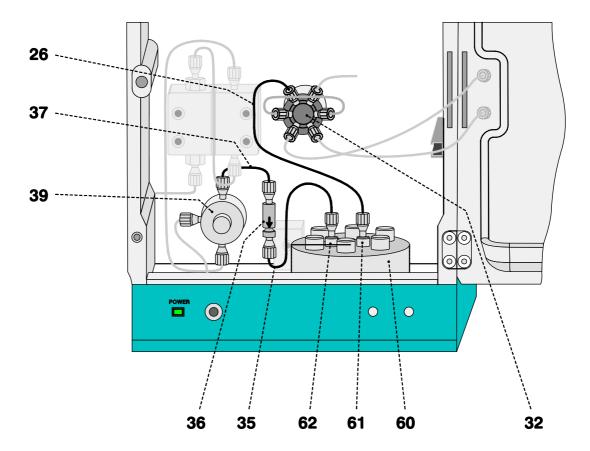


Fig. 10: Connection of the pulsation dampener

26	Inlet capillary for injector 6.1831.010 PEEK capillary, length $L=24$ cm	39	Purge valve
32	Injection valve	60	Pulsation dampener (6.2620.150)
35	Connection capillary 6.1831.010 PEEK capillary, length $L = 13$ cm	61	Connection to injection valve
36	Filter unit PEEK (6.2821.120)	62	Connection to purge valve

## 37 Connection capillary

6.1831.010 PEEK capillary, length L = 13 cm

26

#### 2.6.3 Connecting the eluent bottle

The eluent supply line from the storage bottle to the high-pressure pump is connected as follows (see *Fig. 11*):



Only degassed (with  $N_2$ , He or vacuum) and microfiltered (0.45  $\mu$ m filter) eluents should be used!

The 6.1608.070 Eluent bottle (2 L) supplied is **not suitable for vacuum degassing**. Use a pressure-resistant container for this.

Care must be taken that the **eluent** used is **freely miscible** with any solvent remaining in the pump head (the pump head leaves the factory filled with either isopropanol or methanol/water). If this is not the case then the pump must first be rinsed with a solvent which is miscible with both the previous eluent and the following eluent (e.g. acetone).

#### 1 Prepare eluent bottle

- Prepare, microfilter (0.45 μm microfilter) and degas (with N<sub>2</sub>, He, or vacuum) the suitable eluent for the required application and separating column.
- Fill eluent into eluent vessel 67 (clear glass, 2 L).
- Place eluent bottle **67** at the front in bottle holder **10** on the 761 Compact IC (see *Fig. 1*).

#### 2 Install bottle attachment

- Firmly screw threaded stopper **65** (6.1446.040; part of 6.1602.160) into the smaller threaded opening (M6) of bottle attachment **66** (6.1602.105; part of 6.1602.160).
- Firmly screw aspirating filter **68** onto aspirating tubing **63**.
- Pull the other end of aspirating tubing **63** through the larger threaded opening (M8) of bottle attachment **66** from below.
- Push O-ring (E.301.0021; part of 6.1602.160) over the free end of aspirating tubing **63** and move it towards bottle attachment **66**.
- Push tubing nipple **64** (4.420.4300; part of 6.1602.160) over the free end of aspirating tubing **63**, move it as far as required towards bottle attachment **66** and screw it loosely in the larger opening of bottle attachment **66**.
- Insert aspirating tubing 63 with screwed-on aspiration filter 68 into eluent bottle 67 and screw bottle attachment 66 onto eluent bottle 67.
- Pull aspirating tubing **63** so far through the opening of tubing nipple **64** that aspirating filter **68** is touching the bottom of eluent bottle **67**.
- Fix aspirating tubing **63** in place by screwing shut tubing nipple **64**.



#### 3 Mount CO<sub>2</sub> absorber tube

- First place a piece of cotton wool 70 followed by CO<sub>2</sub> absorber 69 (e.g. Merck soda-lime pellets with indicator, no. 6839.1000) in the large opening of absorber tube 72 and then close it with the plastic lid.
- Fasten absorber tube 72 to bottle attachment 66 with the aid of SGJ clip 71.

#### 4 Connect aspirating tubing to pump

- Insert the free end of aspirating tubing **63** into one of the openings **12** in the interior of the 761 Compact IC (see *Fig. 2*).
- Pull aspirating tubing 63 sufficiently far into the interior of the 761 Compact IC, cut off to the required length and push at least 5 mm of it onto aspirating capillary 40 (see Fig. 3 and Fig. 4) of the high-pressure pump (it may be necessary to use emery paper).
- If necessary, fix aspirating tubing 63 in the required position in the interior with the aid of the Y.107.0150 self-adhesive strap.



Instead of threaded stopper **65**, the 4.420.0311 Tubing nipple (M6) which belongs to the 6.1602.160 Bottle attachment accessories can be used together with the included second E.301.0021 O-ring for recycling operation (see section 5.2.3), in which the outlet capillary from the detector block is led back into eluent bottle **67**.

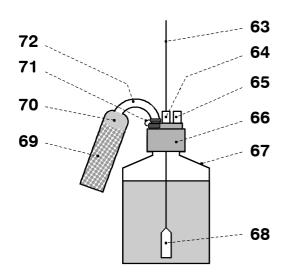


Fig. 11: Connection of eluent bottle

63	Aspirating tubing (6.1834.010)
64	<b>Tubing nipple (4.420.4300; M8)</b> with E.301.0021 O-ring
65	Threaded stopper (6.1446.040; M6)
66	Bottle attachment (6.1602.105)
67	Eluent bottle (6.1608.070)
68	Aspirating filter (6.2821.090)
69	CO <sub>2</sub> absorber
70	Cotton wool
71	SGJ clip (6.2023.020)
72	Absorber tube (6.1609.000)

#### 2.6.4 Deaerating the pump and rinsing the pulsation dampener

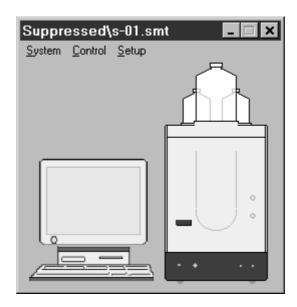
The first time that it is started up the high-pressure pump must be deaerated. Proceed as follows:

#### 1 Prepare for deaeration

- Open the rotary knob on purge valve **39** by approx. ½ turn in the counterclockwise direction (see *Fig. 3* and *Fig. 4*).
- Remove the plastic stopper from connection **8** on the front panel of 761 Compact IC (see *Fig. 1*).
- Push 6.2816.020 Syringe (without needle) into connection 8 until the stop is reached.

#### 2 Open and connect system

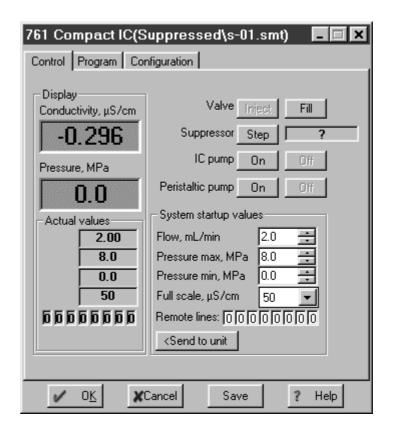
- Start the «761 Compact IC» PC program, if it has not been already been started (see section 2.5.3).
- Select File / Open / System in the main window. Open either the subfolder Non-suppressed (for 2.761.0010 Compact IC without suppressor) or the subfolder Suppressed (for 2.761.0020 Compact IC with suppressor). Select the system file n-01.smt or s-01.smt and click on <Open>. The corresponding system window is opened:



 Select the Connect to workplace item of the Control menu in this window.

#### 3 Set flow rate to 2 mL/min

- Double-click the 761 icon in the system window to open the window for manual control of the 761 Compact IC (see below)
- Set the flow rate to 2 mL/min in the Flow field.
- Click to <Send to unit> to send this value to the 761 Compact IC.



#### 4 Deaerate pump

- Make sure that the aspirating tubing **63** for the high-pressure pump has been immersed into the eluent.
- Click the <On> button for IC pump to switch on the highpressure pump.
- Use the syringe inserted into connection **8** to aspirate air until eluent flows into the syringe.
- Click the <Off> button for IC pump to switch off the highpressure pump.
- Close the rotary knob on purge valve **39** by turning it in a clockwise direction (see *Fig. 3* and *Fig. 4*).
- Remove the syringe from connection 8.

#### **5** Rinse pulsation dampener

- Place a beaker beneath the column connection capillary 28.
- Click the <On> button for IC pump to switch on the highpressure pump and rinse the pulsation dampener 60 filled with isopropanol for ca. 10 min with eluent.
- Click the <Off> button for IC pump to switch off the highpressure pump.

#### 6 Reduce flow rate

- Reset the original flow rate under Flow (e.g. 0.5 mL/min).
- Click **<Send to unit>** to send this value to the 761 Compact IC.

## 2.7 Precolumns and separating columns

#### 2.7.1 General information on precolumns

The use of easily exchangeable precolumns protects the separating columns and prolongs their lifetime. The precolumns available from Metrohm (see section 6.3.2) are either real precolumns or precolumn cartridges, which are used together with the 6.2821.040 Cartridge head or the 6.2828.010 Precolumn cartridge holder.



New IC precolumns are normally filled with solution and sealed at both ends. Before the precolumn is installed in the system, it must be ensured that this solution is freely miscible with the eluent used (check manufacturer's specifications).

#### 2.7.2 Precolumns with cartridge head

The 6.1005.020, 6.1005.040, 6.1005.050, 6.1007.010 and 6.1010.010 Precolumn cartridges are installed in the 6.2821.040 Cartridge head as follows (see *Fig. 12*):

#### 1 Prepare separating column

- Remove end caps from separating column **81**.
- Unscrew fastening screw from column inlet.
- Take steel connector **74** for ferrule out of fastening screw.

#### 2 Install cartridge

- Remove end caps for precolumn cartridge **77** (the steel mesh **76** and gaskets **75** are already installed in the cartridge).
- Mount steel spacer 78 on Manufit pressure screw 80 (the steel mesh 79 and gaskets 75 are already installed in the pressure screw).
- Mount precolumn cartridge **77** on the steel spacer **78** (comply with flow direction if specified on the precolumn).
- Screw Manufit pressure screw 80 firmly to separating column 81.
- Mount steel connector **74** for ferrule on the inlet side of the precolumn cartridge **77**.
- Screw on Manufit housing **73** with Manufit pressure screw **80**.

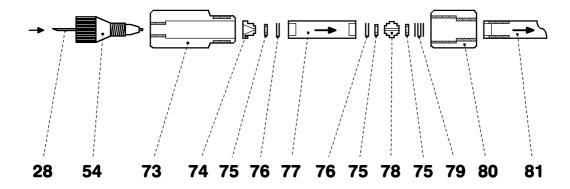


Fig. 12: Installation of precolumn cartridges with cartridge head

28	Column connection capillary from injector	77	Precolumn cartridge
54	Compression fitting (6.2744.010)	78	Steel spacer (6.2821.080)
73	Manufit housing	79	4 Steel meshes (6.2821.020)
74	Steel connector (of IC separating column)	80	Manufit pressure screw
75	PTFE gasket (6.2821.010)	81	IC separating column
76	2 Steel meshes (6.2821.020)		

### 2 Steel meshes (6.2821.020)

#### **Connect precolumn**

- Fit PEEK compression fitting **54** to the column connection capillary 28 mounted on the injection valve (see section 2.3.5).
- Screw column connection capillary **28** to Manufit housing **73**.



The precolumn cartridge built into the cartridge head can only be rinsed together with the separating column (see section 2.7.7/8). This increases the rinsing time by approx. 20 min.

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#### 2.7.3 Precolumn glass cartridges with cartridge holder

The precolumn glass cartridge METROSEP Anion Dual 1 (6.1006.030) is inserted in the 6.2828.010 Precolumn cartridge holder as follows (see *Fig. 13*):

#### 1 Insert cartridge

- Remove end fittings 82 from screw caps 83.
- Insert precolumn cartridge **85** in sleeve **84** (mark flow direction of the column cartridge on sleeve, the column cartridge should always be operated in the same flow direction).
- Screw both screw caps **83** loosely to sleeve **84** by hand.
- Screw both end fittings **82** into screw caps **83** so that their capillary ends are seated in the PTFE seals of the cartridge.
- Tighten both screw caps 83 by hand.

#### 2 Connect precolumn

- Provide the column connection capillary **28** mounted on the injection valve with a PEEK compression fitting **54** (see section 2.3.5) and screw it tightly onto end fitting **82** on the inlet side of the precolumn.
- Cut off as short a piece as possible from PEEK capillary 56
  and fit this with two PEEK compression fittings 54 (see section 2.3.5).
- Screw prepared capillary **56** to end fitting **82** on the other end of the precolumn.

#### 3 Rinse precolumn

- Place a beaker beneath the outlet capillary of the precolumn.
- Open software window for manual system control.
- If necessary, modify Flow rate to the value suited for the inserted separating column and click on <Send to unit> to send this value to the 761 Compact IC.
- Switch on high-pressure pump (IC pump) by clicking <On>
  and rinse precolumn with eluent for ca. 10 min.
- Switch off high-pressure pump by clicking **<Off>**.



The rinsed precolumn can be connected directly to the separating column instead of by means of a PEEK capillary. Proceed as follows:

- Screw off PEEK capillary from end fitting **82** of the precolumn.
- Screw off end fitting **82** from screw cap **83** of the precolumn.
- Screw connection piece 86 onto screw cap 83.
- Screw screw cap 87 of separating column 81 onto connection piece 86.

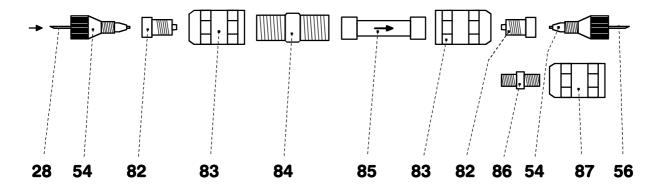


Fig. 13: Installation of precolumn glass cartridges with cartridge holder

28	Column connection capillary from injector	84	Sleeve for precolumn cartridge
54	Compression fitting (6.2744.010)	85	Precolumn cartridge (6.1006.030)
56	PEEK capillary (6.1831.010)	86	Connection piece for connection precolumn – column
82	End fitting	87	Screw cap for column

83 Screw cap for precolumn

#### 2.7.4 IC anion precolumn SUPERSEP

The 6.1009.010 IC Anion Precolumn SUPERSEP has two connections for PEEK capillaries and is installed as follows:

#### 1 Connect precolumn

- Remove end caps from the precolumn.
- Fit PEEK compression fitting **54** to column connection capillary **28** mounted on the injection valve (see section 2.3.5).
- Screw precolumn to column connection capillary 28.
- Cut a piece from the 6.1831.010 PEEK capillary as short as possible and fit with PEEK compression fittings 54 (see section 2.3.5).
- Fasten the prepared capillary to the other end of the precolumn.

#### 2 Rinse precolumn

- Place a beaker beneath the outlet capillary of the precolumn.
- Open software window for manual system control.
- If necessary, modify Flow rate to the value suited for the inserted separating column and click on <Send to unit> to send this value to the 761 Compact IC.
- Switch on high-pressure pump (IC pump) by clicking <On> and rinse precolumn with eluent for ca. 10 min.
- Switch off high-pressure pump by clicking **<Off>**.

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#### 2.7.5 General information on separating columns



New IC separating columns are normally filled with solution and sealed at both ends. Before the column is installed in the system, it must be ensured that this solution is freely miscible with the eluent used (check manufacturer's specifications).

The IC separating columns and precolumns currently available from Metrohm are listed in section 6.3.2. A test chromatogram and an information leaflet is provided with each column. You will find additional information concerning these columns in the 8.732.2003 Metrohm Monograph «Ion chromatography» and in special "Application Bulletins", which are available on request free of charge from your local Metrohm agency.



When you install the column, always ensure that this is inserted correctly in accordance with the flow direction shown (arrow must point <u>upwards</u>).

#### 2.7.6 Selection of the sample loop

Selection of the sample loop depends on the separating column used. Normally, the following sample loops are used:

Cation columns 10  $\mu$ L Anion columns with suppressor 20  $\mu$ L Anion columns without suppressor 10  $\mu$ L

Depending on the instrument version, the following sample loops are installed in the 761 Compact IC 761:

Version	Sample loop	Volume
2.761.0010	6.1825.230 (PEEK)	10 μL
2.761.0020	6.1825.210 (PEEK)	20 μL

If desired, the built-in sample loop can be replaced by one of the sample loops available as an option (see section 6.3.1).



#### 2.7.7 Installation of the separating column without suppressor

With the 2.761.0010 Compact IC without suppressor module, the IC separating column is installed as follows (see Fig. 14):

#### 1 Connect column to injector

- Remove end caps from column 81.
- without precolumn:
   Screw inlet end of separating column 81 (note flow direction) to column connection capillary 28 mounted on the injector.
- with precolumn in cartridge head:
   Install separating column 81 (note flow direction) in the cartridge head (see Fig. 12) as described in section 2.7.2.
- with precolumn in cartridge holder:
   Screw inlet end of separating column 81 (note flow direction) to the precolumn fixed in the cartridge holder as described in section 2.7.3 (see Fig. 13).
- with IC anion precolumn SUPERSEP:
   Screw inlet end of separating column 81 (note flow direction) to the precolumn installed as described in section 2.7.4.

#### 2 Rinse column

- Place a beaker beneath the column outlet.
- Open software window for manual system control.
- If necessary, modify Flow rate to the value suited for the inserted separating column and click on <Send to unit> to send this value to the 761 Compact IC.
- Switch on high-pressure pump (IC pump) by clicking <On>
  and rinse column with eluent for ca. 10 min.
- Switch off high-pressure pump by clicking <Off>.

#### 3 Connect column to detector block

Screw outlet end of separating column 81 to the inlet capillary
 45 permanently mounted on the detector block.

#### 4 Fix column

• Insert one or two column holders **88** (6.2027.030, 6.2027.040 or 6.2027.050) in the mounting rails **27** and fasten separating column **81** in the column holder **88**.

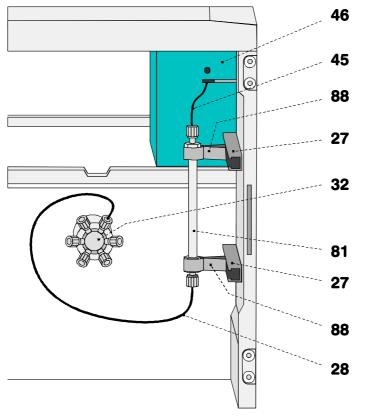


Fig. 14: Installation of column without suppressor

## **27 Mounting rail** for column holder **88**

## 28 Column connection capillary

Connection injection valve **32** – separating column **81** 

#### 32 Injection valve

# 45 Inlet capillary for detector block

PEEK capillary, fixed mounting

## 46 Detector block (1.733.0110)

- 81 Separating column
- 88 Column holder (6.2027.0X0)

#### 2.7.8 Installation of the separating column with suppressor

With the 2.761.0020 Compact IC with suppressor module, the IC separating column is first connected to the injector or precolumn. The connection to the suppressor module and the detector block is described in section 2.8.

#### 1 Connect column to injector

- Remove end caps from column 81.
- Screw inlet end of separating column **81** (note flow direction) to column connection capillary **28** or to the already installed precolumn (procedure see section 2.7.7).

#### 2 Rinse column

- Place a beaker beneath the column outlet.
- Open software window for manual system control.
- If necessary, modify Flow rate to the value suited for the inserted separating column and click on <Send to unit> to send this value to the 761 Compact IC.
- Switch on high-pressure pump (IC pump) by clicking <On>
  and rinse column with eluent for ca. 10 min.
- Switch off high-pressure pump by clicking **<Off>**.

#### 3 Fix column

• Insert one or two column holders **88** (6.2027.030, 6.2027.040 or 6.2027.050) in the mounting rails **27** and fasten separating column **81** in the column holder **88**.

### 2.8 Suppressor module

#### 2.8.1 General information on suppressor module

The **Metrohm Suppressor Module MSM** for chemical suppression installed in the 2.761.0020 Compact IC comprises a total of 3 suppressor units which are in turn used for suppression, regenerated with sulfuric acid and rinsed with water. To record every new chromatogram under comparable conditions, work is normally carried out with freshly regenerated suppressor. Switching is either automatic together with the valve switching or manual.



The suppressor units must never be regenerated with  $H_2SO_4$  in the same flow direction used for the eluent. You should thus always install the inlet and outlet capillaries as described in section 2.8.4 according to the scheme shown in Fig. 18.

For operation of the suppressor module, the **two-channel peristaltic pump** built into the 2.761.0020 Compact IC is used which conveys the regeneration solution (normally **20 mmol/L H\_2SO\_4) and the rinsing solution (normally <b>dist. H\_2O)** to the suppressor units (flow rate of 0.5 mL/min).

The three inlets and outlets numbered 1...3 on the suppressor module each have 2 permanently mounted PTFE capillaries, which must be connected as described in section 2.8.4 (see Fig. 16 and Fig. 18).

To avoid contamination of the suppressor module by foreign particles or bacterial growth, the two **6.2821.120 Filter units PEEK** (see section 2.3.6) supplied with the 2.761.0020 Compact IC must be installed between the peristaltic pump and the inlet capillaries of the suppressor module.



The suppressor module must <u>never</u> be switched in the dry state as there is a danger of blocking. Before every switching operation of the suppressor module, the three suppressor units must have been rinsed for at least ½ h with eluent, regeneration and rinsing solution.

#### 2.8.2 Preparation of the peristaltic pump

Before start-up the accessories for the 2-channel peristaltic pump built into the 761 Compact IC must be mounted according to *Fig. 15*. Proceed as follows:

#### 1 Attach pump tubings

Loosen both tubing cartridges 48 mounted above pump drive
 52 from the holding clamp 50 by pressing down snap-action lever 51 and remove from mounting pin 53 (see Fig. 16).

- Press contact pressure lever **49** on both tubing cartridges down as far as it will go.
- Insert the pump tubings **92** and **93** (6.1826.060) into each of the tubing cartridges as shown in *Fig. 15*. The white-yellow stopper **94** must click into the corresponding holder on the left-hand side of the tubing cartridge.
- Place the tubing cartridges on mounting pin 53 and press down on the right-hand side until snap-action lever 51 clicks into position on holding clamp 50. Take care that no kinks are formed in the pump tubing.

#### 2 Install Filter units PEEK

- Mount a coupling **91** (6.2744.030) to the outlet end of the two Pump tubings **92** and **93**.
- Attach a piece of PTFE tubing **95** (6.1803.020) cut to the required length (normally ca. 10 cm) using a compression fitting **54** (6.2744.010) to the other end of this coupling.
- Attach the PTFE tubing **95** using a compression fitting **54** (6.2744.010) to the filter-screw **57** of the filter unit PEEK (see section 2.3.6).

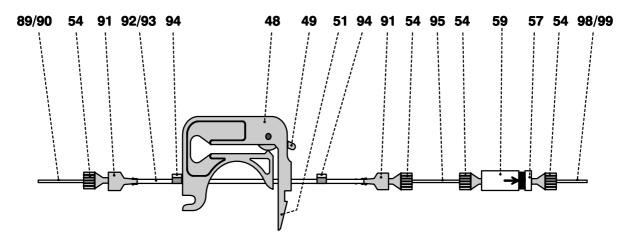


Fig. 15: Installing pump tubings

48	Tubing cartridge	91	Coupling (6.2744.030)
49	Contact pressure lever	92	Pump tubing (6.1826.060) for H <sub>2</sub> SO <sub>4</sub>
51	Snap-action lever	93	Pump tubing (6.1826.060) for H <sub>2</sub> O
54	PEEK compression fitting (6.2744.010)	94	Stopper (white-yellow)
57	Filter-Screw of Filter Unit Part of 6.2821.120 Filter unit	95	PTFE tubing (6.1803.020)
59	Filter-Housing of Filter Unit Part of 6.2821.120 Filter unit	98	Suppressor inlet capillary for H <sub>2</sub> O
89	Aspirating tubing for H₂O	99	Suppressor inlet capillary for H <sub>2</sub> SO <sub>4</sub>
90	Aspirating tubing for H <sub>2</sub> SO <sub>4</sub>		

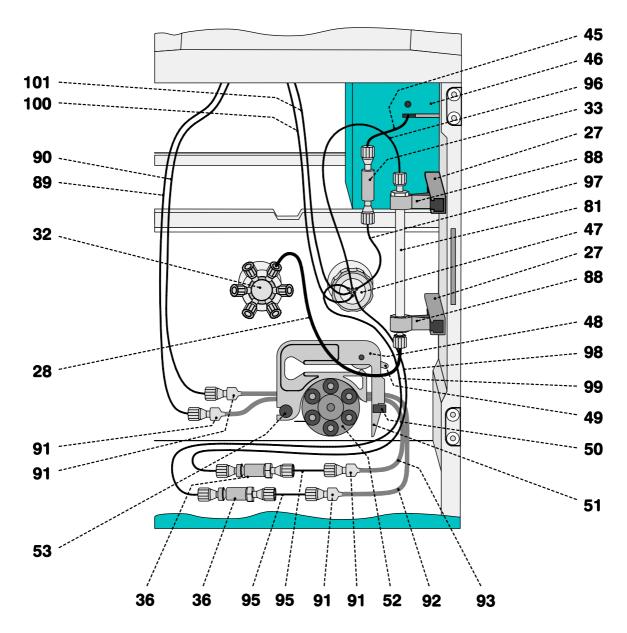


Fig. 16: Connection of the separating column with suppressor

27	<b>Mounting rail</b> for column holder <b>88</b>	81	Separating column
28	Column connection capillary PEEK capillary (6.1831.010; 30 cm)	88	Column holder (6.2027.0X0)
32	Injection valve	89	Aspirating tubing for H₂O 6.1803.020 PTFE tubing
33	PEEK coupling (6.2744.040)	90	Aspirating tubing for H₂SO₄ 6.1803.020 PTFE tubing
36	Filter unit PEEK (6.2821.120)	91	Coupling (6.2744.030)
45	Inlet capillary to detector block (fixed mounting)	92	Pump tubing (6.1826.060) for H <sub>2</sub> SO <sub>4</sub>
46	Detector block (1.733.0110)	93	Pump tubing (6.1826.060) for H <sub>2</sub> O

47	Suppressor module	95	PTFE tubing (6.1803.020)
48	<b>Tubing cartridge (6.2755.000)</b> for pump tubings <b>92/93</b>	96	Suppressor inlet capillary for eluent
49	Contact pressure lever for adjusting the contact pressure	97	Suppressor outlet capillary for eluent
50	Holding clamp for locking the tubing cartridge into place	98	Suppressor inlet capillary for H₂O
51	Snap-action lever for releasing the tubing cartridge	99	Suppressor inlet capillary for H <sub>2</sub> SO <sub>4</sub>
52	Pump drive Roller head with contact rollers	100	Suppressor outlet capillary for H <sub>2</sub> O
53	Mounting pin for attaching the tubing cartridge	101	Suppressor outlet capillary for H <sub>2</sub> SO <sub>4</sub>

#### 2.8.3 Connection of supply bottles

The supply lines for the regeneration and rinsing solution between the storage bottles and the peristaltic pump are installed as follows (see *Fig. 17*):

#### 1 Prepare supply bottle for H<sub>2</sub>SO<sub>4</sub>

- Prepare regeneration solution suited for the desired application and separating column (normally 20 mmol/L H<sub>2</sub>SO<sub>4</sub>).
- Fill regeneration solution into supply bottle **103** (amber glass, 1 L) and label the bottle.
- Screw bottle attachment **102** on to supply bottle **103**.
- Place supply bottle **103** at one of the rear positions of the bottle rack **10** on the 761 Compact IC (see *Fig. 1*).

#### 2 Connect aspirating tubing for H<sub>2</sub>SO<sub>4</sub>

- Prepare aspirating tubing **90**: Cut a piece of the 6.1803.020 PTFE tubing to the required length (normally ca. 120 cm).
- Pull one end of aspirating tubing 90 through a PEEK compression fitting 54 (6.2744.010) so that approx. 30 cm of the tubing projects.
- Screw PEEK compression fitting 54 with tubing into one opening of bottle attachment 102 attached to the regeneration solution storage bottle and tighten it so that the tubing is firmly held.
- Insert the free end of aspirating tubing **90** into one of the openings in **12** of the 761 Compact IC (see *Fig. 2*) from above and pull it sufficiently far into the interior.
- Mount a coupling **91** (6.2744.030) to the inlet end of the rear

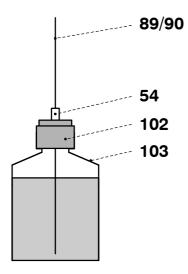
- pump tubing 92.
- Mount a 6.2744.010 Compression fitting at the end of the aspirating tubing **90** and screw this compression fitting on to the coupling **91** (see *Fig. 16*).

#### 3 Prepare supply bottle for H<sub>2</sub>O

- Prepare rinsing solution suited for the desired application and separating column (normally dist. H<sub>2</sub>O).
- Fill rinsing solution into supply bottle **103** (amber glass, 1 L) and label the bottle.
- Screw bottle attachment 102 on to supply bottle 103.
- Place supply bottle **103** beside the other supply bottle on the bottle rack **10** on the 761 Compact IC (see *Fig. 1*).

#### 4 Connect aspirating tubing for H<sub>2</sub>O

- Prepare aspirating tubing **89**: Cut a piece of the 6.1803.020 PTFE tubing to the required length (normally ca. 120 cm).
- Pull one end of aspirating tubing 89 through a PEEK compression fitting 54 (6.2744.010) so that approx. 30 cm of the tubing projects.
- Screw PEEK compression fitting 54 with tubing into one opening of bottle attachment 102 attached to the rinsing solution storage bottle and tighten it so that the tubing is firmly held.
- Insert the free end of aspirating tubing 89 into one of the openings in 12 of the 761 Compact IC (see Fig. 2) from above and pull it sufficiently far into the interior.
- Mount a coupling **91** (6.2744.030) to the inlet end of the front pump tubing **93**.
- Mount a 6.2744.010 Compression fitting at the end of the aspirating tubing 89 and screw this compression fitting on to the coupling 91 (see Fig. 16).



54	PEEK compression fitting (6.2744.010)
89	<b>Aspirating tubing for H₂O</b> 6.1803.020 PTFE tubing
90	Aspirating tubing for H₂SO₄ 6.1803.020 PTFE tubing
102	Bottle attachment (6.1602.150)
103	Supply bottle (6.1608.023)

Fig. 17: Connection of supply bottles

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#### 2.8.4 Connection of the suppressor module

The three inlets and outlets numbered 1...3 on the suppressor module **47** each have 2 permanently mounted PTFE capillaries, which must be connected as described as follows (see *Fig. 16* and *Fig. 18*).

#### 1 Inlet capillary for eluent

 Screw inlet capillary 96 marked with "Eluent" of suppressor module 47 to outlet end of separating column 81 using a 6.2744.010 Compression fitting.

#### 2 Outlet capillary for eluent

- Screw outlet capillary 97 marked with "Detector" of suppressor module 47 to coupling 33 using a 6.2744.010 Compression fitting.
- Screw inlet capillary **45** of detector block **46** to other end of coupling **33**.

#### 3 Inlet capillary for H<sub>2</sub>SO<sub>4</sub>

Attach inlet capillary 99 marked with "H<sub>2</sub>SO<sub>4</sub>" of suppressor module 47 using a 6.2744.010 Compression fitting to the filter unit PEEK 36 connected to the rear pump tubing 92.

#### 4 Outlet capillary for H<sub>2</sub>SO<sub>4</sub>

- Pull outlet capillary 101 marked with "Waste" of the suppressor module 47 from below through one of the openings 13 out of the inner compartment of the 761 Compact IC.
- Lead outlet capillary **101** to a sufficiently large waste container and fix it in place.

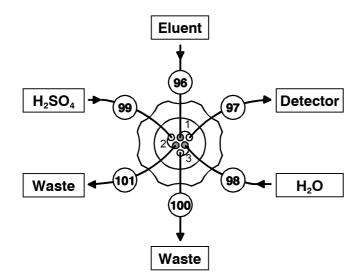


Fig. 18: Connections at suppressor module

96	Suppressor inlet capillary for eluent
97	Suppressor outlet
	capillary for eluent
98	Suppressor inlet
	capillary for H₂O
99	Suppressor inlet
	capillary for H₂SO₄
100	Suppressor outlet
	capillary for H₂O
101	Suppressor outlet
	capillary for H <sub>2</sub> SO <sub>4</sub>



#### 5 Inlet capillary for H<sub>2</sub>O

 Attach inlet capillary 98 marked with "H<sub>2</sub>O" of suppressor module 47 using a 6.2744.010 Compression fitting to the filter unit PEEK 36 connected to the front pump tubing 93.

#### 6 Outlet capillary for H<sub>2</sub>O

- Pull outlet capillary 100 marked with "Waste" of the suppressor module 47 from below through one of the openings 13 out of the inner compartment of the 761 Compact IC.
- Lead outlet capillary **100** to a sufficiently large waste container and fix it in place.

#### 7 Fasten capillaries to the side walls

- If necessary, the two aspirating tubings 89 and 90 can be fixed in the required position in the interior with the help of a Y.107.0150 self-adhesive strap.
- If necessary, the two outlet capillaries 100 and 101 can be fixed in the required position in the interior with the help of a Y.107.0150 self-adhesive strap.

### 2.9 Putting into operation

#### 2.9.1 Putting into operation without suppressor

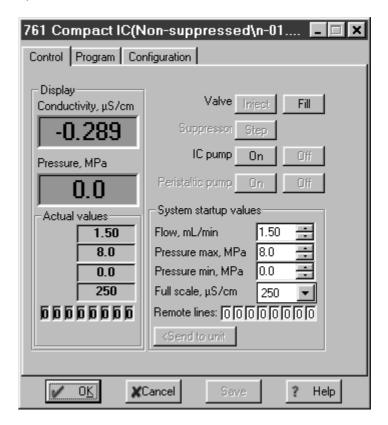
Before sample solutions can be injected at the **2.761.0010 Compact IC** (without suppressor), the entire system must be tested for leaks and then conditioned with eluent until the baseline is stable. Proceed as follows:

#### 1 Open and connect system

- Start the «761 Compact IC» PC program, if it has not already been started (see section 2.5.3).
- Select File / Open / System in the main window. Select the system file suited for the inserted separating column (e.g. n-01.smt) and click on <Open>.
- Select the Connect to workplace item from the Control menu of the system window.

#### 2 Open control window

 Double-click the 761 icon in the system window. The control window for manual control of the 761 Compact IC appears, which indicates conductivity, pressure and current system parameters.





#### 3 Start system

- Make sure, that the aspirating tubing 63 for the high-pressure pump is immersed in the eluent.
- Select Startup hardware (Measure baseline) from the Control
  menu in the system window. The high-pressure pump is
  started, at the same time, a chromatogram window is opened
  where the baseline is recorded continuously.

#### 4 Check for leaks

 Check all capillaries and their connections between the highpressure pump and the detector block for escaping liquid. If eluent escapes anywhere, the appropriate compression fitting must be tightened further or changed.

#### 5 Condition system

- Rinse the system with eluent until the desired stability of the baseline is reached (normally 30...60 min; if the eluent is changed, the establishment of the ion exchanger equilibrium on the separating column can take longer).
- The instrument is now ready for sample determinations using the selected system.

#### 2.9.2 Putting into operation with suppressor

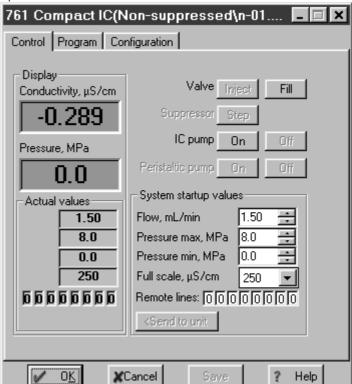
Before sample solutions can be injected at the **2.761.0020 Compact IC** (with suppressor), the entire system must be tested for leaks and then conditioned with eluent until the baseline is stable. At the same time, the suppressor module must be conditioned. Proceed as follows:

#### 1 Open and connect system "Prep-MSM\*.smt"

- Start the «761 Compact IC» PC program, if it has not already been started (see section 2.5.3).
- Select File / Open / System in the main window. Open the folder Suppressed and select the system file Prep-MSM1.smt (for anion column Dual 1) or Prep-MSM2.smt (for anion column Dual 2) suited for the inserted separating column. Click on <Open>.
- Select the Connect to workplace item from the Control menu of the system window.

#### 2 Open control window

 Double-click the 761 icon in the system window. The control window for manual control of the 761 Compact IC appears, which indicates conductivity, pressure and current system



parameters.

#### 3 Start system

- Make sure, that the aspirating tubing **63** for the high-pressure pump is immersed in the eluent.
- Select Startup determination from the Control menu in the system window. The high-pressure pump and the peristaltic pump are started, at the same time, a chromatogram window is opened where the baseline is recorded continuously. The suppressor module is automatically switched to the next position every 20 min and conditioned in this way.

#### 4 Set contact pressure for pump tubings

- Press contact pressure lever 49 on both tubing cartridges 48
  upwards until regeneration and rinsing solution just start to be
  drawn in.
- Then press contact pressure lever **49** upwards until it clicks once more to obtain optimal contact pressure.

#### 5 Check for leaks

 Check all capillaries and their connections between the highpressure pump and the detector block for escaping liquid. If eluent escapes anywhere, the appropriate compression fitting must be tightened further or changed.



#### 6 Condition system

• Rinse the system with eluent until the desired stability of the baseline is reached (normally 30...60 min; if the eluent is changed, the establishment of the ion exchanger equilibrium on the separating column can take longer). After this time the suppressor module is sufficiently conditioned too.

• The instrument is now ready for sample determinations using the selected system.



Pump tubings are consumable material with a lifetime which depends on the contact pressure. This is why the tubing cartridges should be raised completely by loosening snap-action lever on the right-hand side if the pump is to remain switched off for a considerable length of time (the set contact pressure remains unchanged).

#### 2.10 Connection of external devices

#### 2.10.1 Connection of the 750 Autosampler

The 750 Autosampler available from Metrohm as an option is an automatic sampler for ion chromatography. It accommodates max. 128 samples each of 730  $\mu$ L, which are automatically transferred to the sample loop attached to the injection valve of the 761 Compact IC. The samples on the 750 Autosampler are changed by a remote signal which is outputted by the 761 Compact IC (761 Compact IC as "Master").

In order to connect the 750 Autosampler the 6.2128.160 Cable which is available as an option is additionally required. The 750 Autosampler is connected to the 761 Compact IC as follows:

#### 1 Electrical connection 761 – 750

- Connect 6.2140.010 Connector plug to contact closure strip **9** of the 750 Autosampler (see 750 Instructions for Use).
- Connect one end of the 6.2128.160 Cable to the connections 13 "EXTERNAL INJECT INPUT" and 14 "GND" on the connector plug. The two cable ends are appropriately inscribed with "750" (white) and "GND" (brown).
- Connect the other end of the 6.2128.160 Cable to the remote interface **24** of the 761 Compact IC (see *Fig. 2*).

#### 2 Tubing connection 750 - injection valve

- Loosen rotary nipple **31** which is screwed onto the inner side of connection **3** of the 761 Compact IC (see *Fig. 3* and *Fig. 4*).
- Pull aspiration tubing **4** completely out from connection **3** and screw it off from connection "1" of injection valve **32**.
- Cut transfer tubing **8** (see 750 Instructions for Use) to the shortest possible length between the needle of the 750 Autosampler and injection valve **32** of the 761 Compact IC.
- Pull the free end of transfer tubing <u>8</u> through the opening of connection 3 on the 761 Compact IC and screw it onto connection "1" of injection valve <u>32</u> using a 6.2744.010 PEEK compression fitting.
- Retighten rotary nipple on the interior side of connection **3** to fix the transfer tubing.

#### 3 Tubing connection injection valve - waste

- Insert 6.2744.020 Coupling (from 761 accessories) into connection **3** of the 761 Compact IC.
- Screw aspirating tubing 4 onto the 6.2744.020 Coupling and lead it into the waste container.

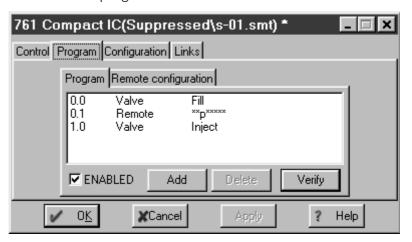


#### 4 Settings at the 750 Autosampler

• Set parameter "Run time" to 0.0 to activate the external start of the 750 Autosampler (see section 3.5 of the 750 Instructions for Use). Filling the sample loop of the injection valve only takes place when the impulse produced by the 761 Compact IC at remote output 3 (see point 5) makes contact at the "EXTERNAL INJECT INPUT" of the 750 Autosampler. The run time will then be determined externally.

#### 5 Settings in the «761 Compact IC» program

 A time program must be drawn up for the selected system which first switches the injection valve to the "Fill" position, then produces an impulse at remote lead 3 to start sample addition by the 750 Autosampler and finally switches the injection valve to the "Inject" position. The following example shows such a program:



#### 2.10.2 Connection of the 766 IC Sample Processor

The 766 IC Sample Processor available from Metrohm as an option is an automatic sampler for ion chromatography. It accommodates max. 127 samples each of 2.5 mL or 11 mL, which are automatically transferred to the sample loop attached to the injection valve of the 761 Compact IC. Sample changing and filling the sample loop are started by a remote signal produced by the 761 Compact IC (761 Compact IC as "Master").

The 6.2141.110 Cable is required for connecting the 766 IC Sample Processor. The 766 IC Sample Processor is connected to the 761 Compact IC as follows:

#### 1 Electrical connection 761 - 766

 Connect the end of the 6.2141.110 Cable marked with "766" to the remote interface <u>22</u> of the 766 IC Sample Processor (see 766 Instructions for Use). • Connect the end of the 6.2141.110 Cable marked with "732/1" to the remote interface **24** of the 761 Compact IC (see *Fig. 2*).

#### **2** Tubing connection 766 – injection valve

- At the 761 Compact IC, loosen the rotary nipple **31** screwed onto the interior side of connection **3** (see *Fig.* 3 and *Fig.* 4).
- Take aspirating tubing 4 completely out of connection 3 and unscrew from connection "1" of injection valve 32.
- Cut PEEK capillary **18** installed at the 766 IC Sample Processor (see 766 Instructions for Use) to the desired length.
- Pull the free end of PEEK capillary tubing <u>18</u> through the opening of connection 3 of the 761 Compact IC and screw it onto connection "1" of injection valve <u>32</u> with the help of a 6.2744.010 PEEK compression fitting.
- Retighten rotary nipple 31 on the interior side of connection 3 to fix the capillary.

#### 3 Tubing connection injection valve - waste

- Insert 6.2744.020 Coupling (from 761 accessories) into connection 3 of the 761 Compact IC.
- Screw aspirating tubing 4 onto the 6.2744.020 Coupling and lead it into the waste container.

#### 4 Settings at the 766 IC Sample Processor

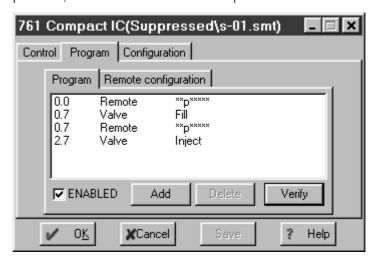
 For operation with the 761 Compact IC it is recommended that the following program "761" is used with the 766 IC Sample Processor:

parameters			
method		761	
number of sam	ples:	Rack	
>start sequence	!		
1 CTL:Rm:		INIT	
>sample sequenc	e		
1 SCN:Rm	:	Wait1	
2 MOVE 1	:	sample	
3 LIFT: 1 :		work	mm
4 SCN:Rm	:	Wait1	
5 PUMP 1.1	:	120	S
>final sequence	!		
>changer settin	ıgs		
rack number		0	
lift rate 1		12	mm/s
shift rate		20	
>manual stop			
CTL Rmt:	*****	******	
CTL RS232:			
		-	

- Method name
- Number of samples to be processed (entire sample rack)
- Initialize remote interface
- Waiting until 761 Compact IC sends signal on remote line 3
- Move needle to sample position
- Place lift with needle to working position
- Waiting until 761 Compact IC sends signal on remote line 3
- Fill sample loop with sample during 120 s
- Settings for changer functions
- Reaction to manual stop

#### 5 Settings in the «761 Compact IC» program

A time program must be drawn up for the selected system
which first produces an impulse at remote lead 3 to start
sample changing on the 766 IC Sample Processor and then
switches the injection valve to the "Fill" position. A further impulse is then produced to start filling the sample loop within
120 s. At the end the injection valve is switched to the "Inject"
position, which also starts data acquisition.



#### 6 Run conditions

In order that the joint operation of 761 – 766 functions properly the following conditions must be fulfilled:

- The program must be started first on the 766 IC Sample Processor, then the "Sample Queue" on 761 Compact IC.
- Remote lead 3 on the Compact IC must be set to 0 at the start of each determination (Set System startup values:
   Remote line 3 = 0).

#### 2.10.3 Connection of other devices

Any external devices can be connected to the 25-pin remote interface **24** (see *Fig. 2*). The 8 output lines can be used to control these external devices.



Before an external device is connected to the remote interface **24**, the 761 Compact IC must always be switched off using mains switch **19**!

The pin assignment of the remote interface, its functions and the electrical requirements and the conditions are described in section 6.1.

## 3 Operating tutorial



This section introduces you to the operation of the 761 Compact IC by means of a brief operating tutorial which describes the basic operating steps needed for the recording of an ion chromatogram using one of the system files supplied.

The determination of the anionic content of a drinking water sample with the METROSEP Anion Dual 2 column with chemical suppression is used an illustrative example. Please note that the steps and parameter settings described apply only to this example. If you use a different separating column and system file, the procedures described in the tutorial must be modified appropriately.

For further explanations of the operation, please refer to section 4.

## 3.1 Requirements

For the determination of anions in drinking water described in this tutorial, the following instruments, accessories and solutions are required:

- 2.761.0020 Compact IC with suppressor module
- 6.1825.210 Sample loop (20 μL, PEEK) already integrated in the 2.761.0020 Compact IC
- 6.1006.100 IC anion column METROSEP Anion Dual 2
- Eluent

2 mmol/L NaHCO $_3$  / 1.3 mmol/L Na $_2$ CO $_3$  in dist. H $_2$ O flow: 0.8 mL/min

Standard

Standard solution with 0.5 mg/L F<sup>-</sup>, 5 mg/L Cl<sup>-</sup> and 10 mg/L each of  $NO_3^-$  and  $SO_4^{2-}$  (in dist.  $H_2O$ )



## 3.2 Preparations

Before you start this brief tutorial, the entire IC system must be correctly installed as described in section 2. In what follows, the most important points for the installation are described once again (for details, see the sections mentioned).

$\Rightarrow$	Setting up instrument	section 2.2
$\Rightarrow$	Installing and connecting detector block	section 2.3.1
$\Rightarrow$	Mounting syringe and aspirating tubing	section 2.3.2
$\Rightarrow$	Mounting drain tubes	section 2.3.3/4
$\Rightarrow$	Mains connection	section 2.4
$\Rightarrow$	Connection to PC	section 2.5

#### 2 Prepare eluent

⇒ Preparing eluent:
 2 mmol/L NaHCO<sub>3</sub> / 1.3 mmol/L Na<sub>2</sub>CO<sub>3</sub> in dist. H<sub>2</sub>O

⇒ Microfiltering and degassing eluent section 5.1.3

#### 3 Install high-pressure pump

$\Rightarrow$	Removing transport security screws	section 2.6.1
$\Rightarrow$	Mounting pulsation dampener	section 2.6.2
$\Rightarrow$	Installing eluent supply	section 2.6.3
$\Rightarrow$	Degassing pump	section 2.6.4

### 4 Connecting separating column and suppressor

$\Rightarrow$	Connecting IC anion column	section 2.7.8
$\Rightarrow$	Connecting peristaltic pump	section 2.8.2
$\Rightarrow$	Connecting supply bottles	section 2.8.3
$\Rightarrow$	Connecting suppressor module	section 2.8.4
$\Rightarrow$	Conditioning system	section 2.9.2

#### 3.3 Calibration

After the complete IC system has been installed as described in section 3.2, the first calibration can be started. A standard solution is required for this; it should contain the substances to be determined in approximately the same concentrations in which they can be expected in the sample.

In our example of drinking water determination using the METROSEP Anion Dual 2 IC-anion column a 20  $\mu L$  sample loop is used; this is filled with the following standard solution:

```
0.5 mg/L F^-
5 mg/L Cl^-
10 mg/L NO_3^-
10 mg/L SO_4^2^-
(as Na^+- or K^+ salts in dist. water)
```

Please note that all program displays refer to the condition in which the system **s-03.smt** is loaded for the first time. If you want to work through this tutorial at a later date and the system **s-03.smt** has been altered in the meantime then differences with respect to the program display and the parameter values may occur.

In the description of the calibration procedure it is assumed that the PC and 761 Compact IC are not in operation and that the system must first be conditioned again. If this is not the case (e.g. if you start the tutorial immediately after conditioning) then you can skip steps **1** to **6**.

#### 1 Switch on 761 Compact IC

⇒ Switch on 761 Compact IC with mains switch **19** on the rear of the instrument. After the instrument has been switched on the mains pilot lamp **9** lights up.

#### 2 Switch on PC

⇒ Switch on PC and start «761 Compact IC» program.

#### 3 Open and connect system "PrepMSM2.smt"

- ⇒ Start PC program «761 Compact IC», if it is not already running (see section 2.5.3).
- ⇒ Select File / Open / System in the main window. Select the system file PrepMSM2.smt in the folder Suppressed and click on <Open>.
- ⇒ Select Connect to workplace of the Control menu in the system window.

#### 4 Start system "PrepMSM2.smt"

⇒ Select Start determination of the Control menu in the system window. High-pressure pump and peristaltic pump are started, at the same time, a chromatogram window opens where the baseline is recorded continuously. The suppressor



module is switched to the next position every 20 min and conditioned in this way.

#### 5 Condition system "PrepMSM2.smt"

⇒ Rinse IC system with eluent until the desired stability of the baseline is achieved and the suppressor module is sufficiently conditioned (at least 1 h).

#### 6 Stop system "PrepMSM2.smt"

⇒ Select Stop determination of the Control menu in the system window. Data recording and time program are stopped, highpressure pump and peristaltic pump keep running on.

#### 7 Open and connect system "s-03.smt"

⇒ Select Change of the System menu in the system window. Select the system file s-03.smt suited for the METROSEP Anion Dual 2 column in the folder Suppressed and click on <Open>. The system PrepMSM2.smt is closed, the system s-03.smt is opened and automatically connected.

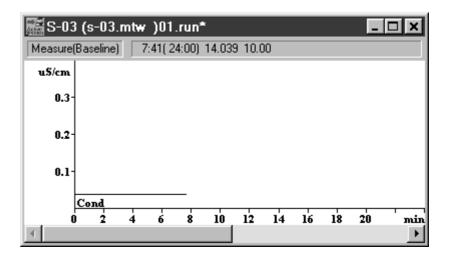
#### 8 Start system "s-03.smt"

- ⇒ Select Start determination of the Control menu in the system window. High-pressure pump and peristaltic pump are started, at the same time, a chromatogram window opens where the baseline is recorded continuously. The status bar of this window shows the message Measure(Baseline), beside this running time, analysis time, absolute conductivity and number of measuring points per second are displayed. The messages Running [761 Compact IC] and Waiting for INJECT [Suppressed] appear in the SYSTEM STATE window
- $\Rightarrow$  Let the system run for several minutes. If the system has already been conditioned then a stable conductivity value of approx. 14  $\mu$ S/cm will be obtained after a few minutes.
- ⇒ Double-click the chromatogram or select **View all** of the **View** menu. The sensitivity is set automatically so that all measuring points are visible.



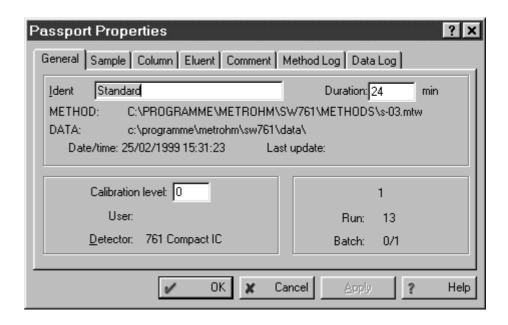
This function is only available if the integration has been started. In the present example the integration only starts after a delay period **Delay = 2.6 min**.

⇒ Select the desired sensitivity using the cursor keys <♠> or <♠>. In the chromatogram window the following display is seen, for example:



#### 9 Enter information for determination

⇒ Click on or select **Passport** of the **Method** menu. The passport opens with the **General** page, which contains the general information about the determination.



- ⇒ Enter a title for the calibration as **Ident** (e.g. **Standard**).
- ⇒ Change the analysis time under **Duration** if required.
- ⇒ Change other parameters on the passport tabs to characterize sample and acquirement conditions if required.

#### 10 Switch injection valve to "FILL" position

⇒ Click on the button of the 761 icon in the system window to switch the injection valve to the "FILL" position. At the same time, the suppressor module is switched to the next position.



#### 11 Fill sample loop

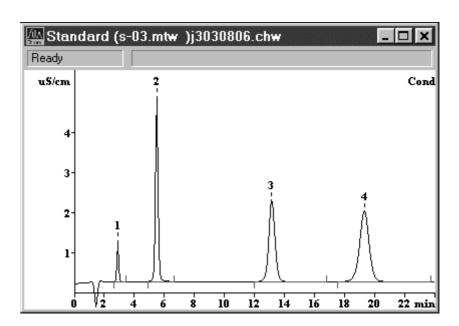
- ⇒ Immerse the aspirating tubing **4** attached to connection **3** in the standard solution.
- ⇒ Using the syringe fixed to connection **2** siphon in ca. 1 mL standard solution.

#### 12 Switch injection valve to "INJECT" position

- ⇒ Click on the Inject button of the 761 icon in the system window to switch the injection valve to the "INJECT" position. At the same time, the data recording is started automatically. The status bar of the chromatogram window shows the message Measure, the SYSTEM STATE window shows the messages Running [761 Compact IC] and INJECT done [Suppressed].
- ⇒ When the analysis time of 24 min set in the method has expired the recorded chromatogram is automatically evaluated and stored. The chromatogram window then closes.

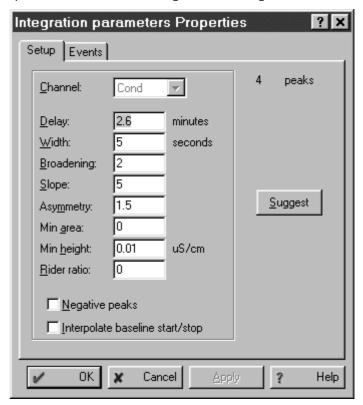
#### 13 Open chromatogram

⇒ Click on or File / Open / Chromatogram in the main window. Select the chromatogram file \*.chw just recorded and click on <OK>. The chromatogram window is opened where the found peaks are numbered and the baselines are drawn in.



#### 14 Modify integration parameters

- ⇒ Click on or select **Integration** from the **Method** menu in the main window to open the integration parameters window.
- ⇒ If required (e.g. if the fluoride peak is not detected), change the time delay **Delay** before starting peak integration.

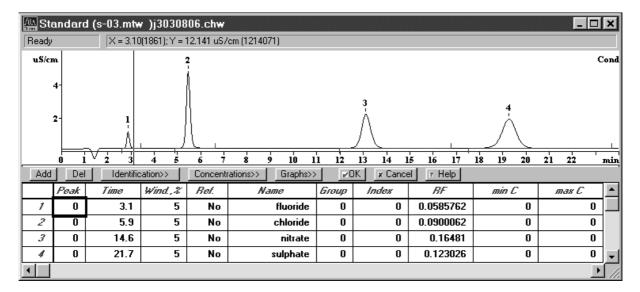


⇒ Click on <**Apply>**. The **Integration parameters** window is opened and the chromatogram is reintegrated.

- ⇒ Repeat this procedure for all other integration parameters until the result satisfies your expectations. Click on <Apply> for each parameter change.
- ⇒ Close the Integration parameters window with <OK>.

#### 15 Modify peak allocation

- ⇒ Click on or select Calibration / Components from the Method menu. The prepared table for the components fluoride, chloride, nitrate, and sulfate is displayed below the chromatogram. For all peaks which could be allocated unambiguously to a component this table contains the peak number in the Peak column and the retention time in the Time column. In the chromatogram itself this retention time is indicated by the cursor (vertical line).
- ⇒ If the **Peak** column contains a **0** the peak must be manually allocated to a component. In this case, insert the peak number from the chromatogram into the **Peak** column and click the **Time** field on this row with the mouse. The corresponding retention time is then inserted automatically as the new time for this component.
- ⇒ For each correctly allocated peak click on the retention time in the *Time* column. This time is automatically shown in the chromatogram by the cursor. If the retention time is not located in the area of the middle of the peak then the value in the *Time* column should be adapted accordingly.



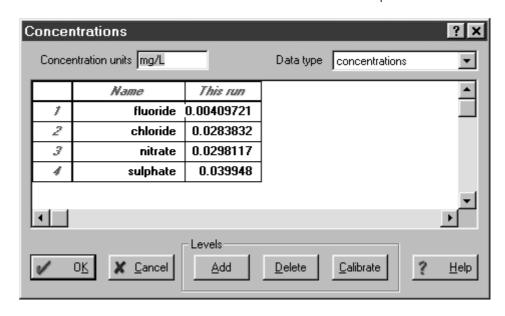
This is done by clicking on the chromatogram and moving the cursor with the aid of the cursor keys < and < to the middle of the peak and reading off the corresponding retention time in the status line. Enter this value (rounded off if necessary) into the *Time* column.

The optimized table could, for example, then appear as follows:

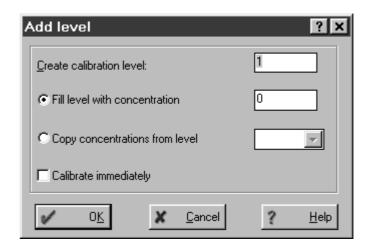
	Peak	Time	Wind, 2	Ref.	Name	Group	Index	RF	min C	так С
1	1	2.9	5	No	fluoride	0	0	0.0585762	0	0
2	2	5.5	5	No	chloride	0	0	0.0900062	0	0
3	3	13.1	5	No	nitrate	0	0	0.16481	0	0
4	4	19.3	5	No	sulphate	0	0	0.123026	0	0

#### 16 Start calibration

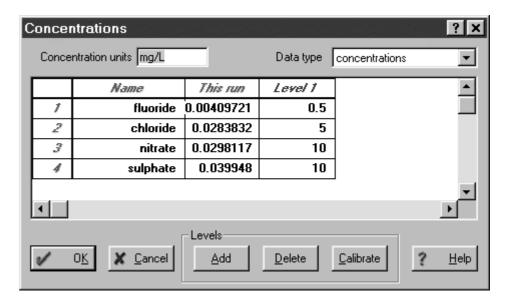
⇒ Click the **<Concentrations>** button in the component window.



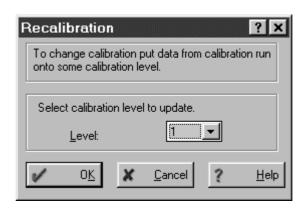
⇒ Click on <Add>. The Add level window appears.



- ⇒ Confirm Calibration level 1 in this window with <OK>. A new column Level 1 is added in the Concentrations window.
- ⇒ Enter the concentrations of all components of the standard solution used in this column:



⇒ Click on **<Calibrate>**. The following window appears:

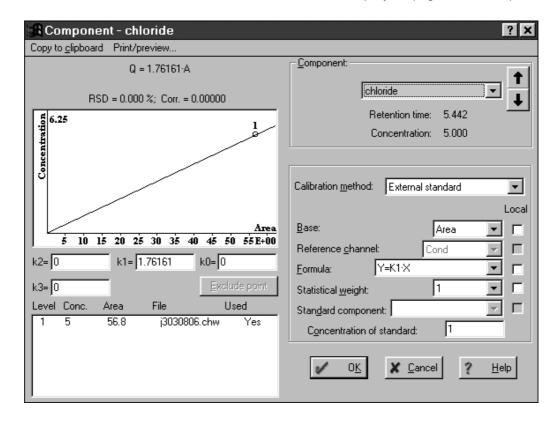


- $\Rightarrow$  Confirm Level 1 with <OK>.
- $\Rightarrow$  Click on **<OK>** in the **Concentrations** window.



#### 17 Display calibration curves

- ⇒ Click on **<Graphs>** in the **Components** window.
- ⇒ Select the desired component in the **Component** field for which the calibration curve should be displayed (e.g. for **chloride**).



- ⇒ Close the Component chloride window with <OK>.
- ⇒ Close the **Components** window below the chromatogram by clicking on **<OK>**.

#### 18 Save chromatogram and method

- ⇒ Close the chromatogram window. A window appears with the message Changes in \*.chw. Modified: Calibration. Save changes?.
- ⇒ Click on <Yes>. A window appears with the message \*.chw already exists. Overwrite?.
- ⇒ Click on <Yes>. A window appears with the message Method s-03.mtw was modified. Save changes?.
- ⇒ Click on <Yes>. The File Save As window appears.
- ⇒ Save the method under the previously given name **s-03.mtw**.



If you want to store the method under a different name then this must be newly selected for the system under PC icon / Setup / Processing method.

# 3.4 Sample determination

Following the calibration of the IC system as described in section 3.3 the first sample solution can be injected.

### 1 Enable "Verify sample"

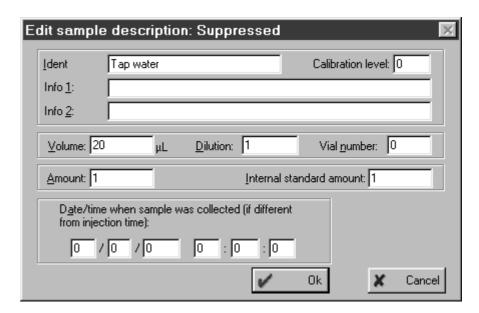
⇒ Switch on the **Verify sample** option of the **Control** menu in the system window. This causes the **Edit sample description** window to open at the start of each determination; the most important data for the sample can then be entered in this window.

### 2 Start system "s-03.smt"

⇒ Select Start determination of the Control menu in the system window. An empty chromatogram window opens where the baseline is recorded continuously. The messages Ready [761 Compact IC] and Initialisation [Suppressed] appear in the SYSTEM STATE window. Because of the Verify sample option switched on the Edit sample description window is opened automatically.

#### 3 Enter information for determination

⇒ Enter the desired information concerning the sample in the Edit sample description window and confirm with <OK>.



# 4 Switch injection valve to "FILL" position

⇒ Click on the button of the 761 icon in the system window to switch the injection valve to the "FILL" position. At the same time, the suppressor module is switched to the next position.

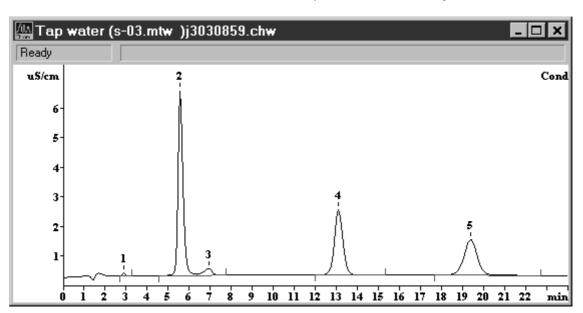


# 5 Fill sample loop

- ⇒ Immerse the aspirating tubing **4** attached to connection **3** in the vessel containing the drinking water sample.
- ⇒ Using the syringe fixed to connection **2** siphon in ca. 1 mL drinking water.

# 6 Switch injection valve to "INJECT" position

- ⇒ Click on the Inject button of the 761 icon in the system window to switch the injection valve to the "INJECT" position. At the same time, the data recording is started automatically. The status bar of the chromatogram window shows the message Measure, the SYSTEM STATE window shows the messages Running [761 Compact IC] and INJECT done [Suppressed].
- ⇒ When the analysis time of 24 min set in the method has expired data recording is automatically stopped and the chromatogram is integrated and evaluated. In the chromatogram window the peaks which have been found are numbered and the baseline drawn. In the SYSTEM STATE window the message Finished [Suppressed] appears. It is now possible to record further samples with the same system.



⇒ At the end of the determination, the chromatogram window is closed and the chromatogram is saved automatically. The file name is generated automatically and contains date and time in a coded form:

1<sup>st</sup> numeral: Alphabetical code for year

(e.g. j = 1999, k = 2000, l = 2001, etc.)

2<sup>nd</sup> numeral: Code for month

(1...9 = Jan....Sept., a...c = Oct....Dec.)

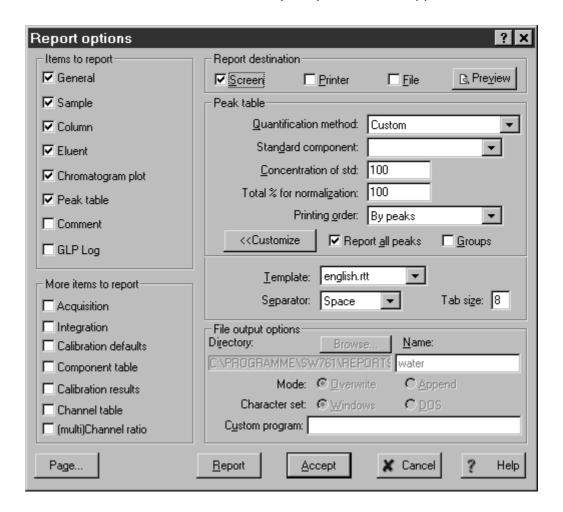
3<sup>rd</sup>+4<sup>th</sup> numeral: Day (01 ... 31) 5<sup>th</sup> - 8<sup>th</sup> numeral: Time (hh:mm)

# 7 Open chromatogram

⇒ Click on or File / Open / Chromatogram in the main window. Select the chromatogram file \*.chw just recorded and click on <OK>. The chromatogram window is opened where the found peaks are numbered and the baselines are drawn in.

# 8 Report output

⇒ Click on or select **Make report** of the **Process** menu in the main window. The **Report Options** window appears.



- ⇒ Under **Report destination**, switch on the desired target option for report output.
- ⇒ Under Items to report and More items to report, click all desired elements for the report output.
- ⇒ Click on <Report> to start the report output to the selected target.



# 9 Print report

⇒ Click on or select **Print** of the **File** menu in the main window. The standard printing window is opened where printer, printing range and number of copies can be selected. After confirmation with **<OK>** the results including the chromatogram are printed out.

# 4 Operation



This section describes the most important points concerning the operation of the 761 Compact IC. For further details please refer to the on-line help in the PC program which can provide you with the required information rapidly and conveniently from any place in the program.

# 4.1 Fundamentals of the operation

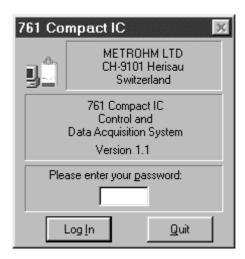
# 4.1.1 Starting/closing the program

### Start the «761 Compact IC» program



### Start the program

Double-click this icon or the **Metro761.exe** file to start the «761 Metro761.exe Compact IC 1.1» program. The Login window appears:



Enter your password and click on <Log In>.



After software installation, the program can be started without entering a **Password**. For the definition of users, see section 4.2.2.

## Close the «761 Compact IC» program

761 COMPACT IC / File / Exit

Exit the «761 Compact IC» program.

The program is also quit by clicking on in the upper right part of the main window **761 COMPACT IC**.



### 4.1.2 Glossary

#### **System**

The term "system" is used to describe the combination of **instrument settings**, **time program** and **process method** which have been optimized for the specific separating column and the determination to be carried out with it. A system is used to start single determinations or determinations with the help of a sample queue.

Systems are stored as **system files** (\*.smt) in the **Systems** directory.

#### Method

A method contains all information necessary for **data acquisition**, **integration**, **peak evaluation** and **quantification**. It can be considered as the chromatogram template, i.e. chromatogram without raw data.

Methods are stored as **method files** (\*.mtw) in the **Methods** directory.

Each system is linked to a method. This method is called **processing method** and is opened automatically at the start of a new determination.

#### **Chromatogram**

A chromatogram is a graphic plot of the elution curve (signal vs. time) recorded following a chromatographic separation on a separating column.

Chromatograms are stored as **chromatogram files** (\*.chw) in the **Data** directory. As well as the measuring data the chromatogram files also contain the method parameters and system settings which have been used for data recording, data processing and remote control.

#### **Determination**

In order to carry out a determination a suitable **system** must be selected for the separating problem. The result of the determination is a **chromatogram**, in which the measuring data and results of the determination are stored.

#### Calibration

Calibration is used to describe the method of determining the relationship between the peak height or peak area found for one component and its concentration in the sample. The result of the calibration is a **calibration function** (calibration curve), which shows the relationship between the amount of sample and the evaluated quantity.

The determination of the calibration function with reference solutions can be carried out as a **one-point** or as a **multiple-point calibration**. The calibration method which is mainly used in ion chromatography is the **external standard calibration** (absolute calibration); calibration with an **internal standard** (relative calibration) or **tabulated calibration** are also possible.

### **Integration**

Integration is to be understood as being the method for determining the peak area and peak height with the aid of approximate baselines. The integration algorithm included in the program is influenced by the **integration parameters** and the optionally programmable **integration events** which are defined in the method. In addition, the integration can be manually corrected later with the aid of the **peak editor**.

# Sample queue

A sample queue is used for the automated processing of series of samples, particularly in combination with a sample changer.

### **Batch reprocessing**

Batch reprocessing is understood to be the subsequent reprocessing of a series of chromatograms which have been loaded in a batch reprocessing queue. During reprocessing with a selected method the settings for calibration, integration, passport, appearance and report can be altered at will.

# 4.1.3 Overview of program windows

The «761 Compact IC» program consists of different windows whose functionality is linked together. The different windows are:

761 COMPACT IC	Main program	window for	file administration,
----------------	--------------	------------	----------------------

printing, opening of systems, methods and chromatograms, login and user rights, optional

settings and window handling.

**CHROMATOGRAM** Window for graphic plot of running or recorded

chromatograms.

**SYSTEM** Window for loaded system with possibility for

manual control of the 761 Compact IC.

**SYSTEM STATE** Window for display of status messages for the

connected system.

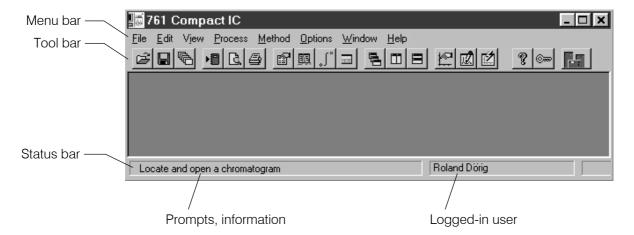
**WATCH WINDOW** Window for display of conductivity and pressure.

**QUEUE EDITOR** Window for edition of sample queues and batch

reprocessing queues.

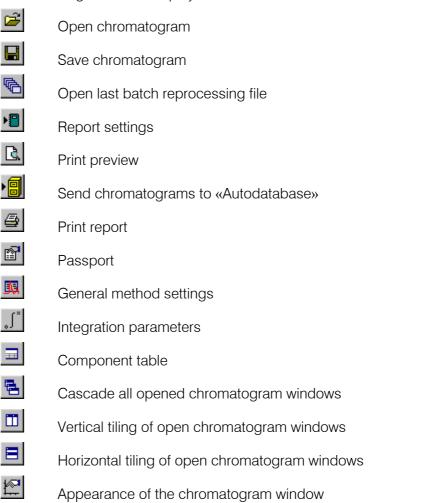
### 4.1.4 Main window elements

The elements of the main window **761 COMPACT IC** are the menu bar, the tool bar and the status bar, indicating prompts and logged-in user.



### 4.1.5 Icons of the main window

The following icons are displayed in the **761 COMPACT IC** main window:





Enable/disable peak editor mode



View whole chromatogram



Help



Lock system



Connected system

# 4.1.6 Overview of file types

The following file types are produced by the «761 Compact IC» software:

### \*.bar Batch reprocessing file

This binary file contains data of the batch reprocessing queue. The \*.bar file is stored automatically in the Data folder.

#### \*.cal Calibration file

This binary file contains calibration data, which can be exported with **761 COMPACT IC / Method / Calibration / Export calibration**. The **\*.cal** file is stored automatically in the **Methods** folder.

#### \*.chw Chromatogram file

This binary file contains chromatogram, system and method data of a determination. The \*.chw file is stored automatically in the Data folder.

#### \*.mtw Method

This binary file contains the data acquisition method, which can be linked to a system. The \*.mtw file is stored automatically in the Methods folder.

### \*.que Sample queue file

This binary file contains sample table queue. The \*.que file is stored automatically in the Methods folder.

#### \*.rtt Report template

This ASCII file contains a report template. The \*.rtt file is stored in the program folder.

#### \*.smt System file

This ASCII file contains the system settings. The \*.stm file is stored automatically in the Systems folder.

#### \*.dev Device file

This ASCII file contains drivers for devices. The \*.dev file is stored in the **Devices** folder.



#### 4.1.7 Context sensitive menus

Some of the menu functions of the program windows are also accessible by clicking on the desired window or item and pressing the **right mouse button**. The pop up windows have different contents and functions depending on the selected active window or item type.

# 4.1.8 Keyboard and mouse functions

The **mouse** can be used to carry out the normal program operating functions such as the selection of menu items and fields. It can additionally be used for magnifying a section of a chromatogram (**zooming**). To **zoom** a portion of the plot it is necessary to place the mouse cursor to the upper left corner of the square to zoom, press the left mouse button and drag the cursor to the lower right corner of the rectangle. After releasing of the left mouse button the selected region will be zoomed full-screen. If the cursor is active in the peak editor mode then it can be moved by pressing down the right-hand mouse key.

The **keyboard** can also be used to scale a chromatogram in the window, as described below.

### **Keyboard quick reference**

#### **Cursor** is inactive:

[ up ]	Increases sensitivity on the Y axis.
[down]	Reduces sensitivity on the Y axis.
[right]	Expands a chromatogram on the X axis.
[left]	Shrinks a chromatogram on the X axis.
[Ctrl] + [Home]	Autoscale procedure on the X axis (shows all on X).
[ Ctrl ] + [ End ]	Autoscale procedure on the Y axis (shows all on Y).
[ PageUp ]	Shifts a chromatogram on <sup>1</sup> / <sub>10</sub> part of a screen upwards.
[ PageDown ]	Shifts a chromatogram on <sup>1</sup> / <sub>10</sub> part of a screen downwards.
[Shift] + [up]	Increases a distance between channels of a chromatogram.
[Shift] + [down]	Reduces a distance between channels of a chromatogram.
[ 0 (Zero) ]	Adjusts a zero on the last point of a chromatogram (running chromatogram) or its lowest level (finished run).

#### Only part of the chromatogram is on screen:

**72** 



[Ctrl] + [right] Moves one window right (without change of

scale on X and Y axes).

[Ctrl] + [left] Moves one window left (without change of

scale on X and Y axes).

[ Home ] Shows the beginning of a chromatogram (with-

out change of scale on X and Y).

[ End ] Shows the end of a chromatogram (without

change of scale on X and Y).

[ 0 (Zero) ] Adjusts a zero on the lowest level in the win-

dow.

### **Cursor is active:**

[ 0 (Zero) ] Adjust a zero in site of the cursor.

[ right ] Moves cursor left to right.

[Shift] + [right] Quickly moves cursor left to right.

[ left ] Moves cursor to the left.

[Shift] + [left] Quickly moves cursor to the left.

[ Home ] Moves cursor to beginning of a window.

[ End ] Moves the cursor to end of a window.

[Shift] + [End] Sets the beginning of a window in site of the

cursor.

[Shift] + [Home] Sets the end of a window in site of the cursor.

# 4.1.9 Help

By clicking on , by clicking on Contents menu item, or by pressing the [F1] key you can get on-line help on the current topic anywhere in the program.

Green texts can be clicked to jump to a different Help topic.

Violet texts identify the dialog item, parameter or button in

the corresponding window.

Blue texts identify important information.



# 4.2 Instrument and software settings

#### 4.2.1 Fonts

#### 761 COMPACT IC / Options / Fonts

This option allows the selection of fonts used by the system.

**Font for dialog...** Selection of font used for dialog boxes.

Default setting: MS Sans Serif / Standard / 8 pt.

**Font for reports...** Selection of font used for report output to the

screen or printer.

Default setting: Courier New / Standard / 10 pt.

**Font for tables...** Selection of font used for data presentation in

tables on the screen.

Default setting: MS Sans Serif / Bold / 8 pt.

Font for plots... Selection of font used for labels on chroma-

togram plots and calibration curves.

Default setting: Times New Roman / Bold / 10 pt.

**Save fonts configuration** Save chosen font configuration.

# 4.2.2 Security system

The «761 Compact IC» program has a security system based on the list of users. Every user has his unique password and one of the following access levels:

**Novice** Restricted access to program functions. Allows

only start and stop of determinations using existing system and method files and manual control of the 761 Compact IC. Modifications of system, method and data files are not allowed.

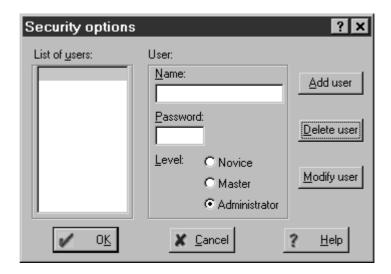
Master Access to all program functions with few excep-

tions: the user cannot set Global preferences,

Hardware settings and security system.

**Administrator** Access to all program functions.

It is recommended to make user lists and enter passwords as a first action after system installation. So, select the menu item **761 COMPACT IC** / **Options / Security** and click on **<Log In>** in the Log In window without entering a password. The **Security options** window appears:



Enter user name, password and access level for all users. Don't forget to make one of the user an **Administrator**, otherwise this window will not open again. At the end, click on **<OK>**.

After configuration of the security system the program prompts for the password every time the system starts. This user name stamps all methods, chromatograms and reports. It is possible at any time to change the user with the menu item **761 COMPACT IC / Options / Lock system**.

# 4.2.3 Global settings

761 COMPACT IC / Options / Global preferences

This window is used for **global program settings**.





This window is only accessible for users with Administrator access level.

**GLP On** 

If this option is enabled, the following parameters are automatically set:



Data file overwrite = Never
If method changed = Don't ask to

save

If disk method is newer = Ask to over-

write

**Data file overwrite** Overwriting of chromatogram files:

**Never** Chromatogram files cannot be overwritten. A

modified chromatogram is saved as a new file

with the file name number raised by 1.

Ask The user is asked if the chromatogram should

be overwritten.

Always Chromatogram files are always overwritten

without confirmation.

**If method changed** Saving of method files:

**Don't ask to save** The method file is not saved automatically. It

can be saved only with File / Save / Method.

Ask to save The user is asked if the method should be

saved.

Overwrite without asking

Method files are overwritten without confirma-

tion.

**If disk method is newer** Overwriting of method files:

**Ask to overwrite** The user is asked if the method should be

overwritten.

Overwrite without asking

Method files are overwritten without confirma-

tion.

**Opening chromatograms** Opening of chromatograms:

Ignore last data directory

If this option is enabled, the default data direc-

tory Data is opened.

OK, Apply buttons mean

Save object to file If this option is enabled, the <Apply> button in

system settings windows is replaced by the <Save> button. The system settings are saved

if the **<Save>** or **<OK>** button is clicked.

with file name confirmation

If this option is enabled, the <Apply> button in system settings windows is replaced by the <Save as> button. The system settings can be saved in a new file if the <Save as> or <OK>

button is clicked.

Chromatogram units Flow Pressure	Units for chromatograms: Unit for flow rate: µL/min, mL/min Unit for pressure: MPa, psi, bar, atm
Print via print spooler	Switch on/off printing via print spooler. Switch off this option if you use a GDI printer.
Measuring priority	Setting the priority of program execution. <b>real- time</b> means that the «761 Compact IC» program has the highest priority, <b>normal</b> means that all active programs have the same priority.
<default colors=""></default>	Default colors for chromatographic windows (details see section 4.5.3).

# 4.2.4 **COM port**

# 761 COMPACT IC / Options / 761 Compact IC:COM1

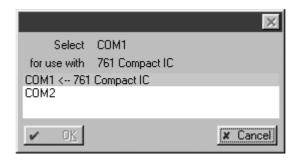
This menu item opens the **Links** window for COM port (serial RS232 interface) selection and settings.



If **COM1** is clicked with the right mouse button, the following menu items appear:

Put on desktop	Possibility for setting COM port parameters for recording data transfer (details see on-line help).
Change	Possibility for changing the COM port (default setting: <b>COM1</b> ). The following window is

setting: **COM1**). The following window is opened, where the interface can be changed by clicking on the desired COM port.



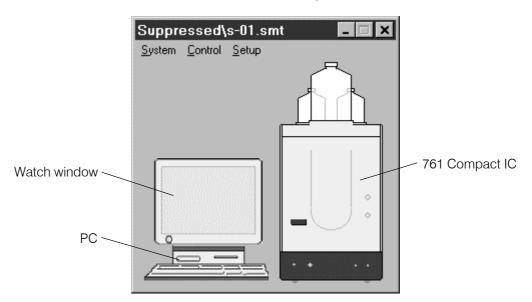


# 4.3 Systems

The term "System" describes the combination of **instrument settings**, **time program** and **process method** which has been optimized for a specific separating column and the determination to be carried out with it. A system is used to start single determinations or determinations with the help of a sample queue. Systems are stored as **system files** (\*.smt) in the **Systems** directory.

# 4.3.1 System window

A system window is opened with **761 COMPACT IC** / **File** / **Open** / **System** and the selection of the desired system file. It contains icons for **PC**, **Watch window** (screen) and **761 Compact IC**.



# 4.3.2 System file handling

The following menu items are used for opening, changing, saving and closing of systems:

#### 761 COMPACT IC / File / Open / System

Load an existing system file (\*.smt) from the Systems directory and open the corresponding SYSTEM window

The directory and name of the opened system file are displayed in the title bar of the **SYSTEM** window. A star (\*) at the end of the name indicates that the system settings have been changed since the last saving.

#### SYSTEM / System / Open other

Load an existing system file (\*.smt) from the Systems directory and open a new SYSTEM window. The system parameters can be changed for this system, but it can not be connected without disconnecting the actual system first.

#### SYSTEM / System / Change

Disconnect the current system, open the selected new system and connect it automatically.

#### SYSTEM / System / Save

Save the current system settings in a system file (\*.smt) in the Systems directory.

#### SYSTEM / System / Close

Disconnect the selected system (if it is connected) and close the system window.

# 4.3.3 System functions

#### **Connect and disconnect system**

To make possible manual control of the instrument and start of determinations the selected system must be connected to the PC. Only one system can be connected at the same time. Systems are connected and disconnected as follows:

#### SYSTEM / Control / Connect to workplace

Connect selected system to the COM port of the PC. If the system is connected, the two buttons <Inject> and <Fill> appear on the 761 icon in the system window for manual control of the injection valve (see section 4.3.7).

At the same time, the icon appears on the tool bar. If this icon is clicked, the **SYSTEM** window is always displayed in front of all other windows.

#### SYSTEM / Control / Disconnect from workplace

Disconnect selected system from the COM port of the PC. If the system is disconnected, the two buttons <Inject> and <Fill> disappear on the 761 icon in the system window. Manual control is not available for this system any more, but all other system settings can be modified and saved.

#### Start/stop hardware and record baseline

**SYSTEM / Control / Startup hardware (Measure Baseline)** 

**Starting the hardware** at the 761 Compact IC includes sending of **System startup values**, starting of the high-pressure pump and (if present) starting of the peristaltic pump.

At the same time, the recording of the measurement signal using the method of the connected system is started. Independently of the set chromatogram **Duration**, the measurement signal is recorded until the data acquisition is stopped with **SYSTEM** / **Control** / **Stop data acquisition** or a new determination is started. Alternatively the baseline recording can be stopped by clicking the icon of the chromatogram window. In this case the user is asked if the recorded baseline should be saved or not.



#### SYSTEM / Control / Shutdown hardware

High-pressure pump and (if present) peristaltic pump at the 761 Compact IC are immediately stopped. A running determination and an active sample queue are also stopped and the remote output lines are set to the values defined under **Hardware / Remote lines after power on**.

#### SYSTEM / Control / Stop data acquisition

Stop recording of the baseline.

### **Start/stop determinations**

#### SYSTEM / Control / Start determination

**Start determination** using the settings of the selected system. At this start command, the **System startup values** are set at the 761 Compact IC. The high-pressure pump and the peristaltic pump are started if they are not already running. The time program and the data recording are started either immediately (**Start with determination**) or after switching the injection valve to the "Inject" position (**Start with inject**) as set in the **Start mode** window.

#### SYSTEM / Control / Stop determination

**Stop running determination**. Data acquisition and time program are terminated immediately. The recorded chromatogram is saved automatically if the **Save chromatogram after the run** option on the **Passport** / **Processing** tab is enabled.

Alternatively the determination can be stopped by clicking the icon of the chromatogram window. In this case the user is asked always if the determination should be saved or not.

#### SYSTEM / Control / Stop data acquisition

**Stop data acquisition** of the running determination immediately and save the recorded chromatogram automatically if the **Save chromatogram after the run** option on the **Passport** / **Processing** tab is enabled. The time program of the running determination is continued normally.

### **Options for determinations**

#### SYSTEM / Control / Auto restart

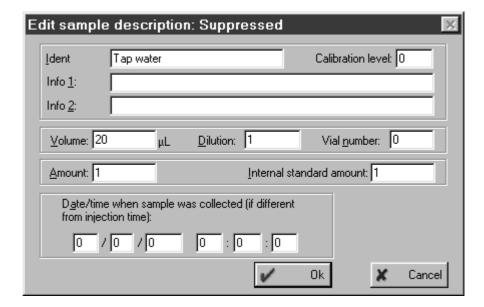
If this option is enabled, a new determination is started automatically using the current system after the preceding determination has been finished normally or stopped manually.

The **Auto restart** option is disabled if determinations are made with an active sample queue.

#### SYSTEM / Control / Verify sample

If this option is enabled, the **Edit sample description** window is opened automatically at the start of each determination for entry of the following sample information:





Ident	User defined identifier (title) for the chromatogram to be displayed in the title bar of the chromatogram window and in the <b>Chromatogram open</b> window.
Calibration level	Calibration level (0 = sample; $1n$ = calibration solutions).
Info 1 / Info 2	Sample description.
Volume	Injected volume in μL.
Dilution	Dilution of the sample.
Vial number	Autosampler vial position to take sample from.
Amount	Sample amount. If this value is different for the calibration run (c) and the sample run (s), the component concentrations of the sample are calculated as follows:
	$\mathbf{C_s} = \mathbf{C_c} ullet Amount_s  /  Amount_c$
Internal standard amount	Concentration of the internal standard component for relative concentration calculations.
Date/time when	Date and time of sample collection (the default values are equal to the date and time when the chromatogram starts).

The **Verify sample** option is disabled if determinations are made with an active sample queue.



# 4.3.4 System settings

#### **Modify system window**

SYSTEM / Setup / Drag icons

If this option is enabled, the system icons can be resized and moved in the **SYSTEM** window and the **SYSTEM** window itself can be resized.

To **resize** an icon or the window, move the cursor to the desired object until  $\ ^{}$  appears. Press the left mouse button and resize the object to the desired size.

To **move** an icon, move the cursor to the desired object until + appears. Press the left mouse button and move the object to the desired place.

## **Watch window display**

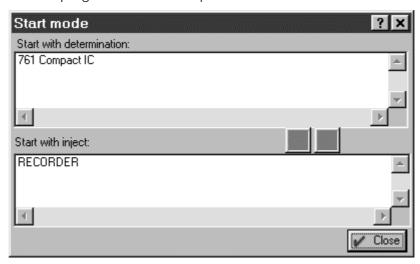
SYSTEM / Setup / Watch window

If this option is enabled, the **WATCH WINDOW** with conductivity and pressure display is automatically opened if a determination or measure baseline is started (see *section 4.3.6*).

#### Set start mode

#### SYSTEM / Setup / Start mode

This menu item opens the **Start mode** window for definition of start mode for time program and data acquisition.



This window contains the following two fields:

Start with determination The loaded object is started at the moment the

determination is started.

Start with inject The loaded object is started at the moment the

sample is injected.

The two objects available are:

**RECORDER** Data acquisition defined in the method.

**761 Compact IC** Time program defined on the Program tab of

the system control window.

These two objects can be moved from one field to the other using the

or buttons.

#### **Print system parameters**

#### SYSTEM / Setup / Parameters / Print

A report of the system parameters including the time program is created and opened using the «Wordpad» program. The \*.txt file opened can be printed, saved and exported into other programs. The system report includes the name of the method linked to the system, the configuration settings for the 761 Compact IC, the system startup values and the time program (incl. Remote configuration), if such a program exists and is switched on (ENABLED).

#### 4.3.5 PC icon

#### Menu options for PC icon



The PC icon is one of the three components of the **SYSTEM** window. If the system is connected and this icon is clicked with the right mouse button, the following menu appears:

Load the processing method linked to the system Open

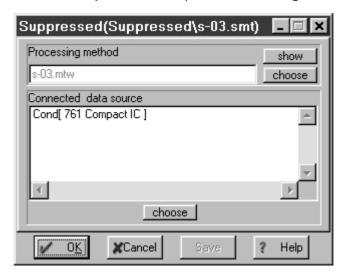
and open an empty chromatogram window.

Open PC setup window for selection of processing Setup

method and data source.

### Select processing method and data source

The **PC** / **Setup** menu item opens the following window:



This window contains the following two fields:

Directory and name of the method file (\*.mtw) Processing method

linked to the system.

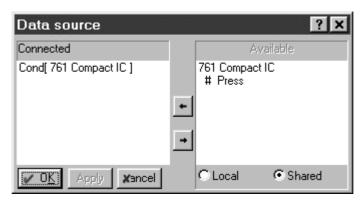
The method file can be selected with <choose>

and opened with <show>.



Connected data source

Data source for main measurement channel. By clicking on **<choose>**, the **Data source** window is opened for selection of the data source:



This window contains the following two fields:

**Connected** Connected data source.

Default setting: Cond[761 Compact IC]

Available Available data sources of the 761 Compact IC:

# Cond Conductivity
# Press Pressure

The data sources can be moved from one field to the other using the or buttons (only one data source can be connected).

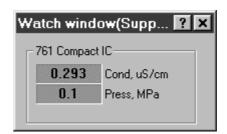
### 4.3.6 Watch window

#### Menu options for screen icon



The watch window icon is one of the three components of the **SYSTEM** window. If the system is connected and this icon is clicked with the right mouse button, the following menu item appears:

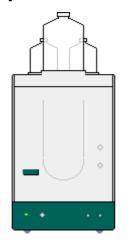
Open Open the **WATCH WINDOW** for live display of conductivity and pressure:



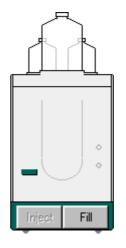
The colors of the watch window fields can be changed by clicking the fields with the right mouse button and selecting the menu item **Choose color / ...** .

### 4.3.7 Instrument icon

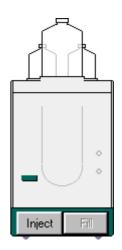
### Menu options for instrument icon



System disconnected



System connected Injection valve in "INJECT" position



System connected Injection valve in "FILL" position

The instrument icon for the 761 Compact IC is one of the three components of the **SYSTEM** window. If the system is connected, the 761 icon contains two buttons for manual control of the injection valve:

<Inject> Switch injection valve to "INJECT" position.
<Fill> Switch injection valve to "FILL" position.

If the system is connected and the 761 icon is clicked with the right mouse button, the following menu appears:

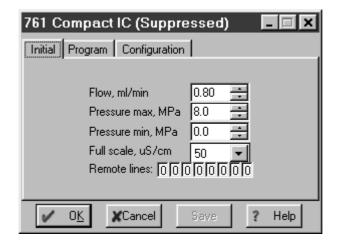
Open Open the system settings window.

Hardware Open the hardware settings window.

Diagnostics Open the diagnostics window.

#### System parameters for disconnected system

A double-click on the instrument icon or selection of the **Open** menu item using the right mouse button opens the system settings window. For a disconnected system, the **Initial** tab is displayed.





Flow, mL/min Startup value for flow rate of the high-pressure

pump.

Entry range: **0.20** ... **2.50** mL/min

Pressure max, MPa Startup value for maximum pressure limit for

high-pressure pump.

Entry range: **0.0** ... **25.0** MPa

Pressure min, MPa Startup value for minimum pressure limit for

high-pressure pump.

Entry range: 0.0 ... 25.0 MPa

Full scale, μS/cm Startup value for full scale range.

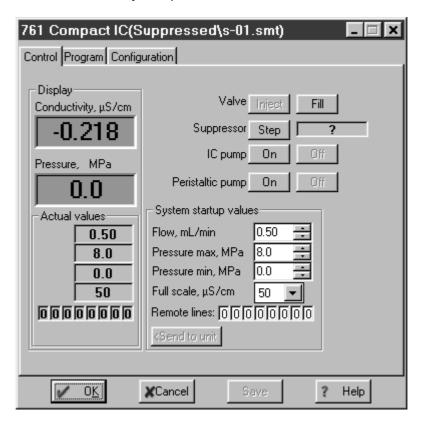
Selection: 50, 250, 1000  $\mu$ S/cm

**Remote lines** Startup value for remote line settings 1...8.

Selection: 0, 1

### Instrument control for connected system

A double-click on the instrument icon or selection of the **Open** menu item using the right mouse button opens the system settings window. For a connected system, the **Control** tab is displayed. It allows manual control of the 761 Compact IC functions and setting of startup values to be sent to the instrument. This tab shows the current measurement values for conductivity and pressure.



Conductivity, µS/cm Live display of measured conductivity.

**Pressure, MPa** Live display of measured pressure.

The color settings for this two fields can be changed by clicking the fields with the right



	mouse button and selecting the menu item $ \textbf{Choose color}  /  \dots  . $
Actual values	Display of actual values
Flow, mL/min	Display of flow rate of the high-pressure pump.
Pressure max, MPa	Display of maximum pressure limit for high- pressure pump.
Pressure min, MPa	Display of minimum pressure limit for high- pressure pump.
Full scale, μS/cm	Display of selected full scale range.
Remote lines	Display of current remote line settings.
Valve	Injection valve
<inject></inject>	Switch injection valve to "INJECT" position.
<fill></fill>	Switch injection valve to "FILL" position
Suppressor	Suppressor module
<step></step>	Switch the suppressor module to the next posi-
	tion. The time since the last switching of the suppressor module is displayed in the field be-
	side the <b><step></step></b> button.
10	High process as purpos
IC pump	High-pressure pump
<0n> <0ff>	Start high-pressure pump.
<uii></uii>	Stop high-pressure pump.
Peristaltic pump	Peristaltic pump
<0n>	Start peristaltic pump.
<off></off>	Stop peristaltic pump.
System startup values	The system startup values are sent and applied to the 761 Compact IC each time the system is connected, a determination is started, or the values are sent manually with <b><send to="" unit=""></send></b> .
Flow, mL/min	Startup value for flow rate of the high-pressure pump. Entry range: 0.20 2.50 mL/min
Pressure max, MPa	Startup value for maximum pressure limit for high-pressure pump. This limit is controlled even without connection to the PC. Entry range: 0.0 25.0 MPa
Pressure min, MPa	Startup value for minimum pressure limit for high-pressure pump. This limit is controlled even without connection to the PC. Entry range: 0.0 25.0 MPa
Full scale, μS/cm	Startup value for full scale range. Selection: <b>50</b> , <b>250</b> , <b>1000</b> μ <b>S/cm</b>
Remote lines	Startup value for remote line settings 18.

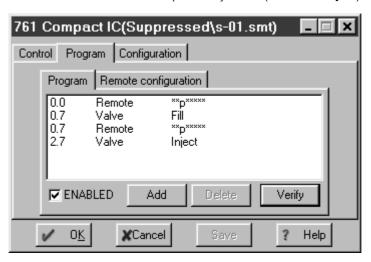
761 Compact IC 87

Selection: 0, 1



### **Time program**

On the **Program** tab of the system settings window a user-defined time program for instrument control can be entered. This program is started automatically as defined in the **Start mode** window (see section 4.3.3) either at the moment the determination is started (**Start with determination**) or at the moment the sample is injected (**Start with inject**).



The Program tab contains the two following subpages:

ProgramMain time program with all program steps.Remote configurationPossibility for creation of user-defined remote

commands.

### **Program**

On the **Program** subpage, program steps including time, program instruction and parameter can be entered.

applied.
truction is ion with
am in-
ctions, be en- juration
ee List of
gram is
es are dis-



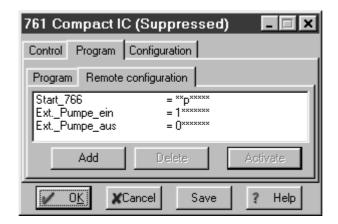
# List of program instructions

The following program instructions can be added to the time program on the **Program** subpage:

Instruction	Parameter entry	Meaning
Valve	Inject, Fill	Switch <b>injection valve</b> to "INJECT" or "FILL" position.
FullScale	50, 250, 1000 μS/cm	Set <b>full scale range</b> to the selected value.
ICPump	on, off	Switch on or off the high-pressure pump.
Flow	0.2 2.5 mL/min	Set <b>flow rate</b> of the high-pressure pump to the desired value.
Pmax	0.0 25.0 MPa	Set <b>maximum pressure limit</b> for the high-pressure pump to the desired value.
Pmin	0.0 25.0 MPa	Set <b>minimum pressure limit</b> for the high-pressure pump to the desired value.
Remote	0, 1, *, p	Set <b>remote output lines</b> 18 to the desired values. For entry of the first value, enter 1, 0, p or *. For entry of the other values, move the cursor in front of the value to be changed and enter 1, 0, p or *.
Program	END, RESET	The <b>END</b> flag can be used to end a program, especially if the program time should be longer than the chromatogram duration. Additional steps after this flag are not allowed. The <b>RESET</b> flag is used to reset the parameters to the system startup values.
Suppressor		Switch <b>suppressor module</b> to the next position.
Peristaltic	on, off	Switch on or off the peristaltic pump.

# **Remote configuration**

On the **Remote configuration** subtab user-defined remote commands can be defined, which can be inserted into a time program.



Name (1st column)

User-definable name of the remote command (e.g. Start\_766).

#### Remote command (2<sup>nd</sup> column)

Setting the remote output lines 1...8.

Selection: 0 (line off, inactive, open)

(line on, active, 0 V)

(pulse) р

(leave line in current status)

For entry of the first value, enter 1, 0, p or \*. For entry of the other values, move the cursor in front of the value to be changed and enter 1, 0,

**p** or \*.

<Add> Add new remote command.

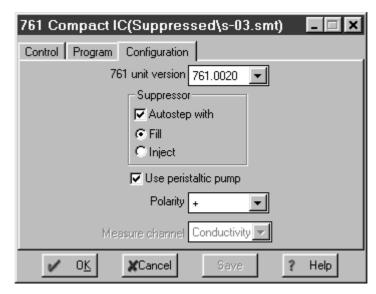
Delete selected remote command. <Delete>

<Activate> Activate the defined remote commands for in-

sertion into the time program.

#### Configuration

The Configuration tab in the system settings window contains configuration settings for the 761 Compact IC.



761 unit version

Selection of instrument version:

761 Compact IC without suppressor 761.0010 761 Compact IC with suppressor

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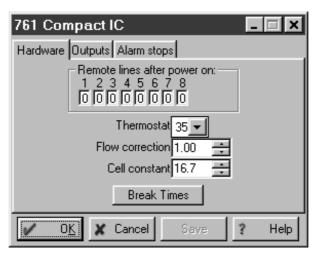
Suppressor	Suppressor module:
Autostep with	Automatic switching to the next position each time the injection valve is switched either to the <b>Fill</b> or to the <b>Inject</b> position.
Use peristaltic pump	If this option is disabled, the peristaltic pump is not switched on automatically at the start of a determination or with <b>Startup hardware</b> .
Polarity	Selection of the polarity of the output signal:  + positive polarity (for anions)  - negative polarity (for cations)
Measure channel	Display of the data source selected in the <b>Data</b> source window (see section 4.3.5).

### **Hardware settings**

Selection of the **Hardware** menu item using the right mouse button opens the hardware settings window consisting of the three tabs **Hardware**, **Outputs** and **Alarm stops**.

#### **Hardware**

This tab of the hardware settings window defines general parameters which are set automatically at power on of the instrument.



#### Remote lines after power on

The remote output lines 1...8 are set to this values after power on or a manual stop with **Shutdown hardware**.

Selection: 0, 1

**Thermostat** Operating temperature of the conductivity cell.

Selection: 25, 30, 35, 40, 45 °C, off



Thermostatting functions only if the ambient temperature is at least 5 °C lower than the operating temperature. It normally takes 30...60 min after power on until a temperature stability of  $\pm 0.01$  °C is attained.

#### Flow correction

Factor for correction of the difference between displayed and actual flow rate of the high-pressure pump.

Range: 0.9 ... 1.09

The flow correction is determined by measurement of the actual flow rate with the aid of a measuring cylinder as follows:

Flow correction = Displayed flow rate

Measured flow rate

#### **Cell constant**

Cell constant of the conductivity cell for correct display of the absolute conductivity. Enter the cell constant printed on the detector block into this field.

Range: **0.1** ... **1000** /cm

For a precise determination of the cell constant, pump a calibration solution of known conductivity through the IC system, observe the displayed conductivity and change the cell constant until the correct conductivity value is displayed.

#### <Break times>

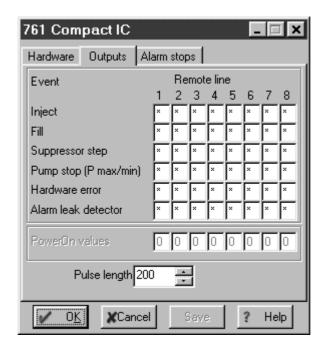
Possibility for changing the break times for injection valve **Valve** and suppressor module **Suppressor**.



These values should only be changed after consulting the Metrohm Service.

#### **Outputs**

The **Outputs** tab of the hardware settings window defines remote output signals to be set automatically if specific events occur.



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Event	Events for automatic remote signal output:
Inject	Switching of the injection valve to the "INJECT" position.
Fill	Switching of the injection valve to the "FILL" position.
Suppressor step	Switching of the suppressor module to the next position.
Pump stop (P max/mi	n)
	Pump stopped because pressure limits are exceeded.
Hardware error	Hardware error detected at the 761 Compact IC (high-pressure pump, injection valve, or suppressor not working correctly).
Alarm leak detector	Leak detector has detected solvent in the instruments interior.
Remote line	Set remote output lines 18.  Selection:   o (line off, inactive, open)  f (line on, active, 0 V)  p (pulse output)  * (leave line in current status)
PowerOn values	Display of the power on startup values for remote output lines set on the <b>Hardware</b> tab.
Pulse length	Length of a pulse in ms.

### **Alarm stops**

The **Alarm stops** tab of the hardware settings window defines the events for which the 761 Compact IC is stopped immediately. At an alarm stop, high-pressure pump and peristaltic pump are stopped immediately, the running determination and the active sample queue are also stopped.



**Event** Events for alarm stop:

Pump stop Pump stopped because pressure limits are ex-

ceeded.

Hardware error detected at the 761 Compact IC

(high-pressure pump, injection valve, or sup-

pressor not working correctly).



Leak detector has detected solvent in the in-

struments interior. This information is also stored in the instrument itself, so that it is stopped automatically even without connection

to the PC.

**Power on** Temporary power failure at the 761 Compact IC

hardware.

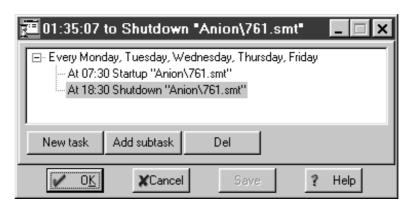
#### 4.3.8 Timer

the timer can be used to program system tasks which are started automatically daily or once at the desired time.



#### Timer icon

Clicking this icon opens the **Timer** window with the following options:



<new task=""></new>	Add a new task to the timer program.
<daily></daily>	Selecting this option opens a subwindow for selection of the days at which the task should be started.
<once></once>	Selecting this option opens a subwindow for selection of time and day at which the task should be started. After the selection of the desired program instruction the system file for which this instruction should be applied must be selected in the <b>Systems</b> folder.
<add subtask=""></add>	Add a new subtask for the selected daily task. The <b>time</b> and program instruction for this subtask has to be entered in the opened subwindow and the system file to which this instruction should be applied must be selected in the <b>Systems</b> folder.
<del></del>	Delete the selected task from the timer pro-

The following **program instructions** can be used for user-defined **event program tasks** in the **Timer program** window:

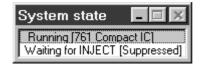
gram.

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Open	Open the selected system.
Close	Close the selected system and disconnect it from the workplace.
Startup	Startup hardware of the selected system.
Start	Start determination using the selected system.
Stop	Stop determination using the selected system.
Shutdown	Shutdown hardware of the selected system.

# 4.3.9 System state window



The **SYSTEM STATE** window is automatically opened if a system is connected. It shows status and error messages for this system. Messages concerning the 761 Compact IC hardware are followed by [761 Compact IC], messages concerning the loaded system are followed by ["folder name"] (name of the folder that contains the system file).

### **Status messages**

Checking on-line	Checking connection between PC and 761 Compact IC.
On-line	Connection between PC and 761 Compact IC OK.
UploadStartupValues	Hardware or system startup values have been loaded to the 761 Compact IC.
Initialization	Hardware or system initialization.
Ready	System is ready for starting a new determination.
Starting	Starting program or chromatogram data acquisition.
Running	Running program or chromatogram data acquisition.
Running program (xxx min left)	Running time program (in brackets: time left).
Waiting for INJECT	Waiting for "INJECT" to start program and/or chromatogram data acquisition.
INJECT done	Injection valve has been switched to the "INJECT" position.
Finished	Determination has been finished.
SHUTDOWN	761 Compact IC hardware is shutdown.



# **Error messages**

Detection of hard- ware failed	Bad connection between PC and 761 Compact IC or instrument switched off (check connecting cable or switch on instrument).
LEAK DETECTED	The leak detector has detected a leak (check IC system and connections).
E1	Program checksum wrong (call Metrohm service).
E2	RAM faulty (call Metrohm service).
E200	Invalid instrument adjustment (call Metrohm service).
E237	Storage of configuration values failed (repeat last action; if error reappears, call Metrohm service).
E238	Storage of instrument number failed (repeat last action; if error reappears, call Metrohm service).
E240	EEPROM faulty (call Metrohm service).
E258	Storage of setup values failed (repeat last action; if error reappears, call Metrohm service).
E295	Storage of memory values failed (repeat last action; if error reappears, call Metrohm service).
E296	Instrument stopped (restart instrument; if error reappears, call Metrohm service).
E297	Storage of remote line values failed (repeat last action; if error reappears, call Metrohm service).
E298	Storage of flow correction value failed (repeat last action; if error reappears, call Metrohm service).
E299	Storage of break time values failed (repeat last action; if error reappears, call Metrohm service).
E300	High-pressure pump faulty (restart pump; if error reappears, call Metrohm service).
E301	Injection valve blocked (check injection valve; if error reappears, call Metrohm service).
E302	Suppressor module blocked (check suppressor; if error reappears, call Metrohm service).
E303	Storage of maintenance information failed (repeat last action; if error reappears, call Metrohm service).

# 4.4 Methods

A method contains all information necessary for **data acquisition**, **integration**, **peak evaluation** and **quantification**. It can be considered as the chromatogram template, i.e. chromatogram without raw data.

Methods are stored as **method files** (\*.mtw) in the Methods directory.

Each system is linked to a method. This method is called **processing method** and is opened automatically at the start of a new determination.

# 4.4.1 Method file handling

The following menu items are used for opening and saving of methods:

### 761 COMPACT IC / File / Open / Method

Load an existing method file (\*.mtw) from the Methods directory and open an empty chromatogram window.

The name of the method is displayed in the title bar of the **SYSTEM** window. A star (\*) at the end of the name indicates that the method has been changed since the last saving.

#### 761 COMPACT IC / File / Save / Method

Save the method of the current chromatogram in a method file (\*.mtw) in the Methods directory.

## 4.4.2 Passport



#### 761 COMPACT IC / Method / Passport

This menu item opens the **Passport** window which includes a detailed textual description of the chromatographic run and consists of the following tabs:

General General information on determination.

Sample Sample information.

Column Column information.

Eluent Eluent information.

**Comment** User comments on chromatogram.

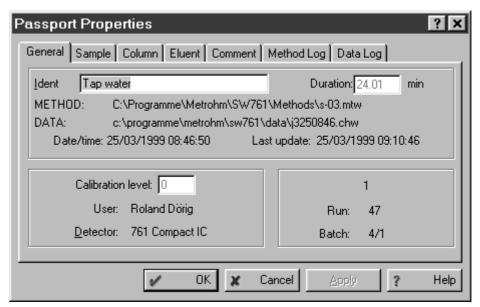
**Method Log** GLP log with date/time stamp for method.

**Data Log** GLP log with date/time stamp for chromatogram.



### General

Tab **General** of the passport with general description of the method and determination.



ldent	User defined identifier (title) for the chromatogram to be displayed in the title bar of the chromatogram window and in the <b>Chromatogram open</b> window.
Duration	Duration of the chromatogram in minutes. You can modify this value when the chromatogram is running.
METHOD	Path and file name of the <b>method</b> used for data acquisition (read-only).
DATA	Path and file name of the current <b>chromatogram</b> (read-only).
Date/time	Date and time of the chromatogram start (read-only).
Last update	Date and time of the last chromatogram modification and saving (read-only).
Calibration level	Calibration level. If the current run is used for calibration, the calibration level is a positive integer, otherwise it equals 0. You can modify this value when the chromatogram is running.
User	Name of the current user. It is taken from the list of users, in accordance with the password entered at the program starting.
Detector	Name of the detector. It can be entered with <b>Method setup</b> / <b>Measure</b> .
Run	Number of the current run starting from the very first one. All runs are automatically numbered by the system.



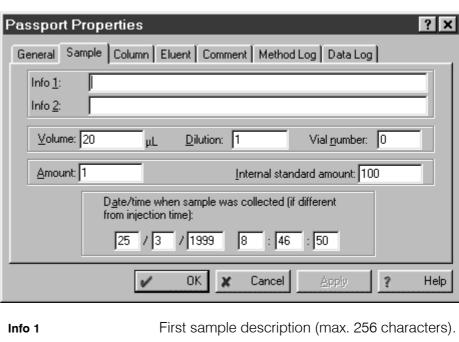
To reset the Run number press [Ctrl] [F8]. The Analysis number window is opened where the run number for the next run can be modified to the desired value.

**Batch** 

X/Y: total number of started sample queues / current analysis number in the sample queue (read-only).

# **Sample**

Tab **Sample** of the passport with sample information.



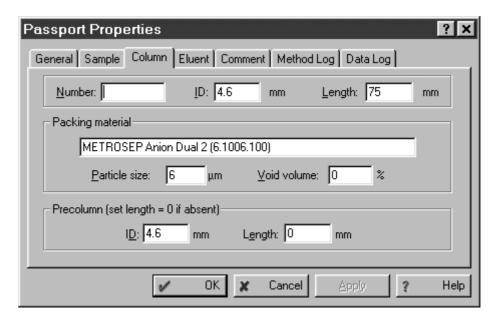
<b>✓</b>	OK Cancel Apply ? Help
Info 1 Info 2	First sample description (max. 256 characters). Second sample description (max. 256 characters).
Volume	Injected volume in μL.
Dilution	Dilution of the sample.
Vial number	Autosampler vial position to take sample from.
Amount	Sample amount. If this value is different for the calibration run (c) and the sample run (s), the component concentrations of the sample are calculated as follows:
	$\mathbf{C_s} = \mathbf{C_c} ullet Amount_s  /  Amount_c$
Internal standard amount	Concentration of the internal standard component for relative concentration calculations.
Date/time when	Date and time of sample collection (the default values are equal to the date and time when the

chromatogram starts).



# Column

Tab **Column** of passport with column information.



Number Serial number of column (max. 256 characters).

ID Internal diameter of the column in mm.

**Length** Length of the column in mm. This parameter is

used to calculate Linear flow.

**Packing material** 

**Column** Column description (max. 256 characters).

 $\mbox{ Particle size } \mbox{ Particle size of the column in } \mu \mbox{m. This value is } \label{eq:particle}$ 

used for the calculation of reduced theoretical

plate height Reduced TP height.

**Void volume** Void volume for the column in %. Used to cal-

culate Logarithmic index, Capacity factor and Linear flow. It is calculated by the system in accordance with the settings defined on Method setup

/ Math.

**Precolumn** 

Internal diameter of the precolumn in mm.

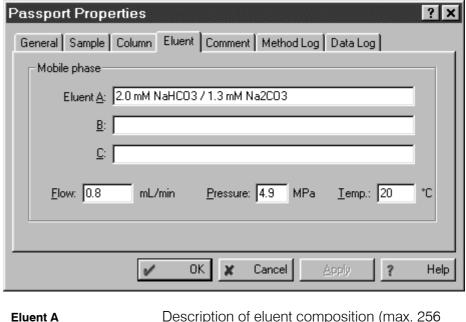
**Length** Length of the precolumn in mm (set length to 0

if no precolumn is used).



# **Eluent**

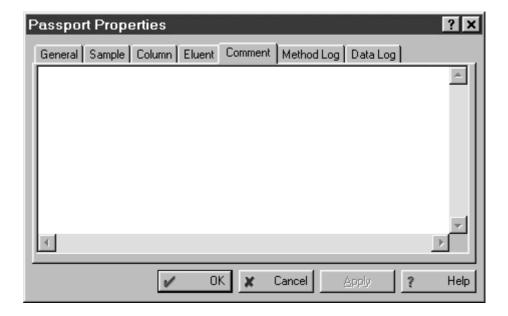
Tab **Eluent** of passport with eluent information.



Eluent A	Description of eluent composition (max. 256 characters).
Eluent B	Description of eluent composition (max. 256 characters).
Eluent C	Description of eluent composition (max. 256 characters).
Flow	Flow rate of the high-pressure pump in mL/min. The flow rate is used to recalculate the time axis into volumetric units. The system startup value for the flow rate is entered automatically into this field at the start of a determination with <b>Start with inject</b> .
Pressure	Pressure in MPa. The measured value for the pressure is entered automatically into this field at the start of a determination with <b>Start with inject</b> .
Temp.	Temperature in °C. Here you can enter the temperature of the column's thermostat or the ambient temperature.

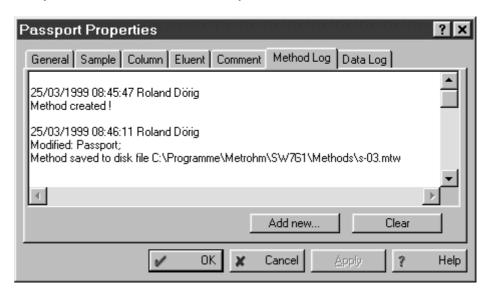
# **Comment**

Tab **Comment** of passport for entry of free-text user comments into the chromatogram description. Use this feature to enter any additional information about the chromatogram not included in other sections of the method.



# **Method Log**

Tab **Method Log** of passport with GLP Log for the method. The **Method Log** displays all automatically produced GLP messages concerning modifications made on a method. In addition to this messages free-text user comments can be entered. All this entries are stamped automatically with date and time of the entry.



<Add new> Add a new GLP comment to the Method Log.

Text can be entered in the **New comment** window, date/time and logged-in user are auto-

matically added.

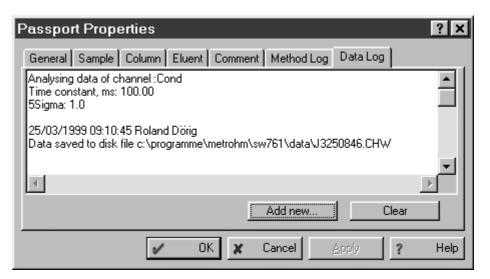
**<Clear>** Clear all GLP messages in the GLP Log and

add a new GLP comment. Text can be entered in the **New comment** window, date/time and logged-in user are automatically added.



# **Data Log**

Tab **Data Log** of passport with GLP Log for the recorded chromatogram. Here a GLP message is automatically produced for every modification made on a recorded chromatogram. In addition to this messages freetext user comments can be entered. All this entries are stamped automatically with date and time of the entry and cannot be cleared.



<Add new>

Add a new GLP comment to the Data Log. Text can be entered in the **New comment** window, date/time and logged-in user are automatically added.



# 4.4.3 Method setup



# 761 COMPACT IC / Method / Method setup

This menu item opens the **Method setup** window which includes the most common parameters for data acquisition and evaluation of the method and consists of the following tabs:

GeneralGeneral information on determination.MeasureImportant data acquisition parameters.

**Filters** Parameters for noise filtration.

**Processing** Set actions that are performed when the chroma-

togram finishes.

Math Parameters that are used for various types of cal-

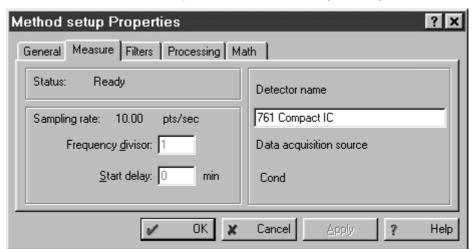
culations.

### **General**

Tab **General** of **Method setup** window with general description of the method and determination. This tab is identical to the **General** tab called with **Method** / **Passport** (see section 4.4.2).

### **Measure**

Tab Measure of Method setup window with data acquisition parameters.



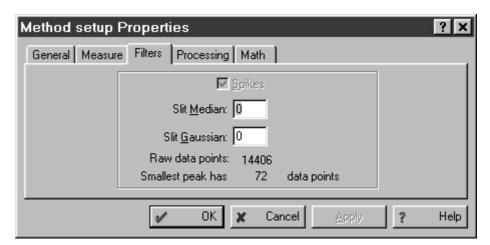
Status	Chromatogram measurement status (read-only).
Sampling rate	Sampling rate for recording the chromatogram (read-only; only displayed correctly for running or recorded chromatograms).
Frequency divisor	Divisor to reduce the sampling rate. Entry range: 19999
Start delay	Time delay before starting data acquisition in min.
Detector	Detector name.
Data acquisition source	Measuring channel (read-only).

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#### **Filters**

Tab **Filters** of **Method setup** window with parameters for noise filtration of raw data. Three different filtration algorithms are available to reduce noise and increase apparent signal-to-noise ratio.



**Spikes** 

**Spikes filter**. The Spikes filter smoothes the first and last points of the chromatogram and the points identified as spikes. The spike is exchanged with half of the sum of two neighboring signal values.

Slit Median

Median filter. Performs filtration if non-zero smoothing degree is entered. Smoothing degree should be a natural number. In the Median filter the values within the window are sorted by increasing response level and the response corresponding to the middle of the window is replaced with the value in the middle of a sorted array. This method affects chromatographic peaks in minimal extent, improves baseline and very effectively eliminates singlepoint spikes. In this case spike will be replaced with one of the neighboring points.

Slit Gaussian

Gauss filter. Performs filtration in non-zero smoothing degree is entered. In case of the Gauss filter the sum of all points within a window with Gauss weights distribution is calculated and used as a new raw data value. Peaks after smoothing become a bit smaller and wider, but their area does not change.

Raw data points

Number of data points per chromatogram (read-only).

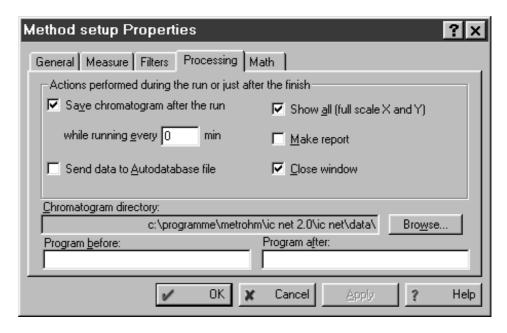
Smallest peak has ...

Number of data points per smallest peak on the chromatogram (read-only). This information is valuable to perform the **Compress** procedure (data compression).



# **Processing**

Tab **Processing** of **Method setup** window for definition of actions that are performed during the run or when the chromatogram finishes. Automatic post-run data processing is especially useful for chromatographic systems that include an autosampler.



#### Save chromatogram after the run

If checked, autosaving of the chromatogram will be performed on data acquisition ending.

#### while running every ... min

Save the chromatogram during the run for data safety at the time interval set (if **o** is set, the chromatogram is not saved during the run).

#### Send data to Autodatabase file

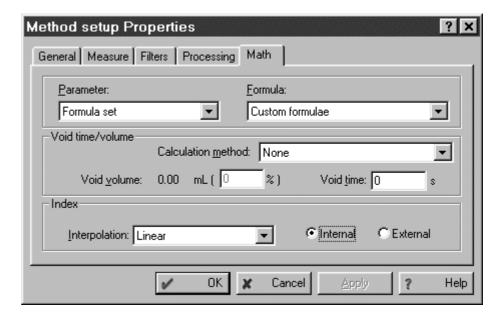
If checked, the chromatogram data are automatically sent to the Autodatabase file specified in the **Autodatabase options** window (see section 4.5.6) at the end of the determination If checked, scales on X and Y axes will be Show all automatically chosen so that the recorded chromatogram fits the window. If checked, report auto-printing will be per-Make report formed on data acquisition ending. If checked, the window is closed automatically Close window after the run (or sample queue) is finished. **Chromatogram directory** Directory where the current chromatogram is saved. Use the <Browse> button to select a new directory. **Program before** Name and path of the program to be started automatically before the run is started. Name and path of the program to be started **Program after** 

automatically after the run is finished.



#### Math

Tab **Math** of **Method setup** window with parameters that are used for various types of calculations.



Selection of calculation parameter: **Parameter** Formula setting by customer (Custom formulae) Formula set or according either to European Pharmacopoeia Or US Pharmacopoeia. Selection of calculation formula for number of Theoretical plates theoretical plates if Formula set = Custom formulae is set. Selection of calculation formula for resolution if Resolution Formula set = Custom formulae is set. Selection of calculation formula for asymmetry if **Asymmetry** Formula set = Custom formulae is set. Selection of calculation formula for the calcu-**Formula** lation parameter selected for Parameter. With Parameter = Formula set the following settings are possible: **Custom formulae** Formula setting by customer. European Pharmacopoeia Formula setting according to European Pharmacopoeia. **US Pharmacopoeia** Formula setting according to US Pharmacopoeia. Calculation of **void time/void volume**: Void time/volume **Calculation method** Selection of the method for void time calculation:

Manual entry in the **Void time** field.

The peak corresponding to the first component

is selected as a void time marker and its reten-

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None

1st component



tion time replaces the previous value of the void time. If the first component is not identified, the

expected retention time is used.

**1st peak** The first detected peak is used as a void time

marker for the run and its retention time is

stored in the **Void time** field.

From void volume %

For the calculation of the void time, the % value entered in the **Void volume** field is multiplied by the ratio of empty column volume and eluent

flow rate.

**Void volume** Void volume in mL and % of column volume.

The % value can be entered manually if Calcula-

tion method = From void volume % is set.

**Void time** Void time in s. This value can be entered

manually if Calculation method = None is set .

Index Calculation of **Retention index**:

**Interpolation** Use of linear or logarithmic interpolation scale

for retention index calculation.

Internal The index scale is constructed on the basis of

the current chromatogram. All components used for index scale calibration should be pre-

sent in the current sample.

**External** The index scale is constructed on the basis of

another standard chromatogram.

# 4.4.4 Integration



# 761 COMPACT IC / Method / Integration

This menu item opens the **Integration parameters** window which contains parameters and events for peak integration and consists of the following two tabs:

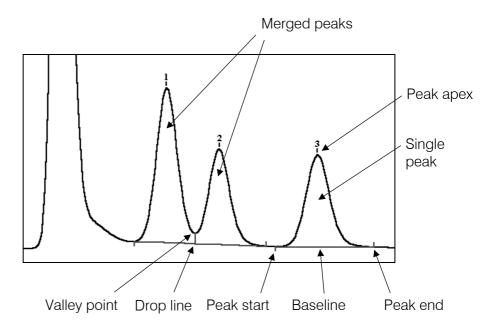
**Setup** Integration parameters.

**Events** Integration events.

The «Compact IC 761» software includes a built-in automatic integration algorithm to detect peaks on the chromatographic curve and to evaluate them using calculated baselines. The integration procedure is tuned by the integration parameters on the **Setup** tab and the integration events on the **Events** tab. Integration events have higher priority than integration parameters. The integration can be modified manually using the peak editor (see section 4.5.4).

The built-in integrator algorithm is based on the use of the first derivative (slope). In order to decide whether the slope at some point is significant, the first derivative value is divided by the baseline noise and the result is compared with a threshold value called "Slope". The threshold values for the upslope and downslope differ.

The baseline and the starting and finishing points are determined for each peak. For separated peaks the baseline consists of a straight line between the starting point and the finishing point. For neighboring peaks which overlap a common baseline is normally constructed; this connects the starting point of the first peak with the finishing point of the last peak. The peaks are separated from one another by dropping a perpendicular line from the lowest point (valley) between the peaks to the common baseline. Alternatively the baseline can also be drawn directly between the valleys of the peaks with the event **Enable valley-to-valley** (see integration events for baseline).



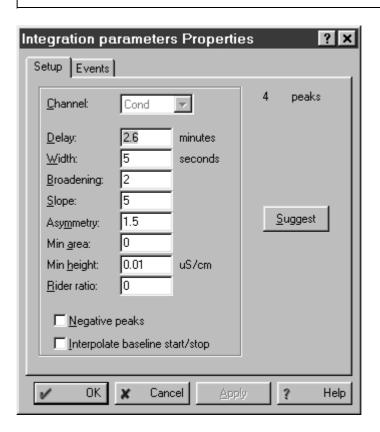


# **Setup**

Tab **Setup** of **Integration parameters** window with integration parameters. The two most important of these parameters for peak recognition are **Width** and **Slope**.



If the signal-to-noise ratio of the chromatogram is high enough (e.g. 100:1 or higher), the integration algorithm is not too sensitive to changes in integration parameters. On the contrary, if the baseline noise is high, careful tuning of these parameters may be needed.



Number of peaks	Number of peaks detected (read-only).
Channel	Data acquisition channel for peak detection (read-only)
Delay	Time delay before starting peak detection (in min). Entry range: 01440 min
Width	Peak width (at the baseline, in s). This parameter is used for setting baseline start and end points. If this value is too low, excessive narrow peaks are detected from the baseline noise. If it is too large, several adjacent peaks or baseline drift can be integrated as a single peak. For an optimal evaluation, it is recommended to enter the width of the narrowest peak on the chromatogram (normally 210 s). Entry range: 0.1480 s



#### **Broadening**

Broadening for peaks in the end of the chromatogram compared to peaks in the beginning.

This parameter is used for automatic adjust-

ment of Width.

Entry range: **0.1...100** 

# Slope

# Threshold for peak recognition.

The value of the first derivation (slope) of the curve is divided by the baseline noise (which is estimated using a special algorithm) and the result is compared with the **Slope** threshold value. Reasonable range of **Slope** parameter is 0.5...5.

Entry range: **0.1...400** 

#### **Asymmetry**

Ratio of slope threshold at the start of the peak to slope threshold at the end of the peak. This parameter is used for automatic adjustment of

Slope.

Entry range: 0.2...5

#### Min area

Minimum area of peaks to be detected.

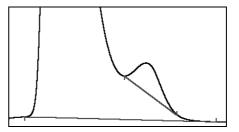
#### Min height

Minimum height of peaks to be detected.

#### Rider ratio

Ratio of peak heights for two adjacent peaks. If this threshold value is exceeded, the smaller peak is separated from the higher one by tangent skimming. This function is switched off al-

ways if 0 is entered. Entry range: **0...100** 



### **Negative peaks**

If this option is enabled, negative peaks are also detected.

# Interpolate baseline start/stop

If this option is enabled, the baseline start and stop points are interpolated. This setting is recommended for very sensitive determinations with high noise



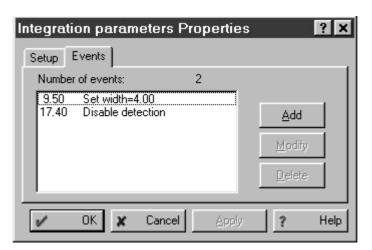
# <Suggest>

This function sets up reasonable integration parameters in the following way (assumes that several peaks are marked manually in the peak editor mode (see section 4.5.4) and wrong peaks deleted):

- Slope is set to 3.
- Asymmetry is set to 1.5.
- Width and Broadening are calculated by fitting a straight line through the measured peak halfwidth values vs. retention time. Width is determined as the y value of this line at the Delay time, Broadening is calculated as the y value at the end of the chromatogram divided by the y value at the Delay time. If this procedure does not provide adequate result,
- Width is set equal to the minimum value of peak width for the chromatogram,
- Broadening is set to 1.

#### **Events**

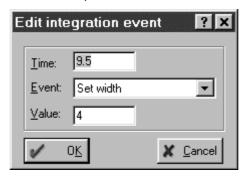
Tab **Events** of **Integration parameters** window for definition of integration events. Integration events have a higher priority than integration parameters and are used for fine tuning of the integration process. They should be used only in the case when problems cannot be solved by tuning a set of parameters from the integration parameters window.



Number of events	Number of integration events (read-only).
<add></add>	Add event to the list.
<modify></modify>	Edit the selected event.
<delete></delete>	Delete the selected event.
<apply></apply>	Accept integration parameters and perform reintegration.



If an integration event is added or modified, the **Edit integration event** window is opened where the following parameters can be set:



**Time** Start time for integration event in min.

**Event** List box with integration events to be selected

(see below).

**Value** Parameter value for selected integration event.

This field appears for events that demand input

of an additional parameter.

# Integration events for peak detection

**Disable detection** Set this mode to stop detection of new peaks. If

a peak is available at the moment of the event it is either finished (downslope peaks) or rejected

(upslope peaks).

**Enable detection** Clear **Disable detection** mode.

**Enable negative peaks** Enable detection of negative peaks. In some

cases this mode may result in instability of de-

tection algorithm.

Disable negative peaks Disable detection of negative peaks. This event

does not influence negative peaks which al-

ready started.

Disable peak reject Set this mode, if a small peak should not be re-

jected because of its flat apex.

**Enable peak reject** Clear **Disable peak reject** mode.

# Integration events for peak start/end

**Set peak start** Force the beginning of a new peak. If a peak is

available at the moment of the event it is either rejected (upslope) or terminated (downslope).

**Set peak end**Force peak end at the time of the event. Up-

slope peaks are rejected (except those born by **Set peak start** event), downslope peaks are ter-

minated.



Stop single peak mode Disable peak end detection. All peaks of any

group after this event will be treated as one peak. The group end will be treated as peak

end.

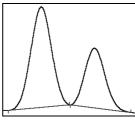
Set normal detection mode, when valley causes Start single peak mode

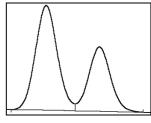
perpendicular drop or skim line separation.

Terminates the current peak and starts a new Split peak

one.

# Integration events for baseline





**Enable valley-to-valley** 

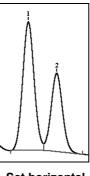
Disable valley-to-valley

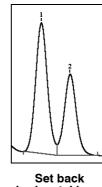
**Enable valley-to-valley** 

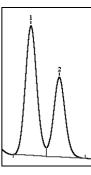
Disable perpendicular drop peak separation. All peaks are considered to be baseline-separated. The bottom of the valley becomes the baseline point.

Disable valley-to-valley

Enable perpendicular drop peak separation (default setting).







Set horizontal baseline

horizontal base

Set normal baseline

Set horizontal baseline

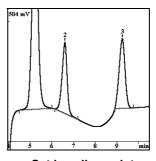
Set horizontal baseline for all peaks except the last one of adjacent peaks that are not separated. The baseline is drawn **forwards** from the peak start point.

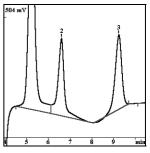
Set back horizontal base

Set horizontal baseline for all peaks except the first one of adjacent peaks that are not separated. The baseline is drawn **backwards** from the peak end point.

Set normal baseline

Set default baseline detection mode.



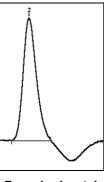


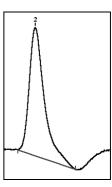
Set baseline point at 4.5, 6, 7.2, 8.9 and 9.8 min

without Set baseline point

# Set baseline point

Set defined baseline point for better evaluation of peaks on descending or ascending baselines. The baseline between two successive baseline points is set to zero for optimum integration.





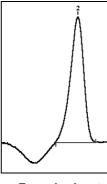
**Force horizontal** baseline

**Cancel horizontal** baseline

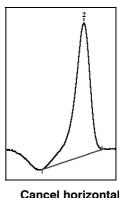
Force horizontal baseline Set horizontal baseline for a single peak. The baseline is drawn forwards from the peak start point. The intersection of the baseline with the signal is defined as peak end point.

# Cancel horizontal baseline

Clear Force horizontal baseline mode.



Force horizontal base back



**Cancel horizontal** base back

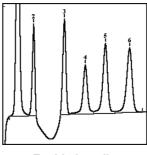


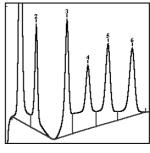
#### Force horizontal base back

Set horizontal baseline for a single peak. The baseline is drawn **backwards** from the peak end point. The intersection of the baseline with the signal is defined as peak start point.

#### Cancel horizontal base back

Clear Force horizontal base backmode.





Enable baseline penetration

Disable baseline penetration

# **Enable baseline penetration**

Allow crossing of the signal by the baseline.

#### Disable baseline penetration

Clear **Enable baseline penetration** mode. The baseline is aligned to the bottom value of the signal.

# Integration events for integration parameters

Set width Sets new Width parameter that supersedes de-

fault linear growth of expected peak.

Set slope Sets new Slope value.

Set min height Sets new Minimum peak height value.

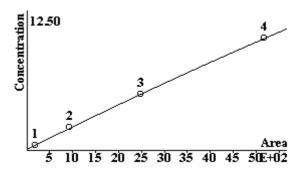
**Set rider ratio** Sets new value of **Rider ratio** parameter.

# 4.4.5 Calibration and quantification

#### **General information**

The aim of any chromatographic analysis is to answer the question "What components are present in the sample and what are their concentrations?". Two procedures are used to achieve this goal: the first step is called **calibration**, the second step includes **quantification**.

**Calibration** has two aims: to get retention characteristics for all components of interest (these data are stored in the component table) and to establish a relation between injected amounts and corresponding instrumental responses for all components of interest (stored in the concentration table). Calibration is performed by running one or several chromatograms of samples with known composition and known concentration of components (standards). For each calibrated component a **calibration curve** is constructed as a result.



With the 761 Compact IC three different procedures can be used for the construction of the calibration curve. By far the most important method for ion chromatography is the **external standard calibration** (absolute calibration) which is described in detail in this section. The other methods of **internal standard calibration** (relative calibration) and **tabulated calibration** (relative gradient factor, a modified method for external standard calibration) are of lesser importance and are not described in detail here (for details please refer to on-line help).

**Identification** is a procedure that enables to decide what peaks on the chromatogram correspond to what components. The identification is performed on the basis of the **Component table** created for calibration.

**Quantification** is a calculation procedure that determines components concentrations, on the basis of instrumental response (peak height or area), using the calibration curves obtained earlier for each component.

# **Notations**

R Stands for response value, either **area** or **height**, de-

pending on setting selected in the Calibration graphs

window.

V Sample Volume injected.

**D Dilution** coefficient, shows number of times to which

the initial solution is dissolved before injection.

V' = V / D Adjusted volume of injected sample. A correction is

made for the dilution coefficient.

C Concentration of the component in the initial solution

(before dilution).

 $Q = C \cdot V'$  Quantity of component, used for calibration curve

construction.

**Retention time**. Time needed by the mobile phase to

flow through the separation system.

 $\mathbf{t_0}$  Void time. Dead time needed by the mobile phase to

flow through the separation system.

 $t' = t - t_0$  Corrected retention time, called also net retention time.

L Column length.

 $v = L / t_0$  Linear Flow rate.

 $\mathbf{W}(\mathbf{R}) = \mathbf{k}_2 \mathbf{R}^2 + \mathbf{k}_1 \mathbf{R} + \mathbf{k}_0$ 

Calibration function (component quantity W vs. detector response R). In the case of the most common linear calibration curve  $Q = W(R) = k_1 R$  it comes through the origin. The concentration of the component in the analyzed mixture is calculated by the for-

mula C = W(R) / V'.

**RSD(Q, R)** Procedure, used for computation of regression coef-

ficients  $(\mathbf{k_0}, \, \mathbf{k_1} \, \text{and} \, \mathbf{k_2})$  of the calibration function  $\mathbf{W(R)}$  using RSD (Residual Standard Deviation). The procedure gets input as a set of calibration points (quantity  $\mathbf{Q}$  vs. response  $\mathbf{R}$ ) and outputs the calibration function  $\mathbf{W(R)}$  used for prediction of the component

quantity  $Q_i = W(R_i)$ .

Subscript values used:

j Stands for j-th calibration level run.

**s** Stands for standard component.

i Stands for component number.

### **External standard calibration**

External standard calibration (absolute calibration) is a key way of calibration. Basically this procedure calculates the dependence of injected component's quantity versus area (or height) of the corresponding peak. This dependence is showed as a calibration plot with injected quantity along the Y axis and peak area (or height) along the X axis. The injected quantity  $\mathbf{Q}_i$  is calculated as product of component's concentration  $\mathbf{C}_i$  to the reduced injected sample volume  $\mathbf{V}'$ :

$$Q_i = C_i \cdot V'$$

The calibration function  $W_i(R)$  for each component is calculated using the RSD method.

$$W_i(R) = RSD(Q_{ii}, R_{ii}) = RSD((C_i \cdot V'), R_{ii})$$

When quantification procedure is performed, a concentration  $C_i$  (absolute concentration of a component in the sample) is determined as a ratio of the computed quantity  $W_i(R_i)$  to the corrected volume of sample injected V':

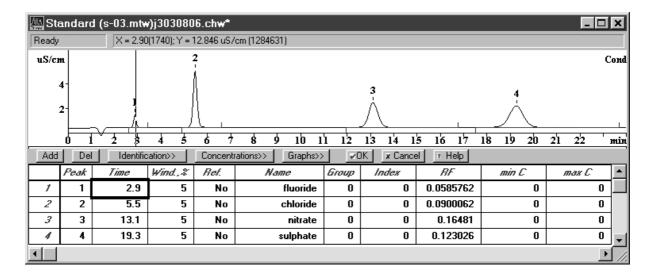
$$C_i = W_i(R_i) / V'$$

# **Component table**



# 761 COMPACT IC / Method / Calibration / Components

If the component table is opened, the chromatogram window is split into two parts. The upper one shows the chromatogram, the component table appears in the bottom part of the screen. When moving into the component table, a special cursor in the upper part of the window jumps to the peak corresponding to the current component.



The **Components table** stores the following information on the components to be analyzed:



Number	Row number (read-only).
Peak	Number of the peak in the chromatogram that corresponds to the given component. If this field contains a <b>o</b> , the <b>peak number</b> must be entered manually.
Time	<b>Expected retention time</b> of the component in the calibration run. After a new entry of the peak number the corresponding retention time is entered automatically as soon as this field is clicked on with the mouse.
Wind.%	Identification window of the component. This value is the maximum allowed difference of the actual retention time of the component and its expected retention time, measured as % of expected retention time. The component will be identified within its identification window only.
Ref.	Reference component. The entry Yes defines components as reference components; these ensure improved peak identification of the other Ordinary components (details see on-line help).
Name	Component's name (cannot include spaces). Rows with empty names are excluded from the ta- ble of components when you leave the component table editor.
Group	Number of the <b>group</b> to which the components are allocated. If group numbers ≥ 1 are entered here then a group report can be outputted for each group; this also contains the intermediate totals for this group. This group report is produced after the main report table.
Index	Retention index for components with known index. In the case of an unknown index this value should be equal to 0. For index calculation the user must define index values for at least two peaks, and all other values will be calculated by the software using linear or logarithmic approximation (details see on-line help).
RF	<b>Response factor</b> . This value corresponds to the slope of the calibration curve (coefficient $\mathbf{k_1}$ of the calibration formula).
min C (max C)	<b>Minimum (Maximum) concentration</b> value for the component. Components whose concentration leaves the range <b>min Cmax C</b> are marked in the peak table by the sign "!".



Additionally, the **Components table** contains the following buttons:

<add></add>	Add a new component (an empty column) to the component table.
<del></del>	Delete the current component from the component table.
<identification></identification>	Open the <b>Peak identification</b> window (see <i>Identification</i> ).
<concentrations></concentrations>	Open the <b>Concentration table</b> (see <i>Concentration table</i> ).
<graphs></graphs>	Open the component window with display of calibration curve and calibration parameters.
<0K>	Accept all changes and close the <b>Component table</b> .
<cancel></cancel>	Reject all changes and close the <b>Component table</b> .

The standard methods supplied with the «761 Compact IC» program all contain a component table with pre-defined components with entries for the names and retention times.



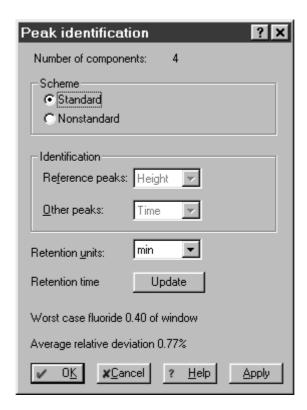
If the output of all peaks is required in the result report then a socalled universal component can be defined in the component table for this purpose; this receives the value **0** in the **Time** field and no peak is allocated to it. The name entered under **Name** (e.g. **unknown**) is then used for all peaks for which no component can be allocated.

# **Peak identification**

# 761 COMPACT IC / Method / Calibration / Identification

Selecting this menu item or clicking on <Identification> in the component table opens the **Peak identification** window with parameters for tuning the peak identification procedure.

Number of components	Number of components in the component table (read-only).
Scheme	Quick choice of identification parameters scheme.
Standard	Set default meanings of identification parameters: <b>Height</b> for reference components, <b>Time</b> for all other components (ordinary components).
Nonstandard	Set custom meanings of identification parameters.



Identification	Identification parameters.
Reference peaks	Identification parameter for reference components. Default is <b>Height</b> .
Other peaks	Identification parameter for ordinary components. Default is <b>Time</b> .
Retention units	Choice of retention units. Default is min.
Retention time	Retention time.
<update></update>	Updates expected retention times of components according to the current chromatogram.
Worst case	Information on the component with the worst (largest) deviation of actual and expected retention time. Deviation is given as a part of component's identification window.

# Average relative deviation

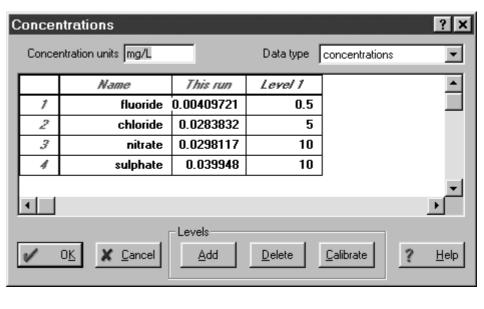
Relative deviation averaged for all components.



# **Concentration table**

# 761 COMPACT IC / Method / Calibration / Concentrations

Selecting this menu item or clicking on **<Concentrations>** in the component table opens the **Concentrations** window with the concentration table containing concentrations of all components for all **Calibration levels**. Each calibration level corresponds to a sample used for calibration and to a point on the calibration curve.

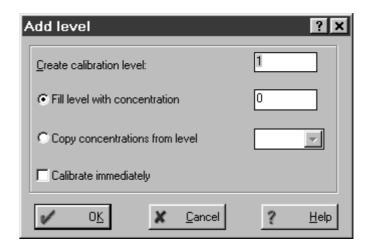


Concentration units	User defined units of concentration which will appear in the report. Nevertheless, entry of new concentration units does not cause recalculation of concentrations.
Data type	Choice of the data type that will be shown in the concentration table: <b>concentrations</b> (default setting), peak <b>heights</b> , peak <b>areas</b> , <b>volumes</b> .
Number	Number of the component (read-only).
Name	Name of the component taken from the component table (read-only).
This run	Contains concentrations (or other chosen values) obtained in the current run (read-only, except for universal component).
Level 1Level N	Contains concentrations (or other chosen values) of components for corresponding Calibration level.



#### <Add>

Add a new **Calibration level** to the concentration table. The following window appears:



#### Create calibration level

Number of calibration level to add.

#### Fill level with concentration

Concentration to be filled in for all components.

#### Copy concentrations from level

Number of calibration level from which concentrations are copied.

### Calibrate immediately

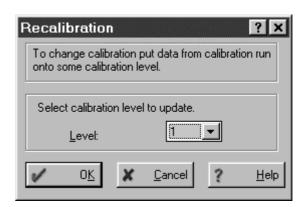
Recalibration with new calibration levels.

# <Delete>

Delete the current calibration level (where cursor is situated) from the concentration table.

#### <Calibrate>

Calibrate the current chromatogram with the selected calibration level. The following window appears:



Level

Calibration level to be used for recalibration.

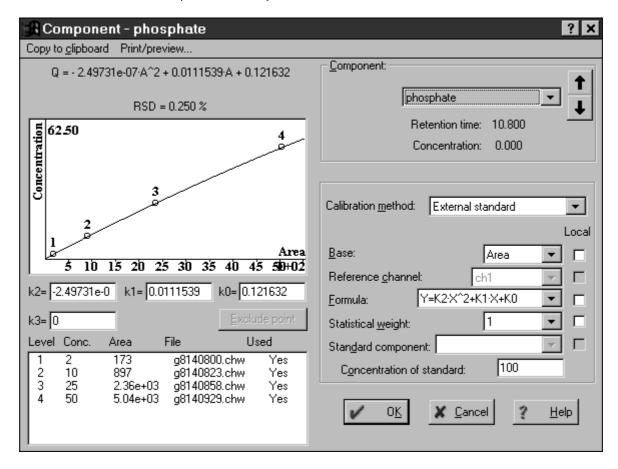
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# **Calibration curve**

# 761 COMPACT IC / Method / Calibration / Graphs

Selecting this menu item or clicking on **<Graphs>** in the component table opens the **Component** window.



In this window the results of the calibration are shown for each component together with the calibration curve. The parameters for calculating the calibration curve can also be entered here.

# **Calibration results and curve**

Analytical expression	A line above the calibration curve looking like $\mathbf{Q} = \mathbf{k_3} \cdot \mathbf{A}^3 + \mathbf{k_2} \cdot \mathbf{A}^2 + \mathbf{k_1} \cdot \mathbf{A} + \mathbf{k_0}$ . This formula is used for approximation of the calibration curve.
RSD	RSD (Residual Standard Deviation) value to evaluate the error of calibration curve approximation.
Corr.	Correlation coefficient value is available only in the case of linear calibrations without weighting of data points.
Calibration curve	Plot of measured calibration points and calculated calibration curve.
k0, k1, k2, k3	Calibration coefficients $\mathbf{k_0}$ , $\mathbf{k_1}$ , $\mathbf{k_2}$ and $\mathbf{k_3}$ (coefficients of the calibration formula).



# **Calibration points table**

This table contains the basic information used to construct the calibration curve:

Level Number of Calibration level.

**Conc.** Concentration of the current component in the

calibration sample. It is taken from the concen-

tration table.

**Area (or Height)** Peak height or area of the current component,

depending on calibration base.

File name of calibration chromatogram that

stores data on the given calibration level.

**Used** Information whether the calibration level is used

for calculation of the calibration curve or not.

**Exclude point>** Exclude the selected calibration point from the

list and recalculate calibration coefficients for the current component. Repeated pressing includes the point again. You can exclude points that drop out of the calibration curve watching

for RSD values.

# **Component information**

**Component** Allows to select the current component from the

list. It is possible also to scroll the list of components by mouse, using the special arrow but-

tons on the right.

**Retention time** Retention time of the selected component

(read-only).

**Concentration** Concentration of the selected component

(read-only).

### **Calibration parameters**

**Calibration method** Method that is used for calibration procedure.

There are three basic methods of calibration:

**External standard calibration** 

Absolute calibration. It is the basic calibration

procedure in ion chromatography.

Internal standard calibration

Relative calibration.

**Tabulated calibration** Relative response factor calibration. It is a sim-

plified method of external standard calibration.

**Local** If this checkbox is marked then the correspond-

ing parameter is valid for the current component only. Otherwise this parameter is global

(i.e. default for all other components).



Base	Base for calculations (Area or Height) in quantification and calibration procedures. Indicates,
	which parameter is to be used as a peak response.
Reference channel	Channel which is used to measure area or height (read-only).
Formula	Formula for calibration function. There are six possible calibration dependencies for linear and nonlinear calibration curves.
Statistical weight	Weighting parameter used to weight calibration points deviations on calculating coefficients of the calibration formula. Apart from the uniform weighting 1 the values 1/x, 1/x², 1/y and 1/y² are also possible.
Standard component	Name of the standard component in the case of Internal standard calibration or Tabulated calibration method (details see on-line help).

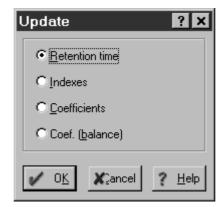
#### **Concentration of standard**

Concentration of the standard component in the case of **Internal standard calibration** or **Tabulated calibration** method (details see on-line help).

# **Update calibration**

# 761 COMPACT IC / Method / Calibration / Update

Selection of this menu item opens the **Update** window.



Retention time	Replace the expected retention times of the components with the retention times of the corresponding peaks obtained in the current run.
Indexes	Fill Index field for components with zero index.
Coefficients	Recalculate all calibration coefficients for all components (performs re-calibration).
Coef. (balance)	Recalculate coefficient of the universal component. The aim of this action is to adjust the re-



sponse factor of this component so, that the total amount of the injected components becomes equal to the declared value. This declared summary amount is entered into the **Concentration** field of the universal component for **This run** level.

#### Load and save calibration data

#### 761 COMPACT IC / Method / Calibration / Load from method

Load the calibration from the method. This option is used to update the calibration for the current run by the one taken from the method used for acquisition.

#### 761 COMPACT IC / Method / Calibration / Save to method

Save the current calibration to the method. This option is used to transfer the updated calibration from the run to the current method (i.e. the method associated with the current chromatogram window).

# Import and export calibration data

### 761 COMPACT IC / Method / Calibration / Import calibration

This option imports a calibration from a calibration file \*.cal to the current method or chromatogram. It is used to transfer the updated calibration from chromatogram-to-chromatogram or from method-to-method. Different methods can be processed in this case using the same calibration.

#### 761 COMPACT IC / Method / Calibration / Export calibration

This option exports a calibration to a in calibration file \*.cal. It is useful to transfer a calibration from chromatogram-to-chromatogram that were obtained by different methods.

#### **Put out calibration curves**

# **COMPONENT / Copy to clipboard**

This option allows to copy the calibration curve of the selected component to the clipboard so that it is available for other Windows applications, such as WinWord, Excel, etc.

### **COMPONENT / Print/Preview / Preview this**

Display calibration curve print preview of the selected component.

### **COMPONENT / Print/Preview / Preview all**

Display calibration curve print preview of all components.

#### **COMPONENT / Print/Preview / Print this**

Print selected calibration curve of the selected component.

# **COMPONENT / Print/Preview / Print all**

Print calibration curves of all components.

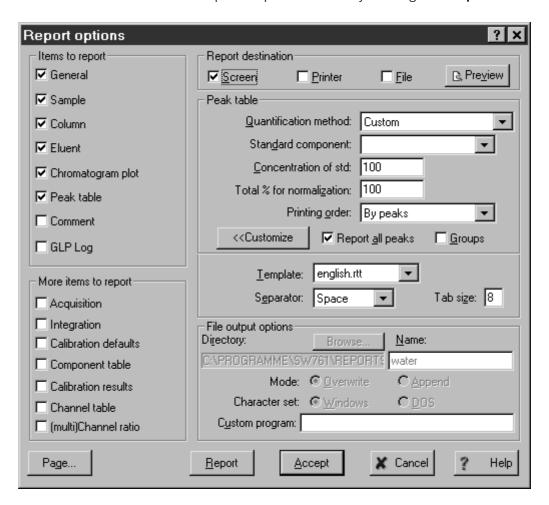
# 4.4.6 Report output

# **Report options**



761 COMPACT IC / Method / Report options 761 COMPACT IC / Process / Make report

The **Report options** window is divided into several regions that combine parameters for report output on their functionality and includes several different areas. The report output is started by clicking on **<Report>**.



# **Items to report**

This part of the **Report options** window contains a list of important report items that can be included into the report on the user's choice:

# General

General description on the analysis from the Passport. The **Report date** and the logged-in user **Printed by** are also put out.

Report date: 05/04/1999 12:34:00
Printed by: Roland Dörig

Ident: Tap water

Analysis from: 19/02/1999 11:06:41

File: j2191106.chw Last save: 23/02/1999 10:02:30

Modified!

Method: s-03.mtw Last save: 23/02/1999 09:57:48

Run operator: Urs Waldburger

Analysis number: 5



Sample Sample information from the Sample tab of the passport.

SAMPLE: Tap water from Application Laboratory

: Herisau, Switzerland

Vial number: 2 Volume: 20.0 µl Dilution: 1.00

**Column** Column information from the **Column** tab of the passport.

COLUMN: METROSEP Anion Dual 2 (6.1006.100)

Size: 4.6 x 75 mm Number: A106 Part.size: 6.0 µm

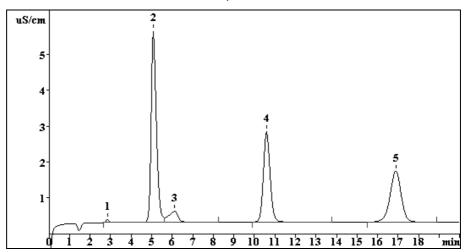
Eluent information from the Eluent tab of the passport.

ELUENT: 2.0 mM NaHCO3 / 1.3 mM Na2CO3

Flow: 0.80 mL/min Temperature: 20.0°C Pressure: 3.4 MPa

# **Chromatogram plot**

Plot of the chromatogram (only for **Printer** or **File** destination).



### Peak table

Peak table with results. The structure of the peak table depends on the selected **Quantification method**. The following report is put out as default:

Quantification method: Standard component: Normalization:		Custom No 100.00			
No	Retention	Height	Area	Conc.	Name
	min	uS/cm	uS/cm*sec	mg/L	
1	2.83	0.09	0.803	0.04	fluoride
2	5.06	5.35	94.134	8.06	chloride
3	6.11	0.31	10.254	0.00	
4	10.57	2.52	57.007	9.06	nitrate
5	16.87	1.43	53.565	6.40	sulfate
5	20.01	9.70	215.763	23.57	
5	20.01	9.70	215.763	23.57	



**Comment** User defined comment from the **Comment** tab of

the passport.

```
Method example for water determination.
```

#### **GLP Log**

Method and Data GLP messages from the **Method Log** and **Data Log** tabs of the passport.

```
METHOD GLP LOG

15/02/1999 17:35:47

Method created !

15/02/1999 17:35:56

Modified: Passport;

Method saved to disk file C:\IC761\Methods\s-03.mtw

DATA GLP LOG

Time constant: 0.100000

19/02/1999 11:26:38 Urs Waldburger

Modified: RAW DATA! Integration; Peaks;

Data saved to disk file C:\IC761\DATA\J2191106.CHW

23/02/1999 10:02:29 Urs Waldburger

Data saved to disk file c:\ic761\data\j2191106.CHW
```

# More items to report

This part of the **Report options** window contains a list of rarely used report items that can be included into the report on the user's choice:

#### Acquisition

Parameters used for data acquisition from the from the **General**, **Measure** and **Filters** tabs of the **Method setup** window.

```
ACQUISITION PARAMETERS
Channels:
Method duration: 20.00min
Run duration: 20.01min
Measurements (method): 12006
Measurements (run):
                      12006
Freq.divisor: 1
Sampling:
                10.00 pts/sec
Device:
                761Compact IC
Interface:
Program before:
Program after:
Spikes filter:
                Yes
Median filter:
                No
  slit:
                0
Gauss filter:
                No
  slit:
                0
```

# Integration

Integration parameters and integration events.

```
INTEGRATION DEFAULTS
Channel:
                 Cond
Delay:
                 2.60 min
Width:
                 5.00 sec
Broadening:
                 2.00
Slope:
                 5.00
Asymmetry:
                 1.50
MinArea:
                 0.00
MinHeight:
                 0.01
Rider ratio:
                 0.00
```



#### Calibration defaults

Calibration parameters defined in the component window and on the **Math** tab of the **Method setup** window.

CALIBRATION Channel: Cond

Method: External standard

Response: Area Standard: No

Index: Linear Internal
Effectivity: 2\*Pi\*(T\*H/A)^2

IDENTIFICATION peaks: Time

Reference peaks: Time Other peaks: Time Retention units: min

# Component table Component table.

No	Retention	Window%	RF	Conc.	Index T	ype Group Name
1	2.85	5.0	1.095e+00	0.04	0.000	0 fluoride
2	4.94	5.0	1.713e+00	8.06	0.000	0 chloride
3	6.78	5.0	2.816e+00	0.00	0.000	0 nitrite
4	8.48	5.0	4.269e+00	0.00	0.000	0 bromide
5	10.56	5.0	3.177e+00	9.06	0.000	0 nitrate
6	13.65	5.0	6.368e+00	0.00	0.000	0 phosphate
7	16.86	5.0	2.391e+00	6.40	0.000	0 sulphate

#### **Calibration results**

Calibration results of the component window. For each component a new page with calibration results and calibration curve is put out.

CALIBRATION OF COMPONENT phosphate Method: Mpoint.mtw  $Q = -2.49731e-07 \cdot A^2 + 0.0111539 \cdot A + 0.121632$ Equation: RSD: 0.250 % Correlation coefficient: 0.999452 Concentration 12.50 Area 10 15 20 25 30 35 40 45 50E+02 K3 = 0K2 = -2.49731e-07K1 = 0.0111539K0 = 0.121632Base: Area Ref.channel: ch1 ISTD: Formula:  $Y=K2 \cdot X^2+K1 \cdot X+K0$ Weight: Level Height Volume Retention Used File Area Conc. 6.46 173 10.8 Yes g8140800.chw 10.8 Yes g8140823.chw 10.8 Yes g8140858.chw 2 34 897 10 1 90.9 2.36e+03 3 25 1 4 198 5.04e+03 50 1 10.8 Yes g8140929.chw



Channel table Parameters of the data acquisition channel

used.

G 3	CHANNELS TABLE								
Cond No	Name	Units Inp	out Minimum	Zero	Maximum	Range	Coefficient	Noise	Shift
1	Cond	uS/cm	1 -8388607	0	8388607	8.389e+01	1.000e-05	5.63	0
# Name 1 Cor uS/		Noise 5.6 5.63e-05	RMS 63047.3 0.63	PeakToPea 796371.8 7.96	3 15119	:/hour 9.570 ).151			

Name Channel's name. This text appears as Channel

label in the chromatogram.

Units Units of detector response (μS/cm).Input Number of data acquisition channel.

Minimum value of the linear range of the AD

converter (in ADC conversion units). This parameter is used to detect an underflow condi-

tion.

**Zero** A signal level on a baseline (in ADC conversion

units). This parameter is a result of the ADC

calibration.

**Maximum** Maximum value of the linear range of the AD in-

terface (in ADC conversion units). This parame-

ter is used to detect overflow condition.

Range An input signal value in maximum in  $\mu$ S/cm.

Range = (Maximum - Zero) • Coef

**Coef** ADC sensitivity coefficient (weight of ADC bit in

 $\mu$ S/cm).

**Noise** Estimated baseline noise value of the channel

in ADC conversion units (bits).

Shift Not used for 761 Compact IC.

# **Report destination**

The following output devices can be selected simultaneously in any combination as targets for report output:

**Screen** Output report (without curves) to screen.

**Printer** Output report to printer.

File Output report to file (see File output options).

#### **Peak table**

The following parameters can be selected for peak table output with determination results:

#### **Quantification method**

Selection of the method to calculate concentrations of components:

#### **Response normalization**

The method normalizes the sum of responses for all peaks of the chromatogram to the **NORM** factor entered in the **Total % for normalization** field:

$$R_i\% = NORM \cdot R_i / \Sigma R_i$$
.

All peaks are calculated and reported no matter whether they have been recognized or not.

The peak table includes the following columns:

Peak number, Retention time, Area + Area% or Height + Height%, Name

#### **Normalized concentration**

The method normalizes the sum of absolute concentrations for all peaks of the chromatogram to the **NORM** factor entered in the **Total % for normalization** field:

$$C_i\% = NORM \cdot W_i(R_i) / \Sigma W_i(R_i)$$

As long as no universal component is defined, only peaks with non-zero concentrations are listed.

The peak table includes the following columns: Peak number, Retention time, Height, Area, Response factor, Concentration%, Name

#### **Absolute concentration**

This quantification method reports raw quantity (absolute concentration) of the component which is calculated directly from the calibration formula:

$$Q_i = W_i(R_i) / V'$$

As long as no universal component is defined, only peaks with non-zero concentrations are listed.

The peak table includes the following columns: Peak number, Retention time, Height, Area, Response factor, Concentration, Concentration%, Name

#### **Relative concentration**

This quantification method uses the internal standard method to calculate the relative concentrations of the components. For this procedure, a **Standard component** must be selected

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	and the concentration of this internal standard must be entered in the <b>Concentration of std</b> field.  As long as no universal component is defined, only peaks with non-zero concentrations are listed.  The peak table includes the following columns: Peak number, Retention time, Height, Area, Response factor, Relative concentration,
	Relative concentration%, Name
Index	This column reports retention indexes of identified components.  The peak table includes the following columns:  Peak number, Retention time, Width/2, Height, Index, Name
Column test	Column test quantification method calculates multiple values that are necessary to evaluate column performance.
	The peak table includes the following columns:  Peak number, Retention time, K' (capacity factor),  TP (number of theoretical plates per column),  TP/m (number of theoretical plates per m),  HETP/dp (reduced height equivalent to theoretical plate), Asym. (peak asymmetry), Name
Custom	This quantification method enables to customize the peak table on the user's choice using the <b><customize></customize></b> button.
	The peak table for the methods supplied contains the following columns as standard:  Peak number, Retention time, Height, Area, Conc., Name
Standard component	Internal standard component for quantification procedure.
Concentration of std	Concentration of the internal standard component for relative concentration calculations. This value is stored in <b>This run</b> column of the concentration table.
Total % for normalization	A value to which the sum of concentrations is normalized. It is used for <b>Response normalization</b> and <b>Normalized concentrations</b> quantification methods. The default value is 100.
Printing order	Determines the order of the components in the peak table.
By peaks	Lists run results for all peaks detected. Unidentified peaks (corresponding to the universal component) are included, but missing components are not listed.



By components	Missing components are always included into the report, even those with concentration equal to zero. In the case of presence of universal component the summary for all unidentified peaks is presented as a single line.
<customize></customize>	Calls up a box with a list of columns that can be included into the report (see <i>Report elements</i> ). It is available only if <b>Quantification method = Custom</b> has been selected.
Report all peaks	Checkbox that includes all peaks irrespective of the concentration of the substance. Otherwise the report generator excludes lines with zero concentration of the component. It is available only if <b>Quantification method = Custom</b> has been selected.
Groups	If this checkbox is checked, a separate peak table report is put out for every group specified in the component table.

# **Template options**

For report output using a report template the following parameters can be modified:

Template	Selection of the report template file *.rtt. Several templates for different languages and applications are available.
Separator	Defines which character separates report col- umns in tables. A separator other than <b>Space</b> is useful when the report is printed to a file and this file is imported into a spreadsheet software.
Tab size	Sets tab stop value for the screen window. Important only when <b>Separator = Tabulation</b> has been selected or there are tab characters in the template file *.rtt.

# File output options

If **File** has been selected for the report destination, the following parameters can be modified:

Directory	Directory for report output. For creation of a new directory, use the <b><browse></browse></b> button.
Name	File name to save report to. The report is saved in text form in the ANSI or ASCII format. So add an extension like *.txt to the file name. If the chromatogram plot checkbox is checked, the plot is saved in a separate file *.wmf in the WMF format under the same name.



Mode The two options Overwrite (overwrite the file) or

Append (append to existing file) are available.

Character set The two options Windows (ANSI) or DOS (ASCII)

are available. This settings are essential for printing of non-English symbols (e.g. ö, ä, ü).

**Custom program** Path and name of the program to be started af-

ter report output. This option enables to transfer the report to a database, an electronic table or another application for further processing.

# Report elements

If **Quantification method = Custom** has been selected, the peak table can be customized on the user's choice. After clicking the **<Customize>** button, the following columns can be included in the peak table.

**number** Peak number.

retention time Retention time of the component (in minutes, ir-

respective of the chosen retention units on the chromatogram graph axes). The total value in the column is equal to the chromatogram dura-

tion.

halfwidth Width of the peak at half height (in minutes).

height Height of the peak (in  $\mu$ S/cm). The total value in

this column is the sum of heights for all identi-

fied peaks.

height% Normalization of peak heights for all peaks us-

ing the **NORM** value entered in the **Total % for normalization** field (default setting 100%):

 $H_i\% = NORM \cdot H_i / \Sigma H_i$ 

area Area of the peak. Depends on the units on the

X and Y axes of the chromatogram. The total value in this column is the sum of areas for all identified peaks (including universal compo-

nent).

area% Normalization of peak areas for all peaks using

the **NORM** value entered in the **Total % for nor-**

malization field (default setting 100%):

 $A_i\% = NORM \cdot A_i / \Sigma A_i$ 

**capacity factor** The capacity factor **k'**; of the component is

equal to the ratio of its corrected retention time

 $(t - t_0)$  to the void time of the system  $t_0$ :

 $\mathbf{k'}_{i} = (\mathbf{t}_{i} - \mathbf{t}_{0}) / \mathbf{t}_{0}$ 

Total for this column is equal to the capacity factor of the last peak of the chromatogram.

**resolution** Resolution **R** for two neighboring peaks is cal-

culated as:

$$\mathbf{R} = (\mathbf{t}_{i+1} - \mathbf{t}_i) / (\mathbf{w}_{0.607i} + \mathbf{w}_{0.607(i+1)})$$

where **i** and **i+1** indexes refer to the neighboring peaks, and  $\mathbf{w}_{0.607}$  stands for the peak width at 60.7 % of the peak height.

effectivity, TP

Effectivity for the peak in number of theoretical plates. The number of theoretical plates  $N_i$  per column for a chosen peak is calculated for a chromatographic peak by one of two formulas:

$$N_i = 2 PI (t_i \cdot H_i / A_i)^2$$

where PI=3.1415926...,  $t_i=$  retention time,  $H_i=$  height,  $A_i=$  area of the peak. The more commonly used formula is:

$$N_i = 5.54 (t_i / w_i)^2$$

where  $\mathbf{w_i}$  is the width on the half-height of the peak. The first formula offers better estimates for fused or unresolved peaks, because the half-width errors for those peaks are much greater than height or area errors.

Total for this column includes average value for the peaks listed.

effectivity, TP/m

Effectivity for the peak in number of theoretical plates per meter. The number of theoretical plates per meter **N'** for the given component is calculated as:

$$N' = N_i \cdot 1000 / L$$

where  $\mathbf L$  is length of the column in mm and  $N_i$  is effectivity of the column for i-th component.

Total for this column includes average value for the peaks listed.

#### reduced TP height, HETP/dp

The height of theoretical plate divided by particle size, called also reduced height, is calculated by formula:

$$H_i = 1000 \cdot L / (N_i dp)$$
.

where  ${\bf L}$  is length of the column in mm,  ${\bf dp}$  is particle diameter in  $\mu m$ .

asymmetry

Peak asymmetry  $A_s$  is calculated at  $^1/_{10}$  of the peak height as a ratio of width after the top of the peak  $w_2$  to the width before the top  $w_1$ .

$$\mathbf{A}_{\mathbf{s}} = \mathbf{w}_2 / \mathbf{w}_1$$

response factor

Coefficient  $\mathbf{k_1}$  of the calibration curve.

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#### raw concentration

Absolute concentrations (raw quantity) of the components, calculated as

$$C_i = W_i(R_i) / V'$$

Total value for the column is the sum of concentrations for all components.

#### concentration%

Normalization of concentrations for all peaks using the **NORM** value entered in the **Total % for normalization** field (default setting 100%):

$$C_i\% = NORM \cdot W_i(R_i) / \Sigma W_i(R_i)$$

Total value for the column equals **NORM** for the main table, and the sum of concentrations in the group for the group table.

#### rel. concentration

Relative concentration of the component relative to the standard component, assumes concentration of one of the components to be known in advance. The concentration of the component is calculated by the formula:

$$C'_i = W_i(R_i) / V' = W_i(R_i) \cdot C_s / W_s(R_s),$$

where  $V' = W_s(R_s) / C_s$  is the effective (reduced) volume of injected sample. The standard component concentration  $C_s$  is entered by the user.

Total value for relative concentration column does not include the concentration of the standard component.

#### rel. concentration%

Relative concentration percent differs from **concentration%** by exclusion of standard component(s) from summation, so that sum of all concentrations (excluding standard) equals **NORM**.

Total value for the column equals **NORM** value for main peak table, and the sum of concentrations in the group for Group table.

index

Linear or logarithmic retention index for identified components.

Total value for the column is a weighted index average with absolute concentration as weight.

$$I = \Sigma (I_i C_i) / \Sigma C_i$$
.

type

The types of components are designated by a one-letter code:

- R Reference component (used for peak identification).
- **s** Standard component.
- **c** Calibration standard in the case it is different from the quantification standard.



? Some points of corresponding peak are out of ADC or detector range – suspicious result.

! Component concentration is outside of minimum (min C) and maximum limits (max C), set in the component table.

Special component (it has one of the special flags checked on in the COMPONENT window).

N The component response is outside of calibrated region.

In the case of peaks additional letters precede the component type information, indicating, how peaks are separated from other peaks, for example:

Peak that starts on the baseline (**B**) and ends on the drop line (**D**) that separates it from another adjacent peak.

Special case of a rider (**R**) peak that is tangentially separated from the main one. A main peak will have a third letter **H** (horse) in this case.

The complete component type may look like: **BBD**: !R.

**group** Number of the group for the component.

spectral ratio Not used for 761 Compact IC.

name Name of the component.

**file name** File name of chromatogram. This column is

very useful for the purpose of processing of ex-

ported data.

ident Title (identifier) for the chromatogram. This col-

umn is useful for the processing of exported

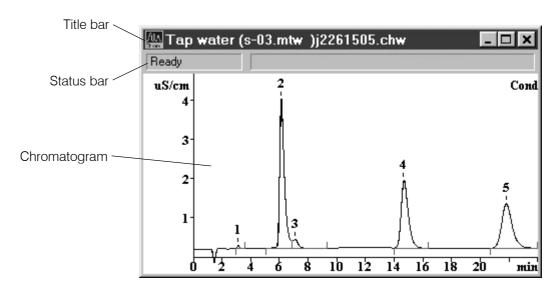
data.

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# 4.5 Chromatograms

# 4.5.1 Chromatogram window

The **CHROMATOGRAM** window is used to show a running or recorded chromatogram.



The **title bar** of the chromatogram window contains buttons for minimizing, maximizing and closing the window. The window name consists of the elements "Ident (method name) chromatogram name". A star (\*) at the end of the name indicates that the chromatogram has been changed since the last saving.

The **status bar** contains two fields. The first field indicates the current measurement status, one of the listed below:

Ready Waiting	Chromatogram is ready to start.  Chromatogram is waiting for the first measuring points.
Measure	Chromatogram is being measured.
Measure(Baseline)	Recording of baseline.
Finished	Measurement finished, but the chromatogram is not processed.
Processing	Chromatogram is being processed after finishing.
Failure	Failure, e.g. unexpected pump stop, etc.
COM error	Error on COM port interface.

During active data acquisition the elapsed time, the duration of the chromatogram or program and the current X and Y values are shown in the second field of the status bar. If the peak editor is switched on then the position of the cursor will be shown.

A chromatogram can be scaled with the help of keyboard or mouse functions or through the **Chromatogram axes** window opened with **761 COMPACT IC** / **View** / **Appearance** / **Chromatogram axes**. Some of the window control functions are collected in the **Window** menu.



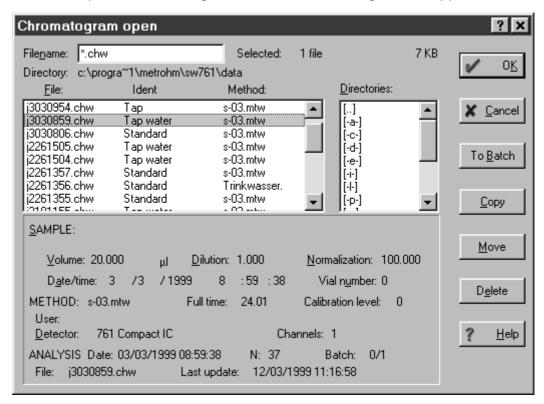
# 4.5.2 Chromatogram file handling

### **Open chromatogram**



# 761 COMPACT IC / File / Open / Chromatogram

Load an existing chromatogram file (\*.chw) from the **Data** directory and open the chromatogram window. The following window appears:



Filename Filename (common wildcards as \* and ? are possible).

Selected Number of selected chromatograms and their total size.

Directory Path name of the working directory.

File window

Box with the list of files residing in the working directory sorted by time (the last recorded chromatogram is at the top of the list). As additional short information on the chromatogram the two parameters **Ident** and **Method** are also displayed. Files in this box can be selected by moving the selection bar to the desired chromatogram and pressing the [Space] key or by a single mouse click.

It is possible to select several chromatograms at once. Press [Shift] and the left mouse button to select all chromatograms up to last item clicked, press [Ctrl] and the left mouse button to add a single chromatogram to the selection already made. All selected items are painted.

Directories	Box with the list of directories. Allows to change the working directory defined in the method, if necessary.
Chromatogram desc	cription
·	A set of fields with the description of the current chromatogram. Selected fields from the passport are listed here.
<0K>	Load all selected chromatogram files (*.chw) and open each in its own window.
<cancel></cancel>	Close this window without any action.
<to batch=""></to>	Add selected chromatograms to the specified batch reprocessing file.
<copy></copy>	Copy selected chromatograms to the specified location.
<move></move>	Move selected chromatograms to the specified location.
<delete></delete>	Delete selected chromatograms and move them to the Windows waste-basket.

# Save chromatogram



#### 761 COMPACT IC / File / Save / Chromatogram

Save the selected chromatogram in a chromatogram file (\*.chw) in the working directory. If this chromatogram has been already stored in this directory, the message File ... exists. Overwrite? appears. If it is preferable to overwrite the previous copy (for example, after the reprocessing), press the <OK> button. Pressing <No> saves the chromatogram into the new file (a new copy of the chromatogram is created).

### **Close chromatogram**

#### 761 COMPACT IC / File / Close

Close the selected chromatogram window. If the data were not saved or the method was modified, a warning appears. Usually it is more convenient to close the window by clicking the button at the top corner area of the window.

# **Delete chromatogram**

# 761 COMPACT IC / File / Delete

Delete the selected chromatogram window. A warning will be generated by the software in any case.

# **Export chromatogram**

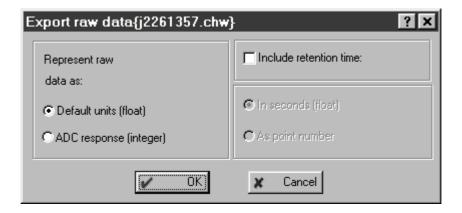
#### 761 COMPACT IC / File / Export / AIA file

Export the selected chromatogram in the AIA format (Analytical Instrument Association) as CDF file (\*.cdf).

### 761 COMPACT IC / File / Export / Raw data to txt

Export chromatographic raw data into an ASCII text file (\*.txt). The following window appears:





**Represent raw data as** Output of raw data:

**Default units (float)** Y values are exported in default units for the

channel (µS/cm).

**ADC** response (integer)

Y values are exported in bits.

Include retention time

In seconds (float)As point numberX values are exported in seconds.X values are exported as point number.

### **Import chromatogram**

# 761 COMPACT IC / File / Import / Chromatogram

Import chromatogram of the following formats:

*.chm	Chromatogram files of the «Chrom&Spec» program (DOS version).
*rd	Chromatogram files of the «714 IC Metrodata» program (DOS version). After selection of the file to be imported you are asked to load a method.
*.chr	Chromatogram files of the «EnviroChrom» program.
*.dar	Chromatogram files of the «AtomChrom» program.
LongInteger (*.*)	Chromatogram files in Long Integer format.

## 761 COMPACT IC / File / Import / AIA file

Import chromatogram in the AIA format (Analytical Instrument Association) from a CDF file (\*.cdf).

# 761 COMPACT IC / File / Import / Raw data from txt

Import chromatographic raw data from a text file \*.txt (ASCII format). This is only possible if an appropriate method has been opened.

# 4.5.3 Graphical representation

### **Appearance**



# 761 COMPACT IC / View / Appearance

This menu item opens the **Appearance** window. It determines the appearance of the chromatogram and consists of four tabs:

**Chromatogram axes** Scaling of chromatogram axes.

**Labels** Settings for peak labels and baseline

drawing.

**Select channel** Select channels to be displayed (only

available for multi-channel chroma-

tograms).

**Colors** Color settings for chromatogram ele-

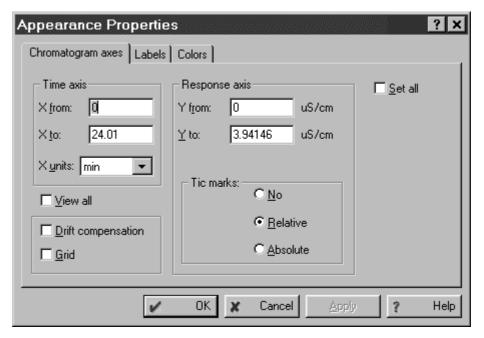
ments.



The appearance settings for a chromatogram are not saved automatically on closing the chromatogram. In order to have the same appearance after reopening the chromatogram, it must be saved explicitly after modification of these parameters with 761 COMPACT IC / File / Save / Chromatogram.

### **Chromatogram axes**

Tab **Chromatogram axes** of the **Appearance** window with parameters for scaling of the chromatogram axes.



### Time axis

**X from** Beginning of the window on X axis.

**X to** End of the window on X axis.

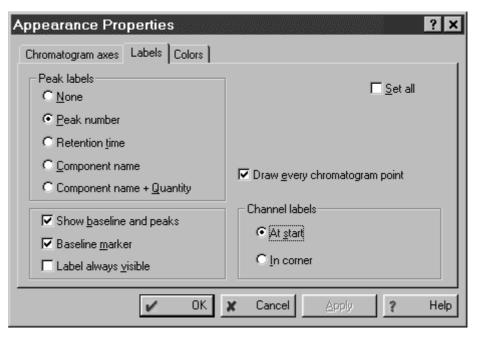
**X units** Selection of unit (Retention unit) for X axis.



Response axis	
Y from	Beginning of the window on Y axis.
Y to	End of the window on Y axis.
Tic marks	Plot axes labels and tic marks on Y axis with <b>no</b> , <b>relative</b> or <b>absolute</b> scaling.
View all	Set scales on X and Y axes so that the whole chromatogram is visible.
Drift compensation	Turns the chromatogram screen image so that the last and first points of the chromatogram are on the same level. This option is useful for gradient chromatograms. Drift compensation has no effect while a chromatogram is running.
Grid	Plot dotted grid lines in the selected chromatogram window.
Set all	Set scales on X and Y axes of all opened chromatograms automatically to the settings in the selected chromatogram window.

### Labels

Tab **Labels** of the **Appearance** window with parameters for peak labeling and baseline drawing.



### Peak labels

None No peak labels.

Peak number Peak number.

Retention time Retention time.

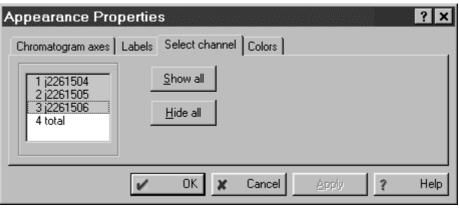
**Component name** Name of the component.



Component name ⊣	<ul> <li>Quantity         Name and quantity of the component.     </li> </ul>
Channel labels	Label for data acquisition channel.
At start	Channel label is shown at the start of the chromatogram.
In corner	Channel label is shown in the upper right corner of the chromatogram.
Show baseline and pea	Baselines are shown.
Baseline marker	Start and end of baselines are marked.
Label always visible	Label is shown always with zooming.
Set all	Set peak labeling of all opened chromatograms automatically to the settings in the selected chromatogram window.
Draw every chromatog	ram point Show chromatogram with all measured points (disable smoothing).

# **Select channel**

Tab **Select channel** of the **Appearance** window for selection of channels for multi-channel chromatograms.

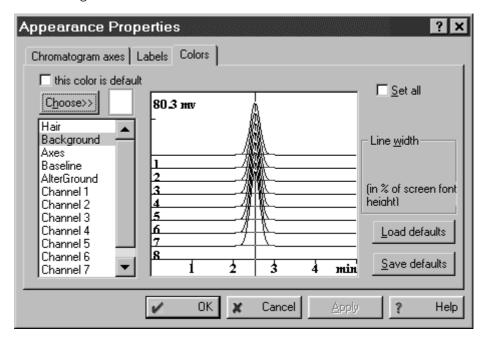


Selection window	The selection window shows all chromatograms of the batch reprocessing queue which can be selected for display. If <b>total</b> is selected, all chromatograms are added and the resulting addition curve is shown.
<show all=""></show>	Select all chromatograms in the selection window.
<hide all=""></hide>	Deselect all chromatograms in the selection window.



# **Colors**

Tab **Colors** of the **Appearance** window with color settings parameters for chromatograms.



this color is default	Reset the selected element to the default color.
<choose></choose>	Choose a new color for the selected element. An example of the chosen color is shown beside the button.
Hair	Cursor's color.
Background	Background color.
Axes	Axes and axes labels color.
Baseline	Baseline color.
AlterGround	Alternative background color for <b>Measure base- line</b> option.
Channel 18	Color of the selected channel. Each channel can be of a unique color.
Line width	Line width for the selected element (Axes or Channel 18) in % of selected plot font height. Range: 0 232
Set all	Set colors of all opened chromatograms automatically to the settings in the selected chromatogram window.
Set all <load defaults=""></load>	matically to the settings in the selected chroma-

# **Other graphical functions**

#### 761 COMPACT IC / View / X full scale

By selecting this menu item or pressing [Ctrl] + [Home] the X axis is scaled so that the chromatogram perfectly fits the window horizontally.

#### 761 COMPACT IC / View / Y full scale

By selecting this menu item or pressing [Ctrl] + [End] the Y axis is scaled so that the chromatogram perfectly fits the window vertically.



# 761 COMPACT IC / View / View all

By selecting this menu item or pressing [Alt] + [V] the X and Y axes are scaled so that the chromatogram perfectly fits the window horizontally and vertically.

#### 761 COMPACT IC / View / Recorder autoscale

This option allows to look at the chromatogram so that the last point is always visible on the screen while data are acquired.

If the **Recorder autoscale** option is switched on, the following autoscaling rules apply:

- If the last acquired point comes out of the window down, autozero is performed.
- If the last point is too high, the recorder scale shrinks twice until the point comes to screen.
- If the last point comes outside of the plotting area to the right, the window is shifted half-screen right.

If the **Recorder autoscale** option is switched off, the window scale does not change automatically during data acquisition.



### 4.5.4 Peak editor

### Switching on/off the peak editor



### 761 COMPACT IC / Process / Peak editor

The peak editor mode is used for subsequent manual correction of the automatic integration for chromatogram peaks (see *section 4.4.4*). It can be used to select the most important points for the peak evaluation (start, end and top of peak, valley between peaks) and move them to the required position.

If the peak editor mode is enabled, the vertical bar cursor appears in the chromatogram window. At the same time, the **Peak** menu appears in the menu bar and the peak editor icons appear in the icon bar of the main window.

The peak editor mode is switched on or off by clicking on pressing [Alt] + [C]. It is also possible to click the right mouse button somewhere in the chromatogram window and to select the **Peak editor** item.

You cannot edit the peak pattern while the component table is active and vice versa, when the peak editor mode is switched on the component table option is disabled.

#### **Peak editor functions**

The functions of the peak editor can be triggered with the corresponding menu items of the **Peak** menu, by the peak editor symbols in the symbol bar or with key combinations.



761 COMPACT IC / Peak / Undo

Undo the last action.



761 COMPACT IC / Peak / Insert peak [Insert]

Add a peak to the chromatogram.



761 COMPACT IC / Peak / Delete peak

[Delete]

Delete selected peak.

### 761 COMPACT IC / Peak / Select nearest point

Move the cursor to the nearest start, top, end, or valley point and select the peak.



# 761 COMPACT IC / Peak / Select start point

Move the cursor to the beginning of the nearest peak and select the peak.



### 761 COMPACT IC / Peak / Select top point

Move the cursor to the top of the nearest peak and select the peak.

150



# 761 COMPACT IC / Peak / Select end point

Move the cursor to the end of the nearest peak and select the peak.



# 761 COMPACT IC / Peak / Select valley point

Move the cursor to the valley of the two nearest peaks.

Δ

### 761 COMPACT IC / Peak / Unselect peak

Delete the selection.

2

761 COMPACT IC / Peak / Move selected point

Move a selected point (start, top, end, valley) to the cursor position.



761 COMPACT IC / Peak / Merge peaks

[+]

Merge two neighboring peaks into a single peak.



761 COMPACT IC / Peak / Fuse peaks

[\*]

Fuse the beginning of the previous and the end of the next peak at the cursor position.



761 COMPACT IC / Peak / Split peaks

[/

Split a single peak into two peaks at the cursor position.



761 COMPACT IC / Peak / Delete all left

Delete all peaks to the left of the selected point.



761 COMPACT IC / Peak / Delete all right

Delete all peaks to the right of the selected point.

# **Moving the cursor**

The cursor can be dragged by the **mouse** when the right mouse button is pressed. Releasing the button will leave the cursor at the new position. The position of the cursor is displayed in the status bar of the chromatogram window.

The cursor can also be moved using the keyboard:

 $[\leftarrow]$  Move cursor left.

[Shift] + [ $\leftarrow$ ] Move cursor left quickly.

 $[ \rightarrow ]$  Move cursor right.

[Shift] + [ $\rightarrow$ ] Move cursor right quickly.

[ Home ] Move cursor to the beginning of the window.

[ End ] Move cursor to the end of window.

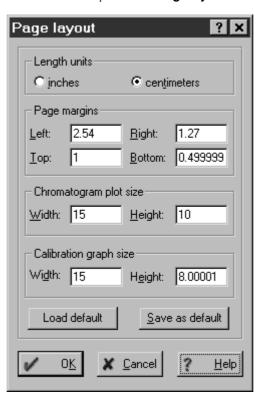


# 4.5.5 Printing

# **Page layout for printing**

761 COMPACT IC / File / Page layout

The selection of this menu item or clicking on <Page...> in the Report options window opens the Page layout window for page layout parameters.



Length units inches centimeters	Selection of length units. Inches. Centimeters.
Page margins	Selection of page margins.
Left Right Top Bottom	Left margin. Right margin. Top margin. Bottom margin.
Chromatogram plot size Width Height	Selection of plot area for chromatograms. Width of plot area. Height of plot area.
Calibration graph size Width Height	Selection of plot area for calibration curves. Width of plot area. Height of plot area.
<load default=""></load>	Load default values for page layout parameters.
<save as="" default=""></save>	Save set page layout parameters as default values.

# **Printer settings**

### 761 COMPACT IC / File / Printer setup

By selecting this menu item the **PRINT SETUP** window is opened where printer, paper size and format can be defined.

# **Print preview**



# 761 COMPACT IC / File / Preview

By selecting this menu item the **Preview** window appears on the screen where the report is shown in the appearance formatted for the desired printer as defined in the **Report options** window.

# **Printing**



### 761 COMPACT IC / File / Print

By selecting this menu item the **Printing** window is opened where printer, printing range and number of copies can be defined. All settings defined in the **Report options** window will be active except the destination of the printout.

In order for the chromatogram to be printed, the option **Chromatogram** plot must be switched on in the **Report options** window under **Items to report**.



# 761 COMPACT IC / Process / Make report

Selection of this menu item opens the **Report options** window for report output to screen, printer or file (details see section 4.4.6).

# 4.5.6 Miscellaneous functions

# Reintegration



# 761 COMPACT IC / Process / Reintegrate

This menu item opens the **Integration parameters** window where the reintegration of the chromatogram can be started (details see section 4.4.4).

# Recalibration

### 761 COMPACT IC / Process / Calibrate

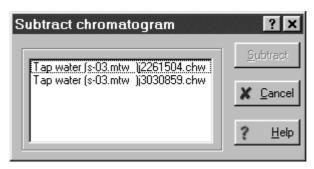
This menu item opens the **Recalibration** window for entry of the calibration level and recalibration of the chromatogram using this level (details see section 4.4.5).



### **Subtraction of a chromatogram**

# 761 COMPACT IC / Process / More / Subtract

This menu item opens the **Subtract chromatogram** window allowing the subtraction of any opened chromatogram from the active, selected chromatogram.

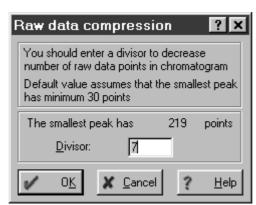


Select the chromatogram that should be subtracted, click on <Subtract> and then on 761 COMPACT IC / View / View all. The result is shown in the active chromatogram window, which is considered as a new chromatogram and will be stored under a new name to avoid overwriting of old data.

### **Data compression**

# 761 COMPACT IC / Process / More / Compress

Selection of this menu item opens the **Raw data compression** window for compression of the raw data of the active chromatogram by summing up several neighboring points.



#### Divisor

Coefficient of compression (the number of data points is reduced by this value). The default value of the compression coefficient is calculated so that the width at half-height of the narrowest peak will be digitized by at least 30 points. If the user enters a coefficient of compression that exceeds the value offered by the software, integration precision may decrease.

### **Invert chromatogram**

#### 761 COMPACT IC / Process / More / Invert!

This menu item inverts the response curves for all channels of the chromatogram so that negative peaks become positive and vice versa (useful for chromatograms with wrong input polarity).

### **Autodatabase options**

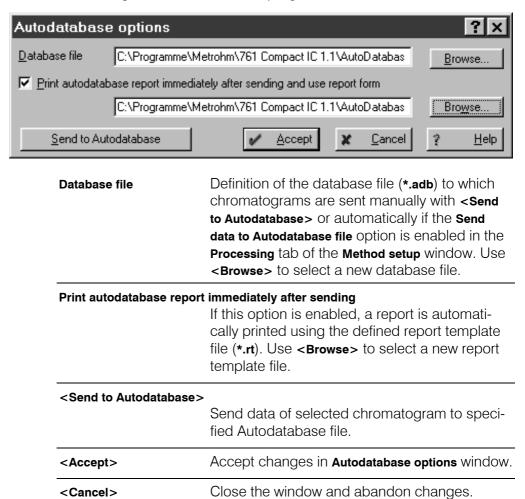


The «Autodatabase 1.0» PC program is a part of the 761 Compact IC standard accessories and is used to save and handle chromatograms and results produced with «761 Compact IC» in a database. You find a detailed description of this database program in the 8.110.8193 Instructions for Use supplied.



#### 761 COMPACT IC / Method / Autodatabase

This menu item opens the **Autodatabase options** window with parameters and settings for the **Autodatabase** program.



# Indicate active Autodatabase program

# 761 COMPACT IC / Options / Indicate AutoDB server

Indicate **Autodatabase** program icon on the task bar if it has been already activated in the background by sending data.



# 4.6 Sample queue

A **sample queue** is a table containing sample-specific data which is used to facilitate work with autosamplers and multiple analyses. The number of rows of the sample queue table defines the number of determinations which are run automatically. Once the queue has been started, the sample-specific data are transferred line-by-line to the running determination overwriting the corresponding fields of the method.

A sample queue is stored in a sample queue file \*.que in a subfolder of the Methods directory.

# 4.6.1 Sample queue file handling

### Open sample queue

761 COMPACT IC / File / Open / Sample queue

SYSTEM / System / Sample queue

These two menu items open the **OPEN** window for opening an existing sample queue file **\*.que** or creation of a new sample queue file **\*.que** by entering a new name. After confirmation with **<OK>** the sample queue overview table is opened (see section 4.6.2).

# Save sample queue



# QUEUE EDITOR / File / Save

Save the sample queue table in a sample queue file (\*.que) in the working directory. The sample queue editor window remains open.



# QUEUE EDITOR / File / Save & exit

Save the sample queue table in a sample queue file (\*.que) in the working directory and close the sample queue editor window.

# SAMPLE QUEUE OVERVIEW / File / Save as

Save a copy of the current sample queue into a new sample queue file (\*.que) in the working directory.

# **Delete sample queue**

#### SAMPLE QUEUE OVERVIEW / File / Delete

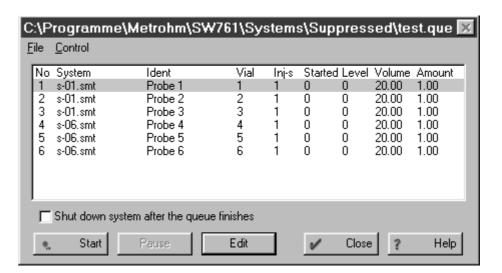
Close the sample queue overview window and delete the sample queue file (\*.que).

# 4.6.2 Sample queue control

### Sample queue overview table

761 COMPACT IC / File / Open / Sample queue SYSTEM / System / Sample queue

These two menu items open the **OPEN** window for opening an existing sample queue file **\*.que** or creation of a new sample queue file **\*.que** by entering a new name. After confirmation with **<OK>** the sample queue overview table is opened showing the current status of the sample queue:



No	Row number.
System	System file to be used for the determination.
Ident	User defined identifier for chromatogram. Will be placed into appropriate passport field when starting the chromatogram.
Vial	Autosampler vial position to take sample from.



This value will not be transferred automatically to the autosampler. Make sure that the vial positions entered in this column are identical to the vial positions used at the autosampler.

Inj-s

Number of injections for the same vial position (only 1 injection is permissible for **Level ≥ 1**).



This value will not be transferred automatically to the autosampler. Make sure that the numbers of injections entered in this column are identical to the numbers of injections used at the autosampler.



Started	Indication whether the sample has been started or not:
	<ul><li>sample not started</li><li>sample started</li><li>number of injections done</li></ul>
Level	Calibration level (see section 4.4.5) for the sample: Level <b>0</b> stays for normal analysis, levels <b>1</b> and greater for calibration runs. Correct filling of this column enables to calculate calibration coefficients automatically.
Volume	Injected volume of sample in $\mu L$ .
Amount	Sample amount. If this value is different for the calibration run (c) and the sample run (s), the component concentrations of the sample are calculated as follows:
	$C_s = C_c \cdot Amount_s / Amount_c$
Shut down system	If this option is enabled, the system hardware is shut down after the sample queue has been finished (all pumps are switched off), but it remains connected.
Shut down system	If this option is enabled, the system hardware is shut down after the sample queue has been fin- ished (all pumps are switched off), but it re-
	If this option is enabled, the system hardware is shut down after the sample queue has been finished (all pumps are switched off), but it remains connected.  Start execution of the sample queue from the
<start></start>	If this option is enabled, the system hardware is shut down after the sample queue has been finished (all pumps are switched off), but it remains connected.  Start execution of the sample queue from the first line with Started = 0.  Stop execution of the sample queue after the

# **Start sample queue**



# SAMPLE QUEUE OVERVIEW / Control / Start

Start execution of the sample queue from the first row with Started = 0. For each row, the sample-specific data are transferred to the method of the current determination and the parameter Started is set to 1 for this row.

If processing the sample queue is interrupted with <Pause> then it can be continued again with <Start> (with a running chromatogram only when data acquisition is finished).



# Pause sample queue



#### **SAMPLE QUEUE OVERVIEW / Control / Pause**

Interrupt execution of the sample queue after the current determination has been finished.

If processing the sample queue is interrupted with <Pause> then it can be continued again with <Start> (with a running chromatogram only when data acquisition is finished).

#### **Cancel last run**

#### SAMPLE QUEUE OVERVIEW / Control / Cancel last run

Cancel the last run (set **Started = 0** for this row). If the queue is restarted, this row is executed again.

This function is only available if the sample queue has been interrupted by pressing the **Pause** button.

# **Reset sample queue**

#### SAMPLE QUEUE OVERVIEW / Control / Reset

Reset sample queue table (set Started = 0 for all rows).

This function is only available if the sample queue has been interrupted by pressing the **Pause** button.

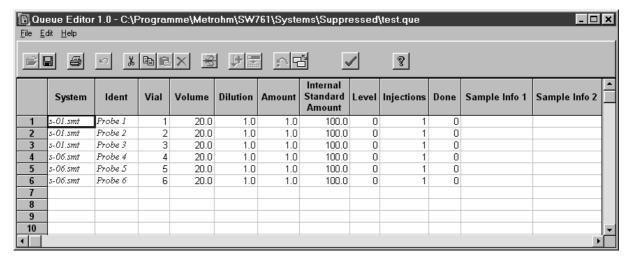
# 4.6.3 Sample queue editor

# Open queue editor window



### SAMPLE QUEUE OVERVIEW / Control / Edit

This menu item opens the **Queue editor** window for editing the sample queue table.





No	Row number.
System	System file to be used for the determination. The system is automatically opened and connected.
Ident	User defined identifier for chromatogram. Will be placed into appropriate passport field when starting the chromatogram.
Vial	Autosampler vial position to take sample from.
0	This value will not be transferred automatically to the autosampler. Make sure that the vial positions entered in this column are identical to the vial positions used at the autosampler.
Volume	Injected volume of sample in μL.
Dilution	Sample dilution prior to injection.
Amount	Sample amount. If this value is different for the calibration run (c) and the sample run (s), the component concentrations of the sample are calculated as follows:
	$C_s = C_c \cdot Amount_s / Amount_c$
Internal standard amount	Concentration of the internal standard component for relative concentration calculations.
Level	Calibration level (see section 4.4.5) for the sample: Level <b>o</b> stays for normal analysis, levels <b>1</b> and greater for calibration runs. Correct filling of this column enables to calculate calibration coefficients automatically.
Injections	Number of injections for the same vial position (only 1 injection is permissible for <b>Level ≥ 1</b> ).
0	This value will not be transferred automatically to the autosampler. Make sure that the numbers of injections entered in this column are identical to the numbers of injections used at the autosampler.
Done	Indication whether the sample determination has been done or not:  o determination not yet done  1n determination or injection done
	···
Sample Info 1	First sample description.

### Sample queue editor functions

The functions in the editor window for sample queues can be triggered with the corresponding menu items of the **Edit** menu or with the corresponding symbols in the symbol bar.

KO

# QUEUE EDITOR / Edit / Undo

Undo the last modification of the sample queue table.



#### QUEUE EDITOR / Edit / Cut row(s)

Cut the selected rows of the sample queue table and copy them into the clipboard.



# QUEUE EDITOR / Edit / Copy row(s)

Copy the selected rows of the sample queue table into the clipboard.



# QUEUE EDITOR / Edit / Paste row(s)

Paste the rows from the clipboard into the sample queue table.



# QUEUE EDITOR / Edit / Delete row(s)

Delete the selected rows from the sample queue table.



# QUEUE EDITOR / Edit / Duplicate row(s)

Duplicate the selected rows of the sample queue table.



#### QUEUE EDITOR / Edit / Increment

Fill selected column fields with values automatically incremented by 1. The last character of the first selected field must be a number. This function is available for the fields **Ident**, **Vial**, **Level**, **Sample Info 1** and **Sample Info 2**.



# QUEUE EDITOR / Edit / Propagate

Fill selected column fields with the same value as the first field.



#### QUEUE EDITOR / Edit / Reset

Reset sample queue table (set **Done = 0** for all rows).



# QUEUE EDITOR / Edit / Change system

Change system file (\*.smt) to be used for the selected rows.

# **Print sample queue**



# **QUEUE EDITOR / File / Print**

Print the sample queue table in landscape format.

### Close sample queue editor



### QUEUE EDITOR / File / Save & Exit

Save the modified sample queue table and close the sample queue editor window.

### QUEUE EDITOR / File / Exit

Close the sample queue editor window. An inquiry appears asking whether or not the sample queue should be saved.



# 4.7 Batch reprocessing

Reprocessing is understood to be the subsequent reprocessing of a series of chromatograms which have been loaded in a **queue (Batch reprocessing queue)**. For reprocessing according to a selected method the settings for calibration, integration, passport, appearance and report can be altered at will.

A batch reprocessing queue is stored in a batch reprocessing queue file \*.bar in the Data directory.

# 4.7.1 Batch reprocessing queue file handling

# Open batch reprocessing queue

### 761 COMPACT IC / File / Open / Batch reprocessing

Load an existing batch reprocessing file (\*.bar) from the Data directory and open the Reprocess window.



### 761 COMPACT IC / File / Open / Last batch

Load the last opened batch reprocessing file (\*.bar) from the Data directory and open the Reprocess window.

# Create new batch reprocessing queue



# 761 COMPACT IC / File / Open / Chromatogram

For creation of a new batch reprocessing queue, open the **Chromatogram open** window. Select the desired chromatograms (\*.chw) and click the <To Batch> button. Enter the name of the new batch reprocessing file and press <OK>. The selected chromatograms are then added to this batch reprocessing queue.



If the reprocessing includes reintegration and/or recalibration, only chromatograms recorded with the same method should be loaded into the batch reprocessing queue.

# Save batch reprocessing queue



### QUEUE EDITOR / File / Save

Save the batch reprocessing queue in a batch reprocessing queue file (\*.bar) in the working directory. The batch reprocessing queue editor window remains open.



# QUEUE EDITOR / File / Save & exit

Save the batch reprocessing queue in a batch reprocessing queue file (\*.bar) in the working directory and close the batch reprocessing editor window.

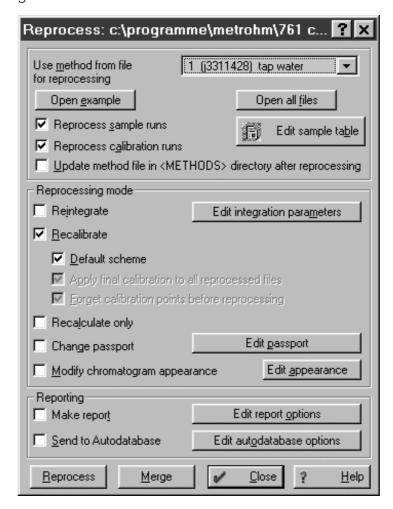
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# 4.7.2 Perform batch reprocessing

# **Reprocess options window**

761 COMPACT IC / File / Open / Batch reprocessing 761 COMPACT IC / File / Open / Last batch

These two menu items are used to open the **Reprocess** window, in which the various options for reprocessing are set and can then be triggered.



#### Use method from file for reprocessing

Selection of the desired chromatogram file whose method should be used for reprocessing.



If the reprocessing includes reintegration and/or recalibration, only chromatograms recorded with the same method should be loaded into the batch reprocessing queue.

<open example=""></open>	Open the chromatogram selected above in the <b>Use method</b> field.
<open all="" files=""></open>	Open all chromatograms of the batch reprocessing queue.



#### <Edit sample table>

Open the batch reprocessing editor program for editing the batch reprocessing table (see section 4.7.3).

#### Reprocess sample runs

Reprocess all sample chromatograms (calibration level = 0).

# Reprocess calibration runs

Reprocess all calibration chromatograms (calibration level > 0).

#### Update method file in <METHODS> directory after reprocessing

Save the method file \*.mtw after reprocessing if the method is changed.

#### Reintegrate

Reintegrate chromatograms according to the current settings of the integration parameters and integration events.

#### <Edit integration parameters>

Open the Integration parameters window for modification of integration parameters and integration events.

#### Recalibrate

Reprocess all calibration chromatograms (if Reprocess calibration runs is switched on) and apply new calibration to all sample chromatograms (if Reprocess sample runs is switched on) by updating the concentration table.

#### Default scheme

Default setting for recalibration reprocessing. The two options Apply final calibration... and Forget calibration points... are switched on. A new calibration is performed with the calibration runs and the resulting new calibration parameters (component table, concentration table and calibration curve) are applied to all sample runs.

### Apply final calibration to all reprocessed files

Apply the updated calibration to all calibration and sample runs. If this option is switched off, the calibration stored in the first chromatogram is used for all other chromatograms.

# Forget calibration points before reprocessing

Forget old calibration points of the calibration curve and perform a new calibration using all calibration runs of the batch reprocessing queue. In this case each calibration chromatogram adds a new point to the calibration curve. If this option is switched off, the calibration curve stored in the first chromatogram remains active. Each further calibration chromatogram in the batch reprocessing queue adds a new point to this calibration curve.

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#### Recalculate only

Enable recalculation of chromatograms with values for **Volume**, **Dilution**, **Amount** and **Internal standard amount** entered in the batch reprocessing table.



The recalculation is done automatically if the Reintegrate and/or Recalculate options are enabled. If the Recalculate only option is enabled, the Reintegrate and Recalculate options are disabled automatically.

#### Change passport

If this option is enabled, those parameters of the passport which have been changed after clicking the **<Edit passport>** button are applied to all chromatograms.

## <Edit passport>

Open the **Passport** window for modification of the passport parameters.



Only some of the passport parameters can be modified. The passport parameters Ident, Sample Info 1 and Sample Info 2 entered in the batch reprocessing table are overwritten if these values are modified in the Passport window.

#### Modify chromatogram appearance

If this option is enabled, the chromatogram **Appearance** parameters which have been changed after clicking the **<Edit appearance>** button are applied to all chromatograms.

#### <Edit appearance>

Open the **Appearance** window for modification of the settings for chromatogram axes, labels and colors.

#### Make report

Print report for all chromatograms using the current report settings of the selected chromatogram in the **Use method...** field.

#### <Edit report options>

Open the **Report options** window for modification of the report settings.

### Send to Autodatabase

Send chromatogram data to Autodatabase file specified in the **Autodatabase options** window (see section 4.5.6).

# <Edit Autodatabase options>

Open the **Autodatabase options** window (see section 4.5.6).

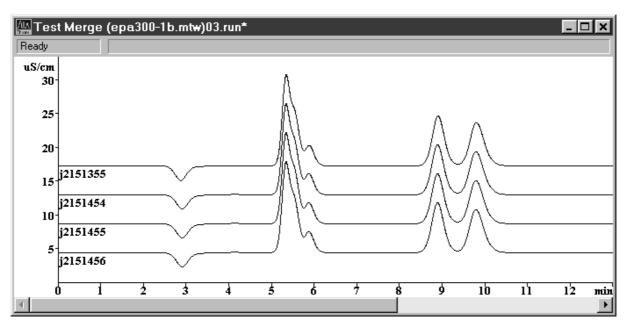


<reprocess></reprocess>	Start reprocessing.
<merge></merge>	Combine all chromatograms of the batch reprocessing queue into a single multi-channel chromatogram.
<close></close>	Close the <b>Reprocess</b> window.

# **Merge chromatograms**

# REPROCESS / < Merge >

Combine all chromatograms of the batch reprocessing queue into a single multi-channel chromatogram.



The chromatograms are displayed in the same order as in the batch reprocessing table slightly displaced one upon the other. The distance between the curves can be increased by pressing [Shift] + [ $\uparrow$ ] and decreased by pressing [Shift] + [ $\downarrow$ ].

The chromatogram axes, labels and colors can be set in the **Appearance** window.

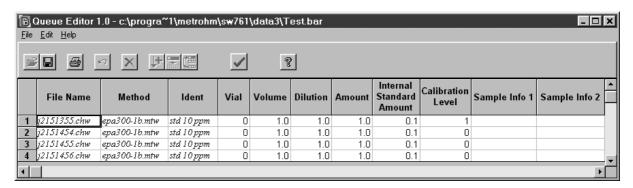
The multi-channel chromatogram can be saved with File / Save / Chromatogram .

# 4.7.3 Batch reprocessing queue editor

# Open batch reprocessing queue editor window

# REPROCESS / <Edit sample table>

This menu item opens the batch reprocessing editor program for editing the batch reprocessing table.



No	Row number.
File name	Name of the chromatogram file (read-only).
Method	Name of the method file *.mtw used for recording the chromatogram (read-only).
Ident	User defined identifier for chromatogram. Will be placed into appropriate passport field when starting the chromatogram.
Vial	Autosampler vial position to take sample from. Will be placed into appropriate passport field.
Volume	Injected volume of sample in $\mu L$ . Will be placed into appropriate passport field.
Dilution	Sample dilution prior to injection. Will be placed into appropriate passport field.
Amount	Sample amount. If this value is different for the calibration run (c) and the sample run (s), the component concentrations of the sample are calculated as follows:
	$C_s = C_c \cdot Amount_s / Amount_c$
	Will be placed into appropriate passport field.
Internal standard amount	Concentration of the internal standard component for relative concentration calculations. Will be placed into appropriate passport field.
Calibration level	Calibration level (see section 4.4.5) for the sample: Level <b>0</b> stays for normal analysis, levels <b>1</b> and greater for calibration runs. Correct filling



	of this column enables to calculate calibration coefficients automatically.
Sample Info 1	First sample description. Will be placed into appropriate passport field.
Sample Info 2	Second sample description. Will be placed into appropriate passport field.

## **Batch reprocessing queue editor functions**

The functions in the editor window for the batch reprocessing queues can be triggered with the corresponding menu items of the **Edit** menu or with the corresponding symbols in the symbol bar.



#### **QUEUE EDITOR / Edit / Undo**

Undo the last modification of the batch reprocessing queue.



#### QUEUE EDITOR / Edit / Delete row(s)

Delete the selected rows from the batch reprocessing queue.



#### **QUEUE EDITOR / Edit / Increment**

Fill selected column fields with values automatically incremented by 1. The last character of the first selected field must be a number. This function is available for the fields **Ident**, **Vial**, **Calibration level**, **Sample Info 1** and **Sample Info 2**.



### **QUEUE EDITOR / Edit / Propagate**

Fill selected column fields with the same value as the first field.



### QUEUE EDITOR / Edit / Rotate rows

Rotate selected rows by one position (the bottom line is moved to the first position; all other lines are moved down by 1 position).

## Print batch reprocessing queue



### **QUEUE EDITOR / File / Print**

Print the batch reprocessing table in landscape format.

# **Close batch reprocessing queue editor**



### QUEUE EDITOR / File / Save & Exit

Save the modified batch reprocessing queue and close the **QUEUE EDITOR** window.

#### **QUEUE EDITOR / File / Exit**

Close the batch reprocessing **QUEUE EDITOR** window without saving the modified batch reprocessing table.

# **5** Notes - Maintenance - Faults

# 5.1 Practical notes on ion chromatography

# 5.1.1 Separating columns

# **Separation efficiency**

The attainable quality of analyses with the 761 Compact IC depends to a large extent on the separation efficiency of the column used. When purchasing an IC column you should ensure that the separation efficiency suffices for the analysis problems at hand. Ascertain the **characteristic data of the IC column** on the standard chromatogram enclosed with the column such as capacity factors, selectivity, plate number and resolution and check these data with your own measurements. If any difficulties arise, you should always first check the quality of the column by recording a **standard chromatogram**.

You will find additional general tips on handling IC separating columns in the **8.732.2003 Metrohm Monograph "Ion chromatography"**, as well as detailed information on the separating columns available from Metrohm (see section 6.3.2) in the leaflets supplied and in the special **Application Bulletins** available free of charge from your nearest Metrohm agency.

#### **Protection**

To protect the column against foreign particles which could have an adverse influence on the separation efficiency, we advise you to subject both the eluents and all samples to **microfiltration** (0.45  $\mu$ m filter) and to siphon the eluent through the **6.2821.090 Aspirating filter**.

To avoid contamination by abrasive particles arising from piston seals of the high-pressure pump, it is advantageous to install an **in-line filter** between the pump and the injection valve. In the 761 Compact IC a **6.2821.120 Filter unit PEEK** is already mounted for this purpose (see section 2.6.2).

The use of readily interchangeable **precolumns** serves to protect the actual separating columns and increase their service life appreciably. The precolumns available from Metrohm (see section 6.3.2) are either actual precolumns or so-called precolumn cartridges which are used together with the 6.2821.040 Cartridge head or the 6.2828.010 Precolumn cartridge holder (see section 2.7).

### **Storage**

Always store the separating columns closed when not in use and filled in accordance with the manufacturer's specifications.

#### **Dead volume**

Dead volume at the end of a column can be the cause of extreme peak broadening or splitting (appearance of double peaks). Filling the column with glass beads ( $\varnothing \le 100~\mu m$ ) frequently improves the separation efficiency.

### Regeneration

If the separation properties of the column have deteriorated, it can be regenerated in accordance with the column manufacturer's specifications. With the separating columns available from Metrohm (see section 6.3.2), the instructions for regeneration can be found on the leaflet enclosed with every column.



In the case of separating columns with carrier material based on silica, **only solutions with pH 2...7** may be used for regeneration, otherwise the columns could be damaged.

# 5.1.2 High-pressure pump

### **Pulsation dampener**

The **MF 6.2620.150 Pulsation dampener** belongs to the standard accessories of the 761 Compact IC; its installation is described in section 2.6.2. It is used to reduce interfering pulsation in highly sensitive measurements and also protects the column material against pressure shocks caused by the injection.

#### **Maintenance**

To protect the pump against foreign particles, we advise you to subject the eluent to **microfiltration** (0.45  $\mu m$  filter) and siphon the eluent through the **6.2821.090 Aspirating filter**.

In many cases, an unstable baseline (pulsation, flow fluctuations) can be traced to contaminated valves or faulty, leaky piston seals.

**Contaminated valves** are cleaned by rinsing with water, RBS solution or acetone (see *section 5.2.6*). When the cleaned valves are reinstalled, you must ensure that the flow direction is correct.

The **replacement of piston seals** is described in section 5.2.6.

Salt crystals between the piston and the seal are the cause of abrasive particles, which can enter the eluent. These lead to contaminated valves, pressure rise and in extreme cases to scratched pistons. It is thus essential to ensure that **no precipitates** can appear (see also section 5.1.3).

#### **5.1.3** Eluents

#### **Treatment**

For the preparation of the eluents only chemicals of a purity degree of at least "p.a." should be used. For diluting please use only **high purity water**.

Fresh eluents should always be **microfiltered** (0.45  $\mu$ m filter) and **degassed** (with N<sub>2</sub>, He or vacuum). For alkaline eluents and eluents with low buffering capacity one should preferably use a **CO<sub>2</sub> absorber** (see section 2.6.3).

The supply vessel containing the eluent must be closed as tightly as possible to avoid excessive evaporation. This is primarily important with eluents containing organic solvents (e.g. acetone), the evaporation of which can lead to drifts in the long term. If work is performed in a very sensitive range, even if one drop of condensate falls back in the eluent this can cause a noticeable change in the background conductivity.

#### Influence of various parameters on anion columns

• Concentration: An increase in the concentration usually leads

to shorter retention times and quicker

separation, but also to a higher background

conductivity.

• pH: pH alterations lead to shifts in the dissociation

equilibrium and thus to changes in the

retention times.

• Organic modifiers: Addition of an organic solvent (e.g. methanol,

acetone, acetonitrile) to aqueous eluents generally accelerates lipophilic ions.

#### **Eluent change**

When the eluent is changed, it must be ensured that **no precipitates** can be formed. Solutions used in direct succession must therefore be miscible. If the system has to be rinsed with an organic solution, several solvents with increasing or decreasing lipophilic character may possibly have to be used (e.g. water  $\leftrightarrow$  acetone  $\leftrightarrow$  chloroform).

### 5.1.4 Peristaltic pump

The pump tubings used by the peristaltic pump are consumable material with a limited lifetime and should be exchanged at regular intervals (approx. every 4 weeks under continuous use; see section 5.2.10).

The working life of pump tubing depends to a considerable extent on the contact pressure. This is why the contact pressure must be correctly set as described in section 2.9.2. If the pump is to remain switched off for a lengthy period of time the tubing cartridges **48** should be raised completely by loosening the snap-action lever **51** on the right-hand side (the set contact pressure remains unchanged).

## **5.1.5** Suppressor module

#### **Protection**

To avoid contamination of the suppressor module by foreign particles or bacterial growth, the two **6.2821.120 Filter units PEEK** (see section 2.6.3) supplied must be installed between peristaltic pump and inlet capillaries of the suppressor module (see section 2.8.2).

#### **Operation**

The **Metrohm Suppressor Module MSM** comprises a total of 3 suppressor units which are in turn used for suppression, regenerated with sulfuric acid and rinsed with water. To record every new chromatogram under comparable conditions, work is normally carried out with freshly regenerated suppressor. Switching takes place automatically together with the valve switching.

For operation of the suppressor module, a **two-channel peristaltic pump** is needed which conveys the regeneration solution (normally **20 mmol/L H\_2SO\_4) and the rinsing solution** (normally **dist. H\_2O**) to the suppressor units (flow rate of 0.5 mL/min).



The suppressor units must never be regenerated with  $H_2SO_4$  in the same flow direction used for the eluent. You should thus always install the inlet and outlet capillaries as described in section 2.8.4 according to the scheme shown in Fig. 18.



The suppressor module must <u>never</u> be switched in the dry state as there is a danger of blocking. Before every switching operation of the suppressor module, the three suppressor units must have been rinsed for at least ½ h with eluent, regeneration and rinsing solution.

The capacity of the suppressor units is exhausted after approx. 2 h. With longer interruptions between the individual measurements it is recommended that, depending on the separating column used, either the **Prep-MSM1** or **Prep-MSM2** system is started during the pauses; this automatically switches the suppressor module further every 20 min.

#### **Maintenance**

In the event of reduced capacity or high counter-pressure the suppressor module must be regenerated (section 5.2.7), cleaned (section 5.2.8) or exchanged (section 5.2.9).

#### 5.1.6 Connections

All connections between injector, column and detector must be as short as possible, have a low dead volume and be absolutely tight. The PEEK capillary after the detector block must be free from constriction (the measuring cell is tested to 5 MPa = 50 bar back pressure).

## 5.2 Maintenance and servicing

#### **5.2.1** General information

#### Care

The 761 Compact IC requires proper care and attention. Excessive contamination of the instrument could possibly lead to malfunctions and a shorter service life of the inherently rugged mechanical and electronic parts.

For protection against escaping liquids the two drain tubes for the inner compartment (section 2.3.3) and for the bottle rack (section 2.3.4) must be mounted.

Spilled chemicals and solvents should be wiped up immediately. It is especially important to protect the plug connections at the rear of the instrument (particular the mains plug) against contamination.



Although constructional measures have been designed to virtually eliminate such a situation, should corrosive media penetrate the interior of the instruments the mains plug of the 761 Compact IC must be immediately disconnected to prevent extensive damage to the instrument electronics. Inform Metrohm service if your instrument(s) have been damaged in such a way.



The instrument must not be opened by untrained personnel. Please comply with the safety notes in section 1.4.1.

#### **Maintenance by Metrohm service**

Maintenance of the 761 Compact IC is best done as part of an annual service performed by specialists from the Metrohm company. If work is frequently performed with caustic and corrosive chemicals, it may be necessary to shorten the interval between servicing.

The Metrohm service department is always willing to offer expert advice on the maintenance and servicing of all Metrohm instruments.

#### 5.2.2 Passivation

Passivation of the complete IC System (without column) by rinsing with  $20...50 \text{ mL } 0.2 \text{ mol/L HNO}_3$  is only required when unusual alterations to the measuring properties of the cell are observed. For passivation the separating column **81** is removed from the 761 Compact IC. The two capillaries **28** and **45** (see *Fig. 14* and *Fig. 16*) are directly connected to each other using the Coupling **33** (6.2620.060).

## 5.2.3 Recycling

To keep the eluent consumption between injections to a minimum when the system is at rest (e.g. overnight), the so-called recycling procedure can be used. In recycling the eluent exiting the outlet capillary of the detector block is led back directly to the eluent supply vessel **67**. The IC system is thus quickly ready for new injections without a long conditioning period.

Instead of threaded stopper **65** the 4.420.0311 Tubing nipple (M6), which belongs to the accessories of the 6.1602.160 Bottle attachment, can be used together with the second E.301.0021 O-ring (which is also included) for returning the eluent (see *section 2.6.3*).



The recycling procedure must **<u>not</u>** be used

- in operation with the suppressor module,
- with alkaline eluents,
- with the 6.1010.000 IC Cation column METROSEP Cation 1-2.

#### 5.2.4 Shutdown

If the 761 Compact IC is shut down for a considerable length of time, the entire IC system (<u>without</u> column and suppressor) must be **rinsed** free from salt with methanol/water (1:4) to avoid crystallization of eluent salts with the corresponding subsequent damage.

For rinsing the connections to the separating column and the suppressor module are removed; the two capillaries **28** and **45** (see *Fig. 13* and *Fig. 14*) are directly connected to each other using coupling **33** (6.2620.060). Rinse with methanol/water (1:4) until the conductivity drops below 10  $\mu$ S/cm.

## 5.2.5 Changing separating columns

## **Identical separation system**

If you wish to replace an IC separating column by a column of the same type, proceed as follows (see *Fig. 14* and *Fig. 16*):

#### 1 Remove old column

- Switch off high-pressure pump and wait for pressure to drop.
- Unscrew column **81** from inlet capillary **45** of the detector block or from suppressor inlet capillary **96**.
- Unscrew column 81 from column connection capillary 28 or the precolumn.

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#### 2 Connect new column to injector

- Remove end caps from column 81.
- Screw inlet end of separating column **81** (note flow direction) to column connection capillary **28** or to the precolumn (see section 2.7.7/section 2.7.8).

#### 3 Rinse column

- Place beaker beneath the column outlet.
- Switch on high-pressure pump and rinse column with eluent for ca. 10 min, then switch off pump.

#### 4 Connect column to detector block

 Screw outlet end of separating column 81 to inlet capillary 45 or suppressor inlet capillary 96.

#### **Changing the separation system**

If you wish to replace an IC separating column by a column of a different type, proceed as follows (see Fig. 14 and Fig. 16):

#### 1 Remove old column

- Switch off high-pressure pump and wait for pressure to drop.
- Unscrew column **81** from inlet capillary **45** of the detector block or from the suppressor inlet capillary **96**.
- Unscrew column 81 from column connection capillary 28 or the precolumn.

#### 2 Rinse with eluent

- Place beaker beneath the column connection capillary 28.
- Rinse IC system with eluent used for the separating column (flow rate 1 mL/min) for ca. 15 min.

#### 3 Connect new column to injector

- Remove end caps from column 81.
- Screw inlet end of separating column 81 (note flow direction) to column connection capillary 28 or to the precolumn (see section 2.7.7/section 2.7.8).

#### 4 Rinse column

- Place beaker beneath the column outlet.
- Set flow rate for new separating column.
- Switch on high-pressure pump and rinse column with eluent for ca. 10 min, then switch off pump.

#### 5 Connect column to detector block

 Screw outlet end of separating column 81 to inlet capillary 45 or suppressor inlet capillary 96.



## 5.2.6 Maintenance work at the pump head

In many cases, an unstable baseline (pulsation, flow fluctuations) can be traced to contaminated valves or faulty, leaky piston seals at the high-pressure pump. For cleaning contaminated valves and/or replacement of wear parts such as pistons, piston seals and valves, proceed as follows:

#### 1 Detach pump head

- Disconnect aspirating tubing **63** from aspirating capillary **40** at the pump head **42** (see *Fig. 3* and *Fig. 4*).
- Unscrew connection capillary **43** from the pump head **42**.
- Remove pump head 42 by loosening the 4 hexagon screws
   41 using the 6.2621.030 Hexagon key. The main piston is on the left (when viewed from front), the auxiliary piston on the right.

## 2 Disassemble pump head

- Strip down pump head 42 in accordance with Fig. 19. Main and auxiliary pistons are identical with the following exceptions:
- The spring 107 of the auxiliary piston (right piston) is more powerful (longer) than that of the main piston (left piston).
- Inlet and outlet valve are not present in the secondary cylinder.



To prevent the piston **105** suddenly jumping out of the piston cartridge **107**, the screw **104** must be undone very carefully by hand.

## 3 Cleaning/replacement of piston 105

- Pistons contaminated by abrasive particles or deposits are cleaned with scouring powder and rinsed free of any particles with dist, water.
- Relatively badly contaminated or scratched pistons must be replaced (spare part: 6.2824.070 Zircon piston).

#### 4 Replacement of piston seal 112

• To remove damaged piston seals **112** the special tool **116** is used. This is screwed into the seal **112**, which can then be pulled out (see *Fig. 20A*).



When the special tool **116** is screwed into the piston seal **112** the latter is completely destroyed!



If you use only aqueous eluents the 6.2741.000 Piston seal can be replaced by the 6.2741.010 PE Piston seal available as an option.

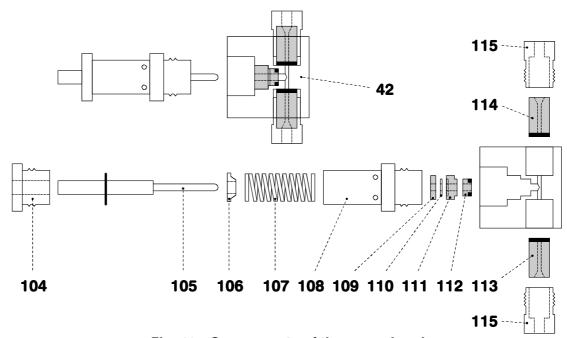


Fig. 19: Components of the pump head

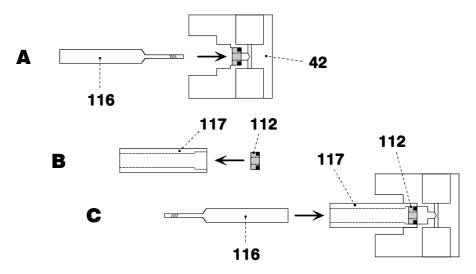


Fig. 20: Replacement of the piston seal 112

42	Pump head (6.2824.100)	111	Piston guide sleeve (4.709.4370)
104	Screw (3.709.1100) for piston cartridge 108	112	Piston seal (6.2741.000/6.2741.010)
105	Zircon piston (6.2824.070) with piston shaft	113	Inlet valve (6.2824.090)
106	Spring retainer	114	Outlet valve (6.2824.080)
107	Spring (6.2824.050) for main piston or Spring (6.2824.060) for auxiliary piston	115	Screw holder for valve
108	Piston cartridge (4.709.0760)	116	Special tool (6.2617.010) to remove the piston seal 112
109	Piston guide sleeve (4.709.4380)	117	Special tool (6.2617.010) to install the piston seal 112

110 Sapphire supporting ring (6.2824.030)



- To install a new piston seal **112** the special tool **117** is used.
- First the new seal is inserted firmly in the recess of tool **117** by hand (see *Fig. 20B*). The seal spring must be located on the outside.
- The tool **117** together with the seal is then inserted in the pump head **42** and the seal pressed into the pump head recess with the aid of tool **116** (see *Fig. 20C*).



The seal surface in the pump head **42** must not be damaged (avoid contact with tool)!

# 5 Cleaning/replacement of inlet valve 113 and outlet valve 114

- Contaminated or blocked valves are cleaned by rinsing with dist. water, RBS solution or acetone. The rinsing effect can be reinforced by brief treatment in an ultrasonic bath (max. 20 s; if longer the sapphire sphere of the valve can be damaged).
- If this does not have the desired effect, the valves can be disassembled as shown in *Fig. 21*. The valve components are pushed out with the aid of a syringe needle inserted through the upper opening in the valve housing **118**. The individual components are rinsed with dist. water and/or acetone, and the sapphire sphere cleaned with a paper towel. The valve is then reassembled in accordance with *Fig. 21*. The components of the inlet and outlet valves are identical, they are distinguished only by the positioning of the sapphire sleeve **121** and the ceramic holder **123** (see *Fig. 21*).
- Valves that fail to function faultlessly after such cleaning must be replaced.
- In the reinstallation of the inlet valve **113** or the outlet valve **114** on no account must the two outwardly identical valves be interchanged. To determine which valve is which, note that the liquid flows through the pump head from the bottom up. The flow direction of the valves can be checked simply by blowing through the clean valve. Both valves are installed with the black face in the direction of the pump head (see *Fig. 19*).



If by mistake an inlet valve 113 is installed instead of the outlet valve 114, an extreme pressure buildup occurs within the working cylinder, which is not detected by the pressure transducer and will destroy the piston seal 112!

### 6 Mounting the pump head

- Reassemble the components of the pump head **42** as shown in *Fig. 19*. Tighten the screw **104** by hand. First screw in piston cartridge **108** manually until the stop is reached and then use a wrench to turn it through a further 15°. Firmly tighten the two valve screw holders **115** with a wrench.
- Reattach pump head 42 to the pump with the help of the 4 fixing screws 41. Firmly tighten them with the 6.2621.030 Allen key.



To ensure that the pump head is not positioned wrongly, the holes at the rear for the clamping bolts have different depths, i.e. 1 clamping bolt is longer than the rest. The deepest hole must naturally accommodate the longest bolt. If this is not the case, the pump will not function properly.

- Screw connection capillary **43** to pump head **42** (see *Fig.* 3 and *Fig.* 4).
- Connect aspirating tubing **63** to aspirating capillary **40** at the pump head **42**.

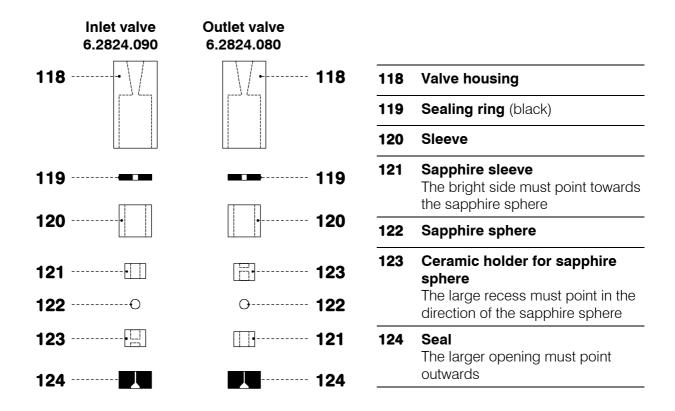


Fig. 21: Components of inlet valve 113 and outlet valve 114

## 5.2.7 Regeneration of the suppressor module

# Regenerating a suppressor operating at reduced capacity

If the suppressor units are exposed to certain heavy metals (e.g. iron) or organic contaminants for long periods of time, these can no longer be completely removed by the regeneration solution normally used (20 mmol/L  $H_2SO_4$ ). This diminishes the capacity of the suppressor units, which, in milder cases, results in a reduced sensitivity to phosphates and, in severe cases, in a strong increase in the baseline. If such capacity problems occur at one or several positions, the suppressor units must be treated as follows:

## 1 Disconnect suppressor from IC system

• Disconnect suppressor from separating column and detector.

## 2 Regenerate suppressor

 Rinse each suppressor unit for about 15 min with one of the following solutions:

#### Contamination with heavy metals

1 mol/L H<sub>2</sub>SO<sub>4</sub>

#### Contamination with organic cationic complexing agents

0.1 mol/L H<sub>2</sub>SO<sub>4</sub>/ 0.1 mol/L oxalic acid / acetone 5%

#### Severe contamination with organic substances

0.2 mol/L H<sub>2</sub>SO<sub>4</sub>/ acetone ≥ 20%



The 6.1826.060 pump tubing is made of PP and must not be used for rinsing with solutions which contain organic solvents. In such cases, rinse with different pump tubing or a different pump.

## 3 Connect suppressor to IC system

• Reconnect suppressor to the IC system. If capacity problems persist, replace the suppressor rotor (see section 5.2.9).

# Regenerating the suppressor in case of high counterpressure

If excessive counterpressure is observed in one or several suppressor units, treat the units as follows:

### 1 Disconnect suppressor from IC system

• Disconnect suppressor from separating column and detector.

#### 2 Regenerate suppressor

Connect the inlet capillary 99 marked "H<sub>2</sub>SO<sub>4</sub>" to the inlet capillary 26 using coupling 33 (see Fig. 10 and Fig. 16). This connects the suppressor module directly to the high-pressure pump.

- Set the flow at the high-pressure pump to 0.5 mL/min and rinse the suppressor unit with 1 mol/L H<sub>2</sub>SO<sub>4</sub> for 5 to 10 min.
- As the pressure falls, slowly increase the flow at the highpressure pump to 2 mL/min. Do not exceed a maximum pressure of 2 MPa (20 bar).
- Switch off high-pressure pump.
- Switch suppressor to the next position using the button.
- Connect the inlet capillary 98 marked "H<sub>2</sub>O" to the inlet capillary 26 using coupling 33 (see Fig. 10 and Fig. 16).
- Set the flow at the high-pressure pump to 0.5 mL/min and rinse the suppressor unit with 1 mol/L H<sub>2</sub>SO<sub>4</sub> for 5 to 10 min.
- As the pressure falls, slowly increase the flow at the highpressure pump to 2 mL/min. Do not exceed a maximum pressure of 2 MPa (20 bar).
- Switch off high-pressure pump.

#### 3 Connect suppressor to IC system

- Connect inlet capillaries **98** and **99** to the peristaltic pump (see section 2.8.4).
- If the pressure problems persist, replace the suppressor rotor (see section 5.2.9).

## 5.2.8 Cleaning the suppressor

It may be necessary to clean the suppressor in the following cases:

- High counterpressure on the suppressor connection tubing
- Irremediable blockage of the suppressor (the suppressor can no longer deliver solutions)
- Irremediable obstruction of the suppressor (the suppressor can no longer be switched to next position)

To clean the connection piece and the rotor, proceed as follows (see Fig. 22):

#### 1 Disconnect suppressor from IC system

- Disconnect input capillary **96** of the suppressor module **47** from the separating column **81** (see *Fig. 16*).
- Disconnect output capillary **97** from inlet capillary **45**.
- Disconnect inlet capillaries **98** and **99** from the filter units **96** (supply from peristaltic pump).

## 2 Dismantle suppressor

- Unscrew nut 125 from suppressor holder 128.
- Pull out connection piece 126 and suppressor rotor 126 from suppressor holder 128 (the connection piece and the rotor normally stick together).
- Loosen connection piece **126** from suppressor rotor **127**.

#### 3 Clean input and output leads

- Connect each of the 6 capillary tubings attached to connection piece 126 to the high-pressure pump one after another, and pump through ultrapure water.
- Check whether solution emerges from connection piece 126.
   If one of the input or output leads remains blocked, replace the connection piece (order number 6.2832.010).

## 4 Clean suppressor rotor

 Clean the sealing surface of suppressor rotor 127 using a lint-free cloth and ethanol.

## 5 Insert suppressor rotor

- Insert suppressor rotor 127 in suppressor holder 128 in such a way that the tubing connections at the rear of the rotor fit in the corresponding openings inside the rotor, and that one of the three holes in the rotor can be seen from below in one of the openings of the holder.
- If the rotor has been inserted correctly, its sealing surface will be about 4 mm inside the holder. If this is not the case, bring the rotor into the correct position from below with the aid of a sharp object (e.g. a screwdriver).

#### 6 Clean connection piece

 Clean the sealing surface of connection piece 126 using a lint-free cloth and ethanol.

#### 7 Insert connection piece

- Insert connection piece 126 in suppressor holder 128 in such a way that connection "1" is at the top, and that the three lugs on the connection piece fit in the corresponding openings of the holder.
- Screw nut **125** onto the thread of suppressor holder **128** manually (do not use tools).

#### 8 Connect and condition the suppressor

- Reconnect the suppressor to the IC system.
- Before switching the suppressor to the next position for the first time, rinse all 3 suppressor units with solution for 5 min.

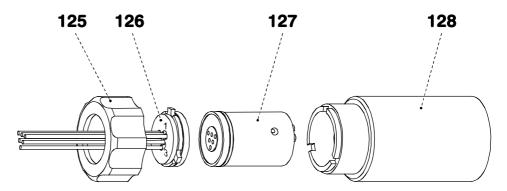


Fig. 22: Assembling the suppressor

125	Screw nut	127	Suppressor rotor (6.2832.000)
126	Connection piece (6.2832.010) with input and output leads	128	Suppressor holder

## **5.2.9 Replacing the suppressor**

The suppressor in the suppressor block may have to be replaced in the following cases:

- Irremediable loss of suppressor capacity (reduced phosphate sensitivity and/or strong rise in baseline)
- Irremediable blockage of the suppressor (the suppressor can no longer deliver solutions)

Both the 6.2832.000 Suppressor rotor and the 6.2832.010 Connection piece with the input and output leads can be replaced. To replace these components proceed as follows (see *Fig. 22*):

### 1 Disconnect suppressor from IC system

• Disconnect all input and output leads of the suppressor from IC system and peristaltic pump.

#### 2 Dismantle suppressor

- Unscrew nut 125 from suppressor holder 128.
- Pull out connection piece **126** and suppressor rotor **127** from suppressor holder **128** (the connection piece and the rotor normally stick together).
- Loosen connection piece **126** from suppressor rotor **127**.

#### 3 Clean suppressor rotor

 Clean the sealing surface of new suppressor rotor 127 (6.2832.000) using a lint-free cloth and ethanol.

#### 4 Insert suppressor rotor

- Insert new suppressor rotor 127 in suppressor holder 128 in such a way that the tubing connections at the rear of the rotor fit in the corresponding openings inside the rotor and that one of the three holes in the rotor can be seen from below in one of the openings of the holder.
- If the rotor has been inserted correctly, its sealing surface will be about 4 mm inside the holder. If this is not the case, bring the rotor into the correct position from below with the aid of a sharp object (e.g. a screwdriver).

#### 5 Clean connection piece

• Clean the sealing surface of new connection piece **126** (6.2832.010) with the aid of a lint-free cloth and ethanol.

## 6 Insert connection piece

- Insert new connection piece 126 in suppressor holder 128 in such a way that connection "1" is at the top and that the three lugs on the connection piece fit in the corresponding openings of the holder.
- Screw nut **125** onto the thread of suppressor holder **128** manually (do not use tools).

#### 7 Connect and condition the suppressor

- Reconnect the suppressor to the IC system.
- Before switching the suppressor to the next position for the first time, rinse all 3 suppressor units with solution for 5 min.

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## 5.2.10 Exchanging the pump tubing

The pump tubings used by the peristaltic pump are consumable material with a limited lifetime and should be exchanged at regular intervals (approx. every 4 weeks under continuous use).

The working life of pump tubing depends to a considerable extent on the contact pressure. This is why the contact pressure must be correctly set as described in section 2.9.2. If the pump is to remain switched off for a lengthy period of time the tubing cartridges **48** should be raised completely by loosening the snap-action lever **51** on the right-hand side (the set contact pressure remains unchanged).

As the pump is always operated on the same side the 6.1826.060 Pump tubings supplied can be used on both sides. To exchange a pump tubing proceed as follows:

## 1 Remove old pump tubing

- Press contact pressure lever 49 on the tubing cartridge 48 down as far as it will go.
- Release tubing cartridge 48 from holding clamp 50 by pressing down snap-action lever 51 and remove from mounting pin 53 (see Fig. 16).
- Remove old pump tubing 92 or 93.

### 2 Insert new pump tubing

- Insert the new pump tubing 92 or 93 in the tubing cartridge
   48 as shown in Fig. 15. The white-yellow stopper 94 must click into the corresponding holder on the left-hand side of the tubing cartridge.
- Place the tubing cartridge 48 on mounting pin 53 and press down on the right-hand side until snap-action lever 51 clicks into position on holding clamp 50. Take care that no kinks are formed in the pump tubing.

#### 3 Set contact pressure

- Switch on peristaltic pump.
- Press contact pressure lever 49 upwards until the solution just starts to be drawn in. Then press contact pressure lever upwards until it clicks once more to obtain optimal contact pressure.
- Switch off peristaltic pump.



## 5.3 Faults and malfunctions

## **5.3.1** Error messages

If any type of malfunction occurs during operation of the 761 Compact IC, this is shown by error messages in the PC program, which appear either in an error window or in the **SYSTEM STATE** window.

Follow the instructions given in the **error window** and close this window with **<OK>**.

You will find further details of the error messages of the **SYSTEM STATE** window, their possible causes and the procedure for their rectification in section 4.3.8.

#### 5.3.2 Malfunctions and their rectification

If difficulties appear with the 761 Compact IC during analyses, their causes are best investigated in the order **separating column**  $\rightarrow$  **high-pressure pump**  $\rightarrow$  **eluent**  $\rightarrow$  **connections**. Several of the malfunctions which may appear are listed in the following table with details of possible causes and countermeasures.

Malfunction	Cause	Rectification
Baseline with high noise level, pulsation	<ul><li>Contaminated pump valves</li><li>Faulty piston seals</li></ul>	<ul> <li>Clean the valves (see section 5.1.5)</li> <li>Replace the piston seals (see section 5.1.5)</li> </ul>
Drift of the baseline	<ul> <li>Thermal equilibrium not yet reached</li> <li>Leak in system</li> <li>Evaporation of organic solvent in eluent</li> </ul>	<ul> <li>Condition system with heating switched on</li> <li>Check connections and make leakproof</li> <li>Ensure better closure of eluent supply vessel</li> </ul>
Considerable pressure drop	Leak in system	Check connections and make leakproof
Considerable pressure rise	<ul> <li>Contamination of the filter in the 6.2821.120         Filter unit PEEK</li> <li>Contamination of the column inlet filter</li> <li>Change of column packing by injection of contaminated samples</li> </ul>	<ul> <li>Replace the 6.2821.130         Filter (see section 2.3.6)     </li> <li>Clean or replace         6.2821.020 Steel mesh(es)     </li> <li>Regenerate column (see section 5.1.1) or replace column         Note:         Samples should always be microfiltered.     </li> </ul>



Malfunction	Cause	Rectification
Chromatograms with poor resolu- tion, change in the retention times	Deterioration in separation efficiency of the IC column	Regenerate column (see section 5.1.1) or replace column
Extreme peak broadening, splitting (double peaks)	Dead volume at the column ends	• Fill column with glass beads (Ø ≤ 100 μm) or replace column
No feed of regeneration or rinsing solution for suppressor	<ul> <li>Contact pressure too low</li> <li>Leak in system</li> <li>Defective pump tubing</li> <li>Contamination of the filter in the 6.2821.120 Filter unit PEEK</li> <li>Counterpressure at suppressor module too</li> </ul>	<ul> <li>Adjust contact pressure (see section 2.9.2)</li> <li>Check connections and make leakproof</li> <li>Replace pump tubing (see section 5.2.9)</li> <li>Replace the 6.2821.130 Filter (see section 2.3.6)</li> <li>Clean or replace suppressor (see section 5.2.65.2.8)</li> </ul>

## 5.4 Diagnostic tests / Validation / GLP

The requirements of **GLP** (**G**ood **L**aboratory **P**ractice) include a periodic check of analytical measuring instruments with regard to their reproducibility and accuracy using **S**tandard **O**perating **P**rocedures, **SOP**.

Under the title **«Application Bulletin No. 277 – Validation of Metrohm ion chromatographs»** an example of such a standard operating procedure is available from Metrohm; it can be adapted and used with the 761 Compact IC.

Further information on the subjects of QA, GLP and validation can also be found in the brochure **«Quality management with Metrohm»**, which is available from your local Metrohm agency.

Testing of the electronic and mechanical function groups of Metrohm instruments can and should be performed as part of a regular service by trained personnel of the manufacturing company (see section 5.2.1). All Metrohm instruments are equipped with start-up-test routines which check for perfect functioning of the relevant assemblies when the instrument is switched on. If no error message is displayed, it may be assumed the instrument is operating without faults.

The Metrohm company also supplies its instruments with an integrated diagnostic program which, in the case of possible malfunctions or faulty behavior, allows the service technician to check the functioning of certain assemblies and localize the fault.

# 6 Appendix

## 6.1 Technical data

## **6.1.1 Conductivity measurement**

Measurement range 1 0...1000  $\mu$ S/cm (resolution: 0.56 nS/cm) Measurement range 2 0...250  $\mu$ S/cm (resolution: 0.14 nS/cm) Measurement range 3 0...50  $\mu$ S/cm (resolution: 0.028 nS/cm)

Maximum error ± 1 % of full scale value and

 $\pm$  1 % of measurement value (k = 16.7/cm)

Linearity Deviations  $< \pm 0.5 \%$  of full scale value

Noise

Measurement range 1 typ. 10 nS/cm Measurement range 2 typ. 2.5 nS/cm Measurement range 3 typ. 0.5 nS/cm

Drift (electronic) typ. < 10 ppm/h of full scale value

Temperature dependence typ. < 40 ppm/°C of full scale value

Reserve range > 33 % (k = 16.7/cm)
Sampling rate 10 measurements/s (fixed)

## **6.1.2 Conductivity detector**

Construction Thermostatted conductivity detector with

2 ring-shaped steel electrodes

Measurement principle Alternating current measurement with 1 kHz

frequency and ca. 1.7 V amplitude (peak to

peak).

Effective cell volume 0.8 µL

Cell constant approx. 17 /cm (the exact value is printed on the

detector)

Maximum back pressure

for measuring cell

5.0 MPa (50 bar)

Thermostatting Connectable dynamic control to adjustable

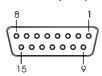
operating temperature

Operating temperature Adjustable in steps of 5°C from 25...45°C

Max. temperature deviation  $\pm 2.5^{\circ}$ C
Heating time  $\geq 30 \text{ min}$ 

Temperature stability ≤ 0.01°C at constant ambient temperature

Connection for detector block Dsub 15 pin (female)





## 6.1.3 Injection valve

Actuator switching duration 100...150 ms

Pressure resistance 25 MPa (250 bar)

## 6.1.4 High-pressure pump

Type Serial dual piston pump with two valves

Pump capacity

Flow range 0.20...2.5 mL/min

Maximum error  $< \pm 2$  % of set value

Flow constancy < 0.5 % of set value

Reproducibility of eluent typ. better than  $\pm 0.1$  %

flow

Pressure measurement

Pressure range 0...25.0 MPa (0...250 bar)

Residual pulsation < 1 % (at 1 mL/min water and 10 MPa pressure,

without pulsation dampener)

Measurement principle Piezoresistive measurement

Response time: 3 ms

Measurement volume: ca. 50 μL

Maximum error ± 3 % of set value

Resolution 0.1 MPa (conductivity measurements)

0.01 MPa (pressure measurements)

Sampling rate 1 measurement/piston stroke (pump running)

1 measurement/s (pump not running)

10 measurements/s (pressure measurements)

28.5 µL

Safety shutdown

Function Automatic shutdown when upper and lower

pressure limits violated

Maximum pressure limit adjustable between 0.1...25.0 MPa (1...250 bar)

Response time: 1 pump cycle

Minimum pressure limit adjustable between 0.1 ... 25.0 MPa (1...250 bar),

inactive at 0 MPa

Main piston:

Response time: 5 pump cycles

Pump head

Pump head volumes Main piston: 40 μL

Priming piston: 20 µL

Pump displacement

volumesPriming piston:14.25 μLLength of strokeMain piston:3.6 mm

Priming piston: 1.8 mm

## 6.1.5 Peristaltic pump

Type 2-channel peristaltic pump

Pump capacity

Rotational speed 20 U/min at 50 Hz

24 U/min at 60 Hz

Flow range 0.5...0.6 mL/min with 6.1826.060 Pump tubing

Maximum error  $\pm 5 \%$ 

Maximum pressure 0.4 MPa (4 bar)

Pumpable liquids Clear liquids with no solid contents

Pump tubing material PP (polypropylene)

## 6.1.6 Suppressor module

Switching duration 140 ms

Pressure resistance 2.5 MPa (25 bar)

## 6.1.7 Leak detector

Type Detector with 2 electrodes approx. 1 mm above

base of interior

Response level Resistance  $< 1 M\Omega$  (for deion. water)

#### 6.1.8 RS232 interface

Connector Dsub 9 pin (male)

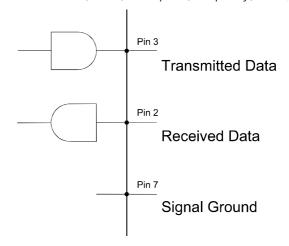


Function TxD and RxD signal for connection with software

handshake

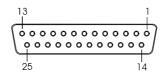
Default settings 9600 baud, 8 bit, 1 stop bit, no parity, XON/XOFF

Pin assignment

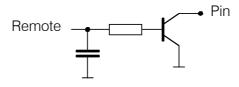


#### 6.1.9 Remote interface

Connector Dsub 25 pin (female)



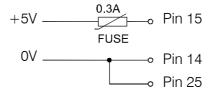
Circuit diagram for output lines 1...8



Assignment of output lines 1...8

Remote 1 Pin 18 Remote 2 Pin 4 3 Remote 3 Pin Remote 4 Pin 1 2 Remote 5 Pin Remote 6 Pin 16 Remote 7 Pin 17 Remote 8 Pin 5

**Potentials** 



## 6.1.10 Analog output

Connector Dsub 9 pin (male)



Pin assignment

Measurement signal (0...1 V for set Full Scale

range 50, 250 or 1000  $\mu$ S/cm)

Accuracy ± 1 %

#### 6.1.11 Mains connection

Voltage 115 V: 100...120 V ± 10 %

230 V: 220...240 V ± 10 %

Frequency 50...60 Hz
Power consumption 100 VA

Fuse 5 mm dia., 20 mm length

100...120 V: 1.0 A (slow-blow) 220...240 V: 0.5 A (slow-blow)

## **6.1.12 Safety specifications**

Construction/testing According to IEC 1010 / EN 61010 / UL 3101-1,

protection class 1, degree of protection IP20

Safety directions The Instructions for Use include information and

warnings which must be heeded by the user to

assure safe operation of the instrument.

## 6.1.13 Electromagnetic compatibility (EMC)

Emitted interference Standards met:

EN55011 (class B), EN55022 (class B),

EN50081-1,

IEC61326 (class B)

Immunity to interference Standards met:

IEC801-2/IEC1000-4-2 (class 3), IEC801-3/ IEC1000-4-3 (class 2), IEC801-4/IEC1000-4-4 (class 4), IEC801-5/IEC1000-4-5 (class 2/3), IEC801-6/IEC1000-4-6 (class 2),

EN50082-1,

EN61000-3-2/IEC1000-3-2, EN61000-3-3/ IEC1000-3-3, EN61000-4-11/IEC1000-4-11,

IEC61326

## **6.1.14** Ambient temperature

Nominal operating range +5...+45°C

(at 20...80 % atmospheric humidity)

*Storage* −20...+70°C *Transport* −40...+70°C

### **6.1.15** Housing

Material of cover Polyurethane rigid foam (PUR) with fire protection

for fire class UL94VO, CFC-free

Material of base Steel, enameled

Width 259 mm
Height 446 mm
Depth 355 mm

Weight 2.761.0010: 13.5 kg (without accessories)

15.7 kg (with accessories)

2.761.0020: 14.7 kg (without accessories)

21.7 kg (with accessories)



# **6.2** Standard equipment



Subject to changes!
All dimensions are given in mm.

The 761 Compact IC is available in two versions:

- 2.761.0010 Compact IC without suppressor module
- 2.761.0020 Compact IC with suppressor module

These instruments include the following parts:

Qua	ant.	Order No.	Description	
2.761.0010	2.761.0020			
1	1	1.732.0110	<b>Detector block (metal-free)</b> with permanently attached connecting cable to 761 Co	ompact IC
-	2	6.1602.150	Bottle attachment GL45 for 6.1608.023 Amber glass bottle (1 L)	GL45
1	1	6.1602.160	For 6.1608.070 Clear glass bottle (2 L), incl. the following accessories:  1 × 6.1446.040 Thread stopper M6 1 × 6.1602.105 Bottle attachment GL 45 1 × 4.420.0311 Tubing nipple M6 1 × 4.420.4300 Tubing nipple M8 2 × E.301.0021 O-ring	A-14/15 GL45
-	2	6.1608.023	Amber glass bottle 1 L Supply bottles for regeneration and rinsing solutions, with GL45 thread	GL45 210
1	1	6.1608.070	Clear glass bottle 2 L Eluent bottle, with GL45 thread	GL45 262

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Qua	ant.	Order No.	Description	
2.761.0010	2.761.0020			
1	1	6.1609.000	CO <sub>2</sub> Absorber tube incl. 6.2701.020 Stopper For 6.1602.160 Bottle attachment.	32 B-14/15
-	1	6.1803.020	PTFE capillary Length = 5 m	0.97
2	2	6.1816.020	Silicone tubing Drain tube for inner compartment and bottle rack, length = 1 m	6 9
-	2	6.1826.060	Pump tubing made of PP (polypropylene) with 2 firmly attached white-yellow stoppers; in.d. = 0.51 mm, ex.d. = 2.31 mm	400
1	1	6.1834.010	Aspirating tubing made of PTFE, with connector for 6.2821.090 aspirating filter Length = 2.5 m For the connection high-pressure pump – eluent bottle	1.5
1	1	6.2023.020	SGJ clip	
1	1	6.2027.030	Column holder Diameter d = 8.5 mm	42
1	1	6.2027.040	Column holder Diameter d = 11.3 mm	42



Qua	ant.	Order No.	Description
2.761.0010	2.761.0020		
1	1	6.2122.0X0	Mains cable           to customer's specifications:           Cable socket         Cable connector           Type IEC 320/C 13         Type SEV 12 (CH)         6.2122.020           Type IEC 320/C 13         Type CEE (7), VII (D)         6.2122.040           Type CEE (22), V         Type NEMA 5-15 (USA)         6.2122.070
1	1	6.2134.100	Connecting cable Connecting cable 761 Compact IC (RS232) – PC
1	1	6.2617.010	Special tool For removing the piston seal of the pump head  M3.5  M3.5  50
1	1	6.2620.150	Pulsation dampener MF Metal-free pulsation dampener to reduce pulsation and prolong the life of separating columns.  52
1	1	6.2621.000	Adjustable spanner  max. 20 150
1	1	6.2621.030	Hexagon key 4 mm  For mounting the pump head of the high-pressure pump.
1	3	6.2744.010	PEEK compression fitting For the connection of 6.1831.010 PEEK capillaries or 6.1822.010 PTFE microcapillaries, set of 5



Qua	ant.	Order No.	Description
2.761.0010	2.761.0020		
1	1	6.2744.020	Coupling 1/16" – Luer Coupling for connection of a 6.1803.000 PTFE capillary to connection 2 of the 761 Compact IC when a 750 Autosampler or a 766 IC Sample Processor is used
-	1	6.2744.030	PEEK coupling Connection between 6.2744.010 PEEK compression fitting and 6.1826.060 Pump tubing; Set of 4
1	1	6.2816.020	Syringe made of PP, volume = 10 mL; for manual filling of the sample loop
1	1	6.2821.090	Aspirating filter Pore dimension 20 μm For 6.1834.000 Aspirating tubing. Set of 5
-	2	6.2821.120	Filter unit PEEK  To avoid contamination due to abrasive particles of piston seals.  Spare part: 6.2821.130 Filter
1	1	6.6030.013	Software CD «761 PC Software 1.1»
1	3	Y.107.0150	Cable strap
1	1	8.110.8213	Instructions for Use (English) for «Autodatabase 1.0» PC program
1	1	8.732.2003	Metrohm Monograph «Ion chromatography» (English)
1	1	8.732.2013	«IC Application Notes» (English)
1	1	8.761.1063	Instructions for Use (English) for 761 Compact IC
1	1	8.761.8007	Registration card (German/English) for PC program «Metrodata 761 PC Software 1.1»



# **6.3** Optional accessories

# **6.3.1** Accessories for capillary connections

Order No.	Description	
6.1825.XXX	Sample loop, made of PEEK For injection valve; incl. two 6.2744.010 PEEK compression fittings 6.1825.210: Volume = 20 μL 6.1825.230: Volume = 10 μL	
6.1831.010	PEEK capillary Length = 3 m	0.25 t <sub>/16</sub> "
6.2620.040	Coupling 1/16" – 1/4" Connector for plastic separating columns with 1/4"-28 thread.	80
6.2621.080	Capillary tubing cutter for plastic capillaries For 6.1831.010 PEEK capillaries and 6.1822.010 PTFE microcapillaries incl. 5 additional blades	55
6.2744.040	PEEK coupling For connection of 1/16" capillaries	9.5
6.2744.070	PEEK compression fitting short Spare part for 6.2824.100 Pump head Set of 5	21
6.2821.130	Filter for filter unit PEEK Spare part for 6.2821.120 Filter unit PEEK. Set of 10	

## 6.3.2 High-pressure pump

Order No.	Description	
6.2824.100	Pump head (metal-free) Complete, with fixations screws	
6.2741.000	Piston seal Spare part for 6.2824.100 Pump head	6.5
6.2741.010	PE piston seal Spare part for 6.2824.100 Pump head (only suitable for aqueous eluents)	6.5
6.2744.070	PEEK compression fitting short Spare part for 6.2824.100 Pump head Set of 5	21
6.2824.070	Zircon piston Spare part for 6.2824.100 Pump head	Ø3 65.5
6.2824.030	Sapphire supporting ring Spare part for 6.2824.100 Pump head	Ø3.2 Ø6.3
6.2824.050	Spring for main piston Spare part for 6.2824.100 Pump head	Ø11
6.2824.060	Spring for auxiliary piston Spare part for 6.2824.100 Pump head	Ø11



Order No.	Description
6.2824.080	Outlet valve (metal-free) Spare part for 6.2824.100 Pump head
6.2824.090	Inlet valve (metal-free) Spare part for 6.2824.100 Pump head

# **6.3.3** Separating columns and precolumns

Order No.	Description	
6.1005.000	IC anion column PRP-X100 (125 mm) For the determination of anions without chemical suppression. Column dimensions: 125 × 4.0 mm Precolumn: 6.1005.020	140
6.1005.010	IC anion column PRP-X100 (250 mm) For the determination of anions without chemical suppression. Column dimensions: 250 × 4.0 mm Precolumn: 6.1005.020	265
6.1005.020	IC precolumn cartridge PRP-X100 To prolong the service life of 6.1005.000 and 6.1005.010 IC anion columns PRP-X100. Column dimensions: 20 × 4.0 mm Installation using 6.2821.040 Cartridge head.	27
6.1005.030	IC exclusion column PRP-X300 For the determination of organic acids without chemical suppression. Column dimensions: 250 × 4.0 mm Precolumn: 6.1005.040	275
6.1005.040	IC precolumn cartridge PRP-X300 To prolong the service life of the 6.1005.030 IC exclusion column PRP-X300. Column dimensions: 20 × 4.0 mm Installation using 6.2821.040 Cartridge head.	27



Order No.	Description	
6.1005.050	IC precolumn cartridge PRP-1 To prolong the service life of the 6.1009.000 IC anion column SUPERSEP and the 6.1006.100 IC anion column METROSEP Anion Dual 2. Column dimensions: 20 × 4.0 mm Installation using 6.2821.040 Cartridge head.	27 ∅6.4
6.1005.100	IC anion column Star-Ion A300 For the determination of anions with chemical suppression. Column dimensions: 100 × 4.6 mm	116
6.1005.200	IC anion column Organic Acids For the determination of organic acids. Column dimensions: 250 × 7.5 mm	275
6.1006.020	IC column cartridge METROSEP Anion Dual 1 For the determination of anions with and without chemical suppression. Column dimensions: 150 × 3.0 mm Installation using 6.2828.000 Glass cartridge holder.	153
6.1006.030	IC precolumn cartridge METROSEP Anion Dual 1 Set of 3 To prolong the service life of the 6.1006.020 IC column cartridge METROSEP Anion Dual 1. Column dimensions: 30 × 3.0 mm Installation using 6.2828.010 Precolumn cartridge holder.	34
6.1006.040	IC column cartridge METROSEP Anion Dual 1 For the determination of anions with and without chemical suppression. Column dimensions: 70 × 3.0 mm Installation using 6.2828.020 Glass cartridge holder.	73
6.1006.100	IC anion column METROSEP Anion Dual 2 For the determination of anions with and without chemical suppression. Column dimensions: 75 × 4.6 mm Precolumn: 6.1005.050 (installation with 6.2821.050 Twin cartridge holder)	107
6.1006.200	Preconcentration cartridge METROSEP Anion For anion preconcentration. Column dimensions: 30 × 3.0 mm Installation using 6.2828.010 Precolumn cartridge holder.	34



Order No.	Description	
6.1007.000	IC cation column Nucleosil 5SA For the determination of divalent cations without chemical suppression. Column dimensions: 125 × 4.0 mm Precolumn: 6.1007.010	140
6.1007.010	IC precolumn cartridge Nucleosil 5SA To prolong the service life of the 6.1007.000 IC cation column Nucleosil 5SA. Column dimensions: 20 × 4.0 mm Installation using 6.2821.040 Cartridge head.	27
6.1008.010	IC cation column Hyperrez Monovalent For the determination of monovalent cations without chemical suppression. Column dimensions: 50 × 4.6 mm	74
6.1009.000	IC anion column SUPERSEP For the determination of anions without chemical suppression. Column dimensions: 100 × 4.6 mm Precolumns: 6.1009.010 IC anion column SUPERSEP or 6.1005.010 IC precolumn cartridge	126
6.1009.010	IC anion precolumn SUPERSEP  To prolong the service life of the 6.1009.000 IC anion column SUPERSEP.	38
6.1010.000	IC cation column METROSEP Cation 1-2 For the determination of monovalent and divalent cations without chemical suppression. Column dimensions: 125 × 4.0 mm Precolumn: 6.1010.010	140
6.1010.010	IC precolumn cartridge METROSEP Cation 1-2 To prolong the service life of the 6.1010.000 IC cation column METROSEP Cation 1-2. Column dimensions: 20 × 4.0 mm Installation using 6.2821.040 Cartridge head.	27
6.1012.X00	Sample pretreatment cartridge IC-RP For non-polar solid phase extraction. Removes organic substances; for the enrichment of heavy metals. With Luer connection. 6.1012.000: set of 50 6.1012.100: set of 10	15.5
6.1012.X10	Sample pretreatment cartridge IC-H Cation exchanger in H <sup>+</sup> form. Removes interfering cations, CO <sub>3</sub> <sup>2-</sup> , HCO <sub>3</sub> <sup>-</sup> or for alkaline samples. With Luer connection. 6.1012.010: set of 50 6.1012.110: set of 10	34



Order No.	Description	
6.1012.X20	Sample pretreatment cartridge IC-Ag Cation exchanger in Ag <sup>+</sup> form. Removes halides. With Luer connection. 6.1012.020: set of 50 6.1012.120: set of 10	15.5
6.1012.X30	Sample pretreatment cartridge IC-OH Cation exchanger in OH <sup>-</sup> form. For highly acidic samples. With Luer connection. 6.1012.030: set of 50 6.1012.130: set of 10	15.5
6.1012.200	Sample pretreatment cartridge Chromafix C18 Removes organic substances (not suitable for fluoride determinations). With Luer connection. 6.1012.200: set of 50	15.5
6.2027.050	Column holder Diameter d = 15.0 mm	42 d
6.2821.010	PTFE gasket Spare part for 6.1005.000, 6.1005.010, 6.1005.030, 6.1007.000 and 6.1010.000 separating columns; set of 4.	5.5
6.2821.020	Steel mesh Spare part for 6.1005.000, 6.1005.010, 6.1005.030, 6.1007.000 and 6.1010.000 separating columns; set of 4.	5.5
6.2821.040	Cartridge head To hold precolumn cartridges; installed in the inlet capillary of the separating column.	42
6.2821.080	Steel spacer Spare part for 6.2821.040 Cartridge head.	Ø0.25 ≈8.5



Order No.	Description	
6.2828.000	Glass cartridge holder To hold the 6.1006.0020 Column cartridge METROSEP Anion Dual 1.	12
6.2828.010	Precolumn cartridge holder To hold the 6.1006.0030 Precolumn cartridge METROSEP Anion Dual 1.	12
6.2828.020	Glass cartridge holder To hold the 6.1006.0040 Column cartridge METROSEP Anion Dual 1.	12 92
6.2832.000	Suppressor rotor Replacement cartridge for Metrohm suppressor module	0
6.2832.010	Connection piece for suppressor rotor with input and output leads	

## 6.3.4 Additional devices and cables

Order No.	Description
2.750.0010	750 Autosampler Sampler for the automation of sample injection. Capacity: 128 sample vessels each with an effective volume of ca. 700 μL; incl. accessories.  Accessories: 6.2413.000 Glass sample vessels, set of 1000 6.2743.000 PP sample vessels, set of 1000 6.2743.010 Polyethylene stoppers, transparent, set of 1000 6.2743.020 Polyethylene stoppers, red, set of 1000 6.2743.030 Filter stoppers, set of 100
2.766.0010	766 IC Sample Processor Sampler for the automation of sample injection. Capacity: 127 sample vessels each with an effective volume of ca. 11 mL; incl. accessories.  Accessories: 6.2743.050 PP sample vessels, set of 2000 6.2743.060 Polyethylene stoppers, transparent, set of 1000
6.2125.010	Cable Connecting cable 761 Compact IC (RS232 interface) – PC Adapter cable 25-pin to 9-pin.
6.2125.020	Cable RS232 extension cable
6.2125.060	Cable Connecting cable 761 Compact IC (RS232 interface) – PC  25 pol. neg.  25 pol. neg.
6.2125.110	Cable Connecting cable 761 Compact IC (RS232 interface) – PC



## 6.4 Warranty and conformity

## 6.4.1 Warranty

The warranty on our products is limited to defects that are traceable to material, construction or manufacturing error which occur within 12 months from the day of delivery. In this case, the defects will be rectified in our workshops free of charge. Transport costs are to be paid by the customer.

For day and night operation, the warranty is limited to 6 months.

Glass breakage in the case of electrodes or other parts is not covered by the warranty. Checks which are not a result of material or manufacturing faults are also charged during the warranty period. For parts of outside manufacture insofar as these constitute an appreciable part of our instrument, the warranty stipulations of the manufacturer in question apply.

With the regard to the guarantee of accuracy, the technical specifications in the instruction manual are authoritative.

Concerning defects in material, construction or design as well as the absence of guaranteed features, the orderer has no rights or claims except those mentioned above.

If damage of the packaging is evident on receipt of a consignment or if the goods show signs of transport damage after unpacking, the carrier must be informed immediately and a written damage report demanded. lack of an official damage report releases Metrohm from any liability to pay compensation.

If any instruments and parts have to be returned, the original packaging should be used if at all possible. This applies above all to instruments, electrodes, burette cylinders and PTFE pistons. Before embedment in wood shavings or similar material, the parts must be packed in a dust-proof package (for instruments, use of a plastic bag is imperative). If open assemblies are enclosed in the scope of delivery that are sensitive to electromagnetic voltages (e.g. data interfaces etc.) these must be returned in the associated original protective packaging (e.g. conductive protective bag). (Exception: assemblies with built-in voltage source belong in a non-conductive protective packaging). For damage which arises as a result of non-compliance with these instructions, no warranty responsibility whatsoever will be accepted by Metrohm.

## 6.4.2 EU Declaration of conformity



# **EU Declaration of Conformity**

The METROHM AG company, Herisau, Switzerland hereby certifies, that the instrument:

# **761 Compact IC**

meets the requirements of EC Directives 89/336/EEC and 73/23/EEC.

### **Source of the specifications:**

EN 50081-1 Electromagnetic compatibility, basic specification;

**Emitted Interference** 

EN 50082-1 Electromagnetic compatibility, basic specification;

Interference Immunity

EN 61010 Safety requirements for electrical laboratory measurement

and control equipment

#### **Description of the instrument:**

Instrument for recording ion chromatograms with electronic or chemical suppression.

Daul & Brown am

Herisau, April 12, 1999

Dr. J. Frank

Ch. Buchmann

Development Manager

Production and

Quality Assurance Manager

## 6.4.3 Certificate of conformity and system validation

# **Certificate of Conformity and System Validation**

This is to certify the conformity to the standard specifications for electrical appliances and accessories, as well as to the standard specifications for security and to system validation issued by the manufacturing company.

Name of commodity: 761 Compact IC

System software: Stored in ROMs

Name of manufacturer: Metrohm Ltd., Herisau, Switzerland Principal technical information: Voltages: 100...120, 220...240 V

Frequency: 50...60 Hz

This Metrohm instrument has been built and has undergone final type testing according to the standards:

IEC801-2/IEC1000-4-2 (class 3), IEC801-3/ IEC1000-4-3 (class 2),

IEC801-4/IEC1000-4-4 (class 4), IEC801-5/IEC1000-4-5 (class 2/3),

IEC801-6/IEC1000-4-6 (class 2), EN50082-1, EN61000-3-2/3/IEC1000-3-2/3,

EN61000-4-11/IEC1000-4-11, IEC61326 (class B), EN55011 (class B),

EN55022 (class B), EN50081-1 — *Electi* 

— Electromagnetic compatibility

IEC1010, EN61010, UL3101-1

— Security specifications

It has also been certified by the Swiss Electrotechnical Association (SEV), which is member of the International Certification Body (CB/IEC).

The technical specifications are documented in the instruction manual.

The system software, stored in Read Only Memories (ROMs) has been validated in connection with standard operating procedures in respect to functionality and performance. The features of the system software are documented in the instruction manual.

Metrohm Ltd. is holder of the SQS-certificate of the quality system ISO 9001 for quality assurance in design/development, production, installation and servicing.

Herisau, April 12, 1999

Dr. J. Frank Ch. Buchmann

Development Manager Production and

Quality Assurance Manager

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